zoosystema

2026 • 48 • 25



Small terrestrial mammal inventory of Assoukoko forest (Togo), with new country records and potential new species for science

Amétépé HOUNMAVO, Gabriel H. SEGNIAGBETO, Kenneth Otieno ONDITI, Holly L. LUTZ, Arnaud DELAPRÉ, Terrence C. DEMOS, Julian C. KERBIS PETERHANS & Violaine NICOLAS

art. 48 (25) — Published on 6 November 2026
www.zoosystema.com

PUBLICATIONS SCIENTIFIQUES

MUSEUM NAT. HIST. NATURELLES

Directeur de la publication / *Publication director*: Gilles Bloch
Président du Muséum national d'Histoire naturelle

Rédactrice en chef / *Editor-in-chief*: Laure Desutter-Grandcolas

Éditrice technique (suivi éditorial) / *Desk editor (editorial process)*: Anne Mabille (zoosyst@mnhn.fr)

Éditrice technique (production) / *Desk editor (production)* : Anne Mabille

Comité scientifique / *Scientific board*:
Nesrine Akkari (Naturhistorisches Museum, Vienne, Autriche)
Maria Marta Cigliano (Museo de La Plata, La Plata, Argentine)
Serge Gofas (Universidad de Málaga, Málaga, Espagne)
Sylvain Hugel (CNRS, Université de Strasbourg, France)
Marco Isaia (Università degli Studi di Torino, Turin, Italie)
Rafael Marquez (CSIC, Madrid, Espagne)
Jose Christopher E. Mendoza (Lee Kong Chian Natural History Museum, Singapour)
Annemarie Ohler (MNHN, Paris, France)
Jean-Yves Rasplus (INRA, Montferrier-sur-Lez, France)
Wanda M. Weiner (Polish Academy of Sciences, Cracovie, Pologne)

Couverture / *Cover*:
Open woodland in Assoukoko Forest, Togo. Photo credit: Amétépé Hounmavo (UL/MNHN, Togo/France).

Zoosystema est indexé dans / Zoosystema *is indexed in:*
– Science Citation Index Expanded (SciSearch®)
– ISI Alerting Services®
– Current Contents® / Agriculture, Biology, and Environmental Sciences®
– Scopus®

Zoosystema est distribué en version électronique par / Zoosystema *is distributed electronically by:*
– BioOne® (http://www.bioone.org)

Les articles ainsi que les nouveautés nomenclaturales publiés dans *Zoosystema* sont référencés par / *Articles and nomenclatural novelties published in* Zoosystema *are referenced by:*
– ZooBank® (http://zoobank.org)

Zoosystema est une revue en flux continu publiée par les Publications scientifiques du Muséum, Paris / **Zoosystema** *is a fast track journal published by the Museum Science Press, Paris*

Les Publications scientifiques du Muséum publient aussi / *The Museum Science Press also publish:*
Adansonia, ***Geodiversitas***, ***Anthropozoologica***, ***European Journal of Taxonomy***, ***Naturae***, ***Cryptogamie*** **sous-sections** ***Algologie***, ***Bryologie***, ***Mycologie***, ***Comptes Rendus Palevol***.

Diffusion – Publications scientifiques Muséum national d'Histoire naturelle
CP 41 – 57 rue Cuvier F-75231 Paris cedex 05 (France)
Tél.: 33 (0)1 40 79 48 05 / Fax: 33 (0)1 40 79 38 40
diff.pub@mnhn.fr / https://sciencepress.mnhn.fr

Les articles publiés dans *Zoosystema* sont distribués sous Licence CC-BY 4.0 / *Articles published in* Zoosystema *are distributed under a CC-BY 4.0 license.*
ISSN (imprimé / *print*): 1280-9551/ ISSN (électronique / *electronic*): 1638-9387

Small terrestrial mammal inventory of Assoukoko forest (Togo), with new country records and potential new species for science

Amétépé HOUNMAVO
Laboratoire d'Écologie et d'Écotoxicologie, Faculté des Sciences,
Université de Lomé, 01BP 1515 Lomé (Togo);
and Institut de Systématique, Évolution, Biodiversité (ISYEB), Muséum national d'Histoire naturelle,
CNRS, Sorbonne Université, EPHE-PSL, Université des Antilles,
case postale 51, 57 rue Cuvier, 75005 Paris (France)
ehounmavo1904@gmail.com (corresponding author)

Gabriel H. SEGNIAGBETO
Laboratoire d'Écologie et d'Écotoxicologie, Faculté des Sciences,
Université de Lomé, 01BP 1515 Lomé (Togo)

Kenneth Otieno ONDITI
National Museums of Kenya, P.O. Box 40658 – 00100 Nairobi (Kenya);
and State Key Laboratory of Genetic Evolution & Animal Models, Kunming Institute of Zoology,
Chinese Academy of Sciences, Kunming 650201 (China)

Holly L. LUTZ
Negaunee Integrative Research Center, Field Museum of Natural History,
1400 S. Lake Shore Drive, Chicago, IL 60605 (United States);
and Denver Museum of Nature & Science, 2001 Colorado Boulevard, Denver, CO 80205 (United States)

Arnaud DELAPRÉ
Institut de Systématique, Evolution, Biodiversité (ISYEB),
Muséum national d'Histoire naturelle, CNRS, Sorbonne Université, EPHE-PSL,
Université des Antilles, case postale 51, 57 rue Cuvier, 75005 Paris (France)

Terrence C. DEMOS
Negaunee Integrative Research Center, Field Museum of Natural History,
1400 S. Lake Shore Drive, Chicago, IL 60605 (United States)

Julian C. KERBIS PETERHANS
College of Arts & Sciences, Roosevelt University,
430 S Michigan Avenue, Chicago, IL 60605 (United States);
and National Museums of Kenya, P.O. Box 40658 – 00100 Nairobi (Kenya)

Violaine NICOLAS
Institut de Systématique, Evolution, Biodiversité (ISYEB), Muséum national d'Histoire naturelle,
CNRS, Sorbonne Université, EPHE-PSL, Université des Antilles,
case postale 51, 57 rue Cuvier, 75005 Paris (France)

Submitted on 13 August 2025 | Accepted on 11 June 2026 | Published on 6 November 2026

urn:lsid:zoobank.org:pub:95797C38-5223-430D-9B5F-8A6BA54EDB25

Hounmavo A., Segniagbeto G., Onditi K., Lutz H., Delapré A., Demos T., Kerbis Peterhans J. & Nicolas V. 2026. — Small terrestrial mammal inventory of Assoukoko forest (Togo), with new country records and potential new species for science. *Zoosystema* 48 (25): 741-781. https://doi.org/10.5252/zoosystema2026v48a25. http://zoosystema.com/48/25

ABSTRACT
Current knowledge of small terrestrial mammal diversity in the semi-deciduous forest of Togo remains incomplete and insufficiently verified taxonomically. Recent records have relied primarily on historical specimens preserved in museum collections, which pose a high risk of taxonomic confusion due to a lack of associated molecular data. Within this context, field surveys were conducted from February 2022 to November 2023 during dry seasons in the Assoukoko Forest, a forest block located at the Ghanaian border near Bismarckburg, the type locality of the endemic Togolese rodent *Leimacomys buettneri* Matschie, 1893 collected in 1890. This survey represents a major contribution to the knowledge of mammalian biodiversity in this poorly explored region. A total of 452 specimens were collected, including 414 Rodentia Bowdich, 1821 (23 species) and 38 Soricidae Fischer, 1814 (five species of *Crocidura* Wagler, 1832). Five taxa are reported for the first time in Togo, including a potentially new species of *Graphiurus* (*sensu lato*). Our study also provides the first molecular confirmation of several previously reported species, including *Mus baoulei* (Vermeiren & Verheyen, 1980), *Crocidura* cf. *jouvenetae*, *Crocidura olivieri* (Lesson, 1827) clade II, *Crocidura foxi* Dollman, 1915, *Mus minutoides* Smith, 1834, *Cricetomys* sp. 1, *Stochomys longicaudatus* (Tullberg, 1893), and *Uranomys ruddi* Dollman, 1909. Habitat-specific analyses revealed higher abundance in gallery and dense semi-deciduous forests, while some generalist species dominated highly disturbed cultivated areas. Local communities contributed additional specimens, including four species not captured by standard trapping. Despite increasing anthropogenic pressure, Assoukoko Forest still hosts a remarkably diverse small mammal community. No *Leimacomys buettneri* individuals were recorded despite intensive targeted trapping, suggesting either local extinction or persistence in less surveyed savanna habitats.

KEY WORDS
Gallery forest,
Leimacomys buettneri,
Gliridae,
Rodentia,
Soricidae,
West Africa.

RÉSUMÉ
Inventaire des petits mammifères terrestres de la forêt d'Assoukoko (Togo), avec de nouvelles observations nationales et une espèce potentiellement nouvelle pour la science.
Les connaissances sur la diversité des petits mammifères terrestres dans la forêt semi-décidue du Togo restent incomplètes et insuffisamment avérées taxonomiquement. Les données récentes reposent principalement sur l'étude des spécimens historiques préservés dans des collections muséales, or ces spécimens présentent un risque élevé de confusion taxonomique du fait de l'absence de données moléculaires associées. Dans ce contexte, des inventaires de terrain ont été menés en saison sèche entre février 2022 et novembre 2023, dans la forêt d'Assoukoko. Ce massif forestier est situé à la frontière du Ghana près de Bismarckburg, la localité type du rongeur endémique *Leimacomys buettneri* Matschie, 1893, collecté en 1890. Cet inventaire apporte une contribution majeure à la connaissance de la biodiversité mammalienne dans cette région peu explorée. Au total, 452 spécimens ont été collectés, comprenant 414 Rodentia Bowdich, 1821 (23 espèces) et 38 Soricidae Fischer, 1814 (cinq espèces de *Crocidura* Wagler, 1832). Cinq taxons sont signalés pour la première fois au Togo, dont une espèce potentiellement nouvelle de *Graphiurus* (*sensu lato*). Notre étude fournit également la première confirmation moléculaire de plusieurs espèces déjà signalées, telles que *Mus baoulei* (Vermeiren & Verheyen, 1980), *Crocidura* cf. *jouvenetae*, *Crocidura olivieri* (Lesson, 1827) clade II, *Crocidura foxi* Dollman, 1915, *Mus minutoides* Smith, 1834, *Cricetomys* sp. 1, *Stochomys longicaudatus* (Tullberg, 1893) et *Uranomys ruddi* Dollman, 1909. Les analyses par habitat montrent une plus grande abondance dans les forêts-galeries et semi-décidues denses, tandis que certaines espèces généralistes dominent dans les zones cultivées perturbées. Les communautés locales ont contribué en collectant des spécimens supplémentaires, dont quatre espèces non capturées par les méthodes standard de piégeage. Malgré une pression anthropique croissante, la forêt d'Assoukoko abrite encore une diversité remarquable. Aucun *Leimacomys buettneri* n'a été capturé malgré un piégeage ciblé intensif, suggérant une possible extinction locale ou une persistance dans des habitats de savane moins explorés.

MOTS CLÉS
Forêt-galerie,
Leimacomys buettneri,
Gliridae,
Rodentia,
Soricidae,
Afrique de l'Ouest.

INTRODUCTION

Small mammals play a key role in the functioning of forest ecosystems, notably by dispersing seeds, serving as prey for various carnivores, raptors, and reptiles, regulating insect populations, and contributing to soil enrichment (Davies *et al.* 2019; Godó *et al.* 2022). Through their functional diversity, they enhance the resilience of forests to both natural and anthropogenic disturbances, thereby maintaining the stability of numerous ecological processes (Čepelka & Dokulilová 2025). This ecological role is particularly critical in ecosystems under high anthropogenic pressure, such as the Assoukoko forest (Togo, West Africa), where human activities, including uncontrolled logging for charcoal production, land conversion for agriculture, and unplanned human settlements, threaten the integrity and long-term sustainability of forest cover. Thus, the conservation of small mammal communities emerges as a major challenge for preserving ecological balance in such habitats.

Togo is divided into five ecological zones (Ern 1979) among which Ecological Zone IV constitutes the only true forested habitats in the country. It includes the Assoukoko forest, one of the last remaining forest blocks in Togo. This zone is also renowned as the type locality of *Leimacomys buettneri* Matschie, 1893, an endemic rodent species never observed again despite several targeted expeditions in the 1960s (USNM, Schmidt *et al.* 2008), 1990s (Hurst *et al.* 1995), and recently in Ghana near the border with Togo by Decher *et al.* (2021). The type locality, formerly known as Bismarckburg, corresponds today to the village of Adélé. In the absence of recent data, the species is currently listed as Data Deficient (DD) on the IUCN Red List. According to Musser & Carleton (2005), *Leimacomys buettneri* is the sole representative of the monotypic subfamily Leimacomyinae Musser & Carleton, 2005. Confirmation of the presence of this endemic Togolese taxon would significantly enhance global understanding of rodent systematics as well as the Togo's faunal value. Togo currently exhibits low mammalian endemism, with only a few endemic species shared with neighboring countries of the Dahomey Gap region, including Walter's duiker *Philantomba walteri* Colyn, Hulselmans, Sonet, Oudé, De Winter, Natta, Nagy & Verheyen, 2010, Dahomey Gap forest mouse *Hylomyscus pamfi* Nicolas, Olayemi, Wendelen & Colyn, 2010, Benin tree hyrax *Dendrohyrax interfluvialis* Oates, Woodman, Gaubert, Sargis, Wiafe, Lecompte, Dowsett-Lemaire, Dowsett, Gonedelé Bi, Ikemeh, Djagoun, Tomsett & Bearder, 2021, *Praomys togoensis* Hounmavo, Demos, Lutz, Delapré, Cornette, Segniagbeto, Kerbis Peterhans & Nicolas, 2026, and a candidate white-bellied pangolin species of the *Phataginus tricuspis* Rafinesque, 1820 group (Gaubert *et al.* 2016). If *Leimacomys buettneri* were to be rediscovered and its habitat accurately characterized, it would greatly strengthen the biogeographic importance and conservation value of these highly fragmented forest habitats. Such a discovery would contribute to a growing body of evidence highlighting the critical role of the Dahomey Gap region as a unique center of biodiversity.

Small terrestrial mammal diversity in Ecological Zone IV of Togo, and in particular within the Assoukoko Forest, remains poorly documented. The main sources currently available are museum specimens from field surveys conducted by Robbins & Van Der Straeten (1996) and the museum-based compilation of Amori *et al.* (2016). The former is based on the collections housed in the USNM (National Museum of Natural History, Smithsonian Institution, Washington D.C.) and MRAC (Musée royal de l'Afrique centrale, Tervuren). The latter primarily relies on specimens housed in the NHM (Natural History Museum, London), IFAN (Institut fondamental de l'Afrique noire, Dakar), MRAC, USNM, MNHN (Muséum national d'Histoire naturelle, Paris), and ZMB (Museum für Naturkunde, Berlin). As part of their study, Robbins & Van der Straeten (1996) conducted field expeditions in Ecological Zone IV of Togo, focusing on the localities of Pagala, Agou, Misahohé, and Ounabé. These surveys revealed high species richness of small mammals, with notable variation across sites.

In Pagala, a total of 112 individuals representing 12 small mammal species were recorded. In Agou, 221 individuals belonging to 10 species were captured, with *Praomys misonnei* Van der Straeten & Dieterlen, 1987 (originally named *Praomys tullbergi* Thomas, 1894) being the most abundant species (75 males and 41 females). In Misahohé, small mammal diversity was particularly high, with at least 15 species recorded and 234 individuals collected, including 166 *P. misonnei*. At Ounabé, only 12 individuals were collected, including a single *P. derooi* Van der Straeten & Verheyen, 1978 and 11 specimens identified as *P. misonnei*. In total, these surveys yielded 579 individuals across the four localities, representing a minimum of 27 species. These data emphasize the diversity of forest-dwelling small mammal communities in the region, with *P. misonnei* being consistently dominant in multiple sites.

In addition, the annotated gazetteer of the USNM African Mammal Project expeditions led by Hank Setzer in the 1960s (Schmidt *et al.* 2008) reported numerous records of mammals from Togo, including rodents, shrews, bats, and hares, with three localities of Ecological Zone IV represented:

Kpalimé: *Heliosciurus* Trouessart, 1880; *Graphiurus* Smuts, 1832; *Lophuromys* Peters, 1874; *Grammomys* Thomas, 1915; *Hylomyscus* Thomas, 1926; *Lemniscomys* Trouessart, 1881; *Mastomys* Thomas, 1915; *Mus* Linnaeus, 1758; *Praomys* Thomas, 1915; *Stochomys* Thomas, 1926; *Crocidura* Wagler, 1832.

Ezimé: *Xerus* Hemprich & Ehrenberg, 1833; *Funisciurus* Trouessart, 1880; *Heliosciurus*, *Graphiurus*, *Lophuromys*, *Uranomys* Dollman, 1909, *Gerbilliscus* Thomas, 1897, *Arvicanthis* Lesson, 1842, *Dasymys* Peters, 1875, *Hylomyscus*, *Lemniscomys*, *Mastomys*, *Mus*, *Praomys*, and *Crocidura*.

Pagala: *Heliosciurus*, *Protoxerus*, *Graphiurus*, *Cricetomys* Waterhouse, 1840, *Uranomys*, *Gerbilliscus*, *Taterillus* Thomas, 1910, *Arvicanthis*, *Dasymys*, *Lemniscomys*, *Mastomys*, *Mus*, *Praomys*, and *Crocidura*. These historical records highlight that the small mammal fauna of the region was already diverse decades before the more recent inventories.

For comparison, Amori *et al.* (2016) reported 34 rodent species and five shrew species in Zone IV based on museum records (see Appendix 1). However, the number of specimens per species is extremely low for many species and typically from very few localities, highlighting the limited sampling effort in this region. Moreover, the Assoukoko forest was not sampled. As a result, previous data are not fully representative and do not allow for an accurate assessment of actual small mammal diversity. Furthermore, taxonomic uncertainties complicate the interpretation of museum data. Several of the species listed by Robbins & Van der Straeten (1996) and by Amori *et al.* (2016) are now known to be part of cryptic species complexes, especially those belonging to the genera *Cricetomys* (Olayemi *et al.* 2012), *Hylomyscus* (Nicolas *et al.* 2010, 2020), *Praomys* (Nicolas *et al.* 2012; Mikula *et al.* 2020), *Lemniscomys* (Nicolas *et al.* 2008; Denys *et al.* 2020), *Mus* subgenus *Nannomys* (Bryja *et al.* 2014), *Graphiurus* (Holden & Levine 2009), and *Mastomys* (Nicolas *et al.* 2012). For these cryptic species, accurate identification of museum specimens can be challenging and often requires confirmation by molecular data, which remains to be collect-

ed for several species (e.g. *P. misonnei*, *H. simus* G. Allen & Coolidge, 1930, *Cricetomys* species). In other cases, species were listed for Togo in the absence of voucher specimens and were based solely on expected range distributions. Such is the case for *Dendromus messorius* (Thomas, 1903), *Crocidura lamottei* Heim de Balsac, 1968, *C. fuscomurina* (von Heuglin, 1865), *C. viaria* (I. Geoffroy Saint-Hilaire, 1834), and *Suncus megalura* (Jentink, 1888), whose presence in Togo is inferred rather than confirmed.

In summary, current data concerning small mammal terrestrial diversity in Ecological Zone IV of Togo are incomplete and lack rigorous taxonomic verification. In this context, our updated inventory from Assoukoko Forest represents a crucial contribution to the knowledge of small mammal biodiversity in Togo's Ecological Zone IV, providing new data that will help clarify the actual species composition of this underexplored gallery forest.

The objectives of this study are as follows:

1) to inventory the species of small terrestrial mammals occurring in the Assoukoko forest, their relative abundance and habitat preferences.

2) to assess the current presence of *Leimacomys buettneri* through an intensive trapping effort within dense and gallery forest since it is assumed to live in the forest (Amori *et al.* 2016). The type locality of *Leimacomys buettneri*, formerly known as Bismarckburg, is located in a partially forested area (Krell 1994), which may suggest an association with forest-type environments. However, because the exact capture site of the holotype is unknown, the possibility that the specimen originated from another locality cannot be entirely excluded. Today, this locality has been transformed into a populated and touristic zone, with the Assoukoko Forest remaining the only significant forested area in the region.

MATERIAL AND METHODS

Study area

This study was conducted in the Assoukoko Forest, a protected area of approximately 100 km² located in the west-central region of Togo, along the border with Ghana (Dowsett-Lemaire & Dowsett 2016). The forest stretches from the village of Yalla in the south (7°51'45"N, 0°37'51"E) to Djikpéléou in the north (8°12'39"N, 0°36'30"E), the last village before the Ghanaian border. The forest is named after the Assoukoko River, which originates within its boundaries and flows westward into Ghana and into Lake Volta.

The vegetation of the Assoukoko forest consists of a mosaic of gallery forests bordering the Assoukoko River and small streams, dense semi-deciduous forests in varying degrees of degradation, and cultivated fields established after deforestation.

The region experiences a tropical climate with a single rainy season from June to September, and a long dry season from November to February, marked by the harmattan, a dry and dusty wind from the Sahara. Annual rainfall ranges from 1300 to 1800 mm, and temperatures vary between 17°C and 39°C (Segniagbeto *et al.* 2017). In 2023, exceptionally heavy rains in August and September caused unprecedented flooding, submerging the only bridge linking Diguingue to Assoukoko and temporarily disrupting travel between the two localities.

Three main sampling sites were selected (Fig. 1): Assoukoko (south), Diguingue (center), and Yegue (north). The selection of these three sites was guided by the intention to sample habitats closest to Bismarckburg, where the dense forest remains relatively well-preserved. We sampled the eastern slopes of the Assoukoko forest because they are considerably more accessible and consist of moderate hills, unlike the western slopes where the terrain is markedly steeper and much more difficult to explore. Furthermore, this choice is also supported by a preliminary survey conducted in November 2022 among local communities living near the forest, during which participants were shown photographs of different small mammal species. This photo-based identification survey revealed a notable richness of rodents and shrews in the eastern part of the forest, particularly between Assoukoko and Diguingue. The immediate surroundings of the sampling sites included cultivated plots, notably with plantain bananas and cocoa trees. At Diguingue and Assoukoko, several small hamlets are located within these cultivated fields. The cultivated area is larger in Yegue and Assoukoko than in Diguingue. Other habitats include dense semi-deciduous forests and, gallery forests.

Collection methodology

Small mammal trapping

Small mammal trapping was carried out using medium-sized Sherman live traps (9 × 7.5 × 23 cm), snap traps (Victor and Museum Special), and pitfall traps with drift fences.

Sherman and snap traps

Despite variations in trapping effort due to logistical constraints, the methods employed remained consistent. The number of traps varied (48-300) between sessions, but the setup always included a combination of snap traps and Sherman live traps. Traps were arranged in pairs spaced 8 to 10 m apart, with each line covering approximately 115 to 120 m depending on the number of traps. Trapping sessions lasted between four and 14 nights, with traps baited daily using a mixture of peanut butter and grated cassava or oats, supplemented with smoked fish. For more details, see the Table 1.

Pitfall trapping

Pitfall trapping was implemented at the Diguingue and Assoukoko sites. Due to technical constraints, no pitfalls were set at Yegue. In Diguingue, a 50-metre-long pitfall line was installed following the protocol described by Nicolas *et al.* (2003). The line consisted of ten 10-liter buckets spaced at 5 m, buried flush with the ground and connected by a plastic drift fence approximately 0.5 metres high. At Assoukoko, five pitfall trap lines were installed in forested areas near the Assoukoko River, following the same methodology as at Diguingue (Nicolas *et al.* 2003), except for the number of buckets per line. Each line at Assoukoko consisted of seven to 11 plastic buckets (10 L), spaced 5 m apart, buried flush with

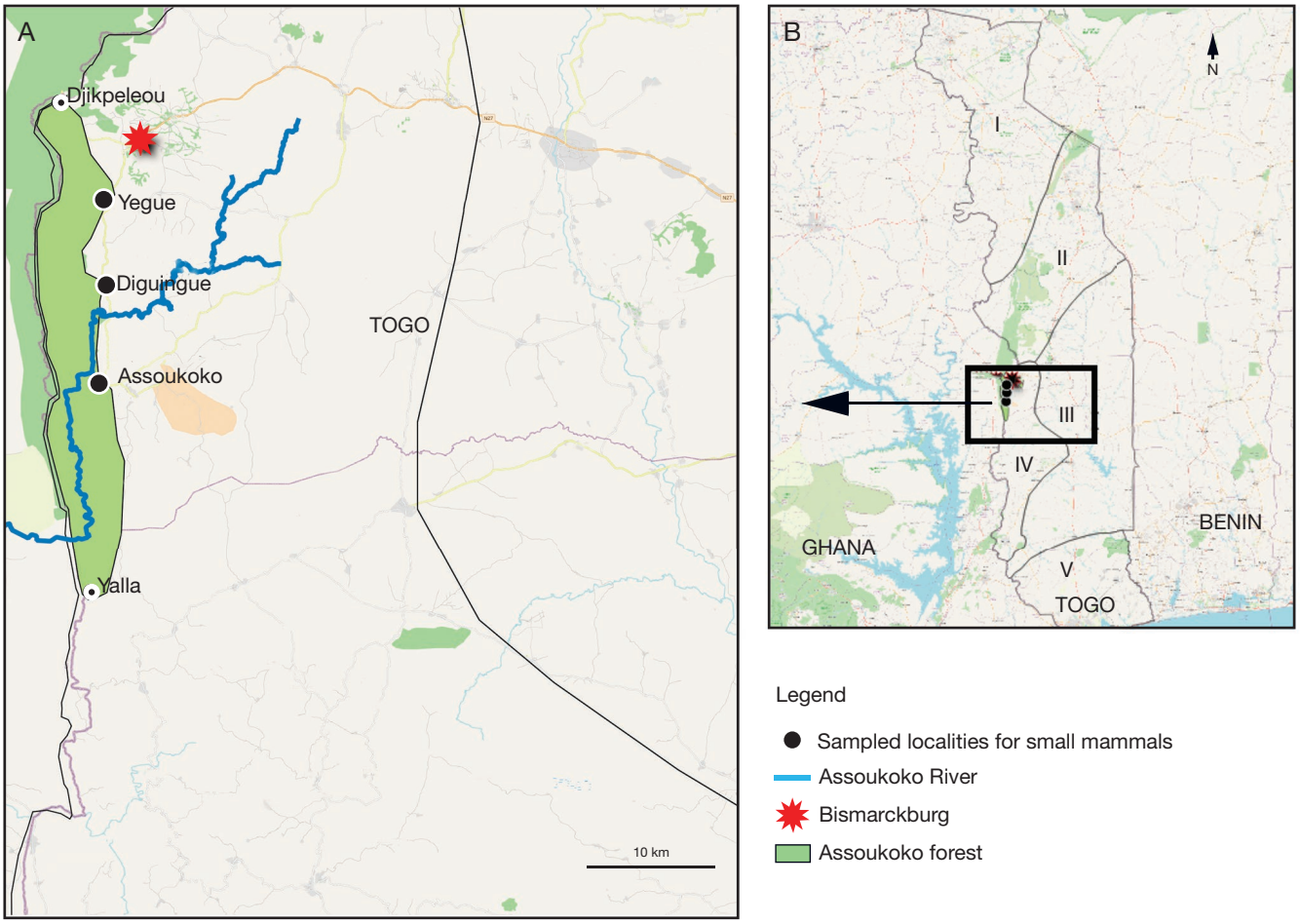


FIG. 1. — **A**, map of sampled localities in Zone IV; **B**, five ecological zones of Togo: **I**, dry savannas of the northern plains; **II**, mosaic of open woodlands, dry dense forests, and montane savannas of the north; **III**, dry dense forests and Guinean savannas of the central plains; **IV**, semi-deciduous dense forests of the southwestern mountains; **V**, dry coastal zone (mosaic of forests, savannas, thickets, and grasslands).

the ground, and connected by a plastic drift fence approximately 0.5 m high. At Diguingue, the trapping took place from 24 to 28 April 2023, 20 to 26 May 2023, and 18 to 24 June 2023, totaling 16 nights. At Assoukoko, pitfall trapping was conducted from 28 February to 7 March 2023. Two lines with 11 buckets and one line with seven buckets were operated for six nights, while one line with seven buckets and one with 10 buckets were operated for five nights.

COMMUNITY-BASED COLLECTION OF SMALL MAMMALS

In addition to direct trapping, a participatory approach was implemented with local communities near the Assoukoko forest to capture small mammals in fields or around dwellings. We provided them twenty snap traps and two Tomahawk live traps. In addition, villagers regularly practice communal drive hunting, which involves groups of three to four individuals using hoes and clubs to capture small mammals for local consumption. Undamaged specimens obtained through this method were brought to us and incorporated into the study. A total of 12 099 trap nights (11 680 Sherman or snap trap nights and 419 pitfall nights) were conducted throughout all trapping campaigns.

SPECIMEN PROCESSING

All specimens captured alive were euthanized by cervical dislocation. Standard external measurements were recorded, including total body length (TOT), tail length (TAIL), ear length (EAR), and hind foot length with claw (HF). Muscle samples were collected and preserved in 95% ethanol or Zymo DNA/RNA Shield (Zymo Research, USA) for subsequent genetic analyses. Specimens were first fixed in 10% formalin, then transferred to 70% ethanol for long-term preservation. All samples were deposited in the Laboratory of Ecology and Ecotoxicology (LaEE) at the University of Lomé (Lomé, Togo) and at the Field Museum of Natural History (Chicago, IL, USA).

SPECIES IDENTIFICATION

The taxonomic nomenclature adopted in this study largely follows the Mammalian Diversity Database (https://www.mammaldiversity.org). Individual identification was first performed on external morphology and then confirmed by molecular analysis of the mitochondrial *Cytochrome b* (*Cytb*) gene for each morphogroup. For identification of species we applied a DNA-barcoding approach and Sanger sequencing of the *Cytb* gene of selected specimens using the DNA extraction,

TABLE 1. — Summary of small terrestrial mammal trapping efforts in Assoukoko, Diguingue, and Yegue (Assoukoko forest in Ecological Zone IV, Togo): Trap types, spatial arrangement, habitat contexts, and sampling durations (2022-2023).

Locality	Session Dates	Total Number of Traps	Trap Configuration / Habitats	Duration (nights)	Notes
Assoukoko	20-26.XI.2022; 28.I-7.III. 2023	48 traps, then 7 lines of 20 to 60 traps each	1st session: 2 lines (riverine mixed cultivated field with plantain and cocoa, avocado trees, and gallery forest; cultivated field only) 2nd session: 7 lines in forest and agricultural habitats with mixed Victor, Museum Special (MS), and Sherman traps	4-14 nights	–
Diguingue	22-26.XI.2022	48 + 48 traps	Gallery forest and cultivated field	4 nights	–
Yegue	8-22.II.2022	300 traps (6 lines of 50 traps)	Gallery forest, semi-deciduous dense forest, and cultivated field	14 nights	–
Diguingue (*Leimacomys buettneri* monitoring)	IV-XI 2023 (monthly)	144 traps (3 × 48 traps)	Gallery forest, semi-deciduous dense forest, and cultivated fields within forest matrix	4-6 nights/month	Targeted monthly monitoring following ethno-mammalogical survey

polymerase chain reaction (PCR), and sequencing protocols as described in Nicolas *et al.* (2012). The DNA sequences obtained were compared to those available in the Genbank database using BLAST. Recent molecular studies are available for the genera *Hylomyscus* (Nicolas *et al.* 2012, 2020), *Mus Nannomys* (Bryja *et al.* 2014), *Praomys* (Bryja *et al.* 2010; Nicolas *et al.* 2011, 2012; Mikula *et al.* 2020); *Lemniscomys* (Nicolas *et al.* 2008; Denys *et al.* 2020; Hánová *et al.* 2021), *Mastomys* (Nicolas *et al.* 2012), *Lophuromys* (Verheyen *et al.* 2007; Igbokwe *et al.* 2016; Onditi *et al.* 2021), *Gerbilliscus* (Granjon *et al.* 2012), *Cricetomys* (Olayemi *et al.* 2012), and *Crocidura* (Jacquet *et al.* 2014, 2015; Igbokwe *et al.* 2019; Nicolas *et al.* 2019) facilitating accurate molecular species determination.

MOLECULAR ANALYSES

DNA was extracted from tissue samples of specimens previously preserved in ethanol using the QIAGEN DNeasy Blood and Tissue Kit, following the manufacturer's instructions. Amplification of the mitochondrial *cytochrome b* (*Cytb*) gene was performed by PCR, primarily using primers L14723 and H15915 (Ducroz *et al.* 2001), and, when necessary, the primer pair L14723 and H14896 (Ducroz *et al.* 2001). The amplification program consisted of 40 cycles with the following parametres: denaturation at 94°C for 30 seconds, annealing at 52°C (or 55°C for the L14723/H14896 pair) for 40 seconds, followed by extension at 72°C for 90 seconds.

For specimens belonging to the genus *Gerbilliscus*, when the initial primers failed, primers L14723 and H6 (Colangelo *et al.* 2015) were used with the same protocol, except that the annealing temperature was set at 55°C. The double-stranded PCR products were subsequently sent to Eurofins laboratory for purification and sequencing. The obtained chromatograms (AB1 files) were cleaned, assembled, and aligned using CodonCode Aligner software (version 9, CodonCode Corporation, 2020). The sequences generated in this study have been deposited in GenBank under accession numbers PX147189-PX147380.

STATISTICAL ANALYSES

Rarefaction curves based on captured individuals were generated using the rarecurve function from the vegan package in R (Oksanen *et al.* 2025) to assess and compare species richness across localities while taking into account the number of individuals captured.

To this end, the raw dataset was first filtered to exclude records originating from community-based collections. These data were not included in the construction of rarefaction curves, as they were collected sporadically and in a non-standardized manner, often during hunting drives involving a variable number of participants. As such, they do not constitute a controlled or reproducible trapping effort and are therefore unsuitable for standardized analyses such as rarefaction.

Following data cleaning, captures were aggregated by locality, combining all trap types (snap traps, Sherman traps, and pitfall lines), in order to construct a species-by-locality abundance matrix. This matrix served as the input for the rarefaction analysis. The shape and inflection of the resulting curves were used to determine whether species richness tended toward a plateau or continued to increase with the number of individuals captured.

ABBREVIATIONS

NHM	Natural History Museum, London (formerly BMNH);
IFAN	Institut fondamental de l'Afrique noire, Dakar;
MNHN	Muséum national d'Histoire naturelle, Paris;
MRAC	Musée royal de l'Afrique centrale, Tervuren;
USNM	National Museum of Natural History, Smithsonian Institution, Washington D.C.;
ZMB	Museum für Naturkunde, Berlin.

TABLE 2. — Summary of capture effort and success by locality and habitat using trapping, pitfall methods, and local contributions.

		Sherman and snap traps			Pitfall traps			Local contribution
Locality	Habitats	No. of trap-nights	No. of captures	Trap success (%)	No. of trap-nights	No. of captures	Trap success (%)	No. of captures
Assoukoko	Cultivated fields	484	22	4.55	–	–	–	8
Assoukoko	Gallery Forest	1572	38	2.42	259	8	3.09	1
Diguingue	Cultivated fields	1968	39	1.98	–	–	–	3
Diguingue	Gallery Forest	3264	64	1.96	160	5	3.12	3
Diguingue	Semi-deciduous dense forest	192	8	4.17	–	–	–	–
Yegue	Cultivated fields	1400	136	9.71	–	–	–	2
Yegue	Gallery Forest	1400	18	1.29	–	–	–	3
Yegue	Semi-deciduous dense forest	1400	94	6.71	–	–	–	–
Total		11680	419	3.59	419	13	3.10	20

SYSTEMATIC

REMARK

Specimens were captured using a combination of snap traps and live traps, pitfall traps, and local resident contributions (Table 2). We captured 452 specimens, of which 414 were rodents (14 genera, 23 species) and 38 were shrews (all in the genus *Crocidura*, five species; Table 3).

Order RODENTIA Bowdich, 1821
Family GLIRIDAE Muirhead, 1819

Genus *Graphiurus* Smuts, 1832

Our dormouse captures include three specimens representing three distinct species-level entities, treated in this manuscript as *Graphiurus* sp. A, *G. kelleni* (Reuvens, 1890), and *Graphiurus* sp. B.

No recent molecular revision is available for *Graphiurus*, although several studies (Musser & Carleton 2005) have highlighted the need for taxonomic reassessment. We captured three specimens of *Graphiurus*, and for two of them we obtained *Cytb* sequences. One of our sequences (specimen JCK11238) did not match any known sequence in GenBank or unpublished datasets and is herein referred to as *Graphiurus* sp. A second sequence (specimen JCK11281) clusters with *G. kelleni* (Genbank sequence MN616984, similarity 90.64%). Unfortunately, no tissue in ethanol is available for the last specimen captured by locals. Based on clear morphological differences, including tail shape and ventral fur coloration, we consider this individual (AH201) to represent a third, distinct species of *Graphiurus*, different from both *G. kelleni* and *Graphiurus* sp. Accordingly, it is treated as a separate taxonomic unit in our estimates of species richness. We referred to this specimen as *Graphiurus* sp. B, as its taxonomic assignment to a described species could not be confirmed with certainty due to the absence of genetic data and the documented high levels of cryptic diversity within the genus.

Family MURIDAE Illiger, 1811

Genus *Gerbilliscus* Thomas, 1897

Our gerbil captures include five specimens assigned to a single species-level entity, treated in this manuscript under the name *Gerbilliscus* sp.

Within the genus *Gerbilliscus*, our four sequences (out of five specimens captured) clusters with 95-99% similarity with the entity (GenBank JF704147) referred to as *Gerbilliscus* sp. by Granjon *et al.* (2012), a clearly delimited lineage not yet formally described. This taxon is characterized by a relatively large body size, a long tail, a distinct karyotype, and forms a distinct genetic clade within the genus. Its distribution covers the Sudano-Guinean forest-savanna mosaic, extending from Benin to Guinea. In this region, it is often found in sympatry with *Gerbilliscus kempi* Wroughton, 1906 and *G. guineae* Thomas, 1910. Granjon *et al.* (2012) discussed the possibility that this species may correspond to *G. kempi*. If confirmed, this would imply that one name, currently included within *G. kempi*, deserves full species status, probably *G. giffardi* (Wroughton, 1906). However, due to the lack of genetic data from type material, and pending a thorough taxonomic revision, we retain the provisional nomenclature used by Granjon *et al.* (2012) for this distinct West African lineage and named it *Gerbilliscus* sp.

Genus *Hylomyscus* Thomas, 1926

Within the genus *Hylomyscus*, our three sequences cluster with 96-99% similarity to the sequence from Benin (GenBank MN845700) identified as *Hylomyscus pamfi* Nicolas, Olayemi, Wendelen, & Colyn, 2010 (Nicolas *et al.* 2020). This taxon belongs to the *alleni* species group, which also includes *H. stella* Thomas 1911, *H. walterverheyeni* Nicolas, Wendelen, Barriere, Dudu & Colyn, 2008, *H. simus* (G. M. Allen & Coolidge, 1930) (two distinct lineages), and *H. alleni* (Waterhouse, 1838). In the phylogeny inferred from complete mitochondrial genomes and five nuclear genes (Nicolas

TABLE 3. — Number of specimens for each rodent and shrew species collected in the three sites Assoukoko, Diguingue and Yegue per habitat type. Abbreviations: **GF**, gallery forest; **CF**, cultivated field; **SF**, semi-deciduous dense forest.

	Assoukoko		Diguingue			Yegue			
	GF	CF	GF	CF	SF	GF	CF	SF	Total
Rodentia Bowdich, 1821									
Cricetomys gambianus Waterhouse, 1840	0	0	0	0	0	0	1	1	2
Cricetomys sp. 1	0	0	1	0	0	0	0	0	1
Cricetomys sp.	1	0	0	0	0	0	3	0	4
Euxerus erythropus (Geoffroy, 1803)	0	1	0	0	0	0	0	0	1
Funisciurus leucostigma Temminck, 1853)	0	0	0	0	0	1	0	0	1
Gerbilliscus sp.	0	0	2	0	0	0	0	3	5
Graphiurus kelleni (Reuvens, 1890)	0	0	0	1	0	0	0	0	1
Graphiurus sp. A	0	0	1	0	0	0	0	0	1
Graphiurus sp.	0	0	0	1	0	0	0	0	1
Hylomyscus pamfi Nicolas, Olayemi, Wendelen & Colyn, 2010	1	0	0	0	0	0	0	2	3
Lemniscomys macculus Thomas & Wroughton, 1910	0	0	0	0	0	0	2	0	2
Lemniscomys striatus (Linnaeus, 1758)	0	4	9	9	1	2	25	0	50
Lophuromys sikapusi (Temminck, 1853)	0	0	2	0	0	5	0	0	7
Mastomys erythroleucus (Temminck, 1853)	0	1	4	5	3	0	89	8	110
Mastomys natalensis (Smith, 1834)	0	0	0	2	0	0	1	0	3
Mus (*Nannomys*) sp.	0	0	2	0	0	0	3	21	26
Mus (*Nannomys*) *baoulei* (Vermeiren & Verheyen, 1980)	1	0	0	1	2	0	0	3	7
Mus (*Nannomys*) *minutoides* Smith, 1834	2	4	1	0	2	0	3	4	16
Praomys daltoni (Thomas, 1892)	2	4	3	16	0	1	0	0	26
Praomys misonnei Van Der Straeten & Dieterlen, 1987	30	0	33	5	0	0	1	48	117
Rattus rattus (Linnaeus, 1758)	0	17	1	1	0	0	4	0	23
Stochomys longicaudatus (Tullberg, 1893)	3	0	1	0	0	0	0	2	6
Uranomys ruddi Dollman, 1909	0	0	0	0	0	0	1	0	1
Soricidae Fischer, 1814									
Crocidura foxi Dollman, 1915	2	0	1	0	0	5	1	0	9
Crocidura cf. *jouvenetae*	1	0	5	1	0	1	2	1	11
Crocidura obscurior Heim de Balsac, 1958	2	0	1	0	0	2	0	0	5
Crocidura olivieri clade II	0	0	3	1	0	2	1	1	8
Crocidura sp.	1	0	1	0	0	2	1	0	5
Total	46	31	71	43	8	21	138	94	452

et al. 2020), *H. pamfi* is closely related to the two lineages of *H. simus*, forming a well-supported clade distinct from other members of the group.

Genus *Lemniscomys* Trouessart, 1881

Our *Lemniscomys* captures include 52 specimens assigned to two species, *Lemniscomys striatus* Linnaeus, 1758 and *L. macculus* Thomas & Wroughton, 1910.

Regarding the genus *Lemniscomys*, the studies by Nicolas *et al.* (2008) and Hánová *et al.* (2021) confirm the existence of distinct genetic lineages within the genus. Twenty-two of our sequences showed 99% similarity with sequence EU422966 deposited in GenBank, corresponding to a specimen collected in Igbéré (Benin) and assigned to clade III of *Lemniscomys striatus* as defined by Nicolas *et al.* (2008). According to these authors *L. striatus* is a monophyletic species subdivided into four major regional clades: clade I, restricted to Central Africa; clade II, to East Africa; clade III, to Nigeria and Benin; and clade IV, to West Africa. These results were later confirmed by Hánová *et al.* (2021), who additionally identified two more mitochondrial lineages based on a expanded geographical sampling (which results in six clades recognized within the monophyletic *L. striatus*): clade A in Guinea, Côte d'Ivoire and southern Mali; clade B in Côte d'Ivoire and Ghana; clade C in Benin and Nigeria; clade D from Cameroon and Gabon to Democratic Republic of Congo; clade E in Kenya; clade F in southwest Kenya, Rwanda and Tanzania. Our sequences fall within lineage C. Accordingly, we refer these specimens to *L. striatus*. Twenty-nine additional specimens are assigned to this taxon based on their morphological similarity to the sequenced individuals.

For two other sequenced specimens (JCK10976 and JCK11059), BLAST analyses revealed 91-99% sequence similarity with GenBank reference sequences assigned to *Lemniscomys bellieri* Van der Straeten, 1975, *L. linulus* Thomas, 1910 and *L. macculus* (Thomas & Wroughton, 1910). Based on multilocus phylogenetic species delimitation analyses and morphological comparisons, Hánová *et al.* (2021) assessed the validity of several historically described taxa within the *macculus* species group and proposed recognizing three species within it: *L. linulus* that can be differentiated based on their stripe patterns and restricted distribution in the western Sudanian savanna, *L. mittendorfi* Eisentraut, 1968 which is endemic to Mt Oku, and *L. macculus* largely distributed from Western (Eastern Guinea) to Eastern Africa. *Lemniscomys bellieri* is considered by these authors as a junior synonym of *L. macculus*. Thus, we exclude the names *L. linulus* and *L. mittendorfi* on the basis of stripe

pattern and/or geographic distribution and *L. bellieri* on the basis of synonymy. In line with the taxonomic conclusions of Hánová *et al.* (2021), we assign the name *L. macculus* to the two sequenced specimens.

Genus *Lophuromys* Peters, 1874

Our *Lophuromys* captures include seven specimens assigned to a single species, *Lophuromys sikapusi* Temminck, 1853. Recent molecular studies have provided new insights into species diversity within the genus *Lophuromys*, a group characterized by its insectivory and dense, spiny pelage. Based on mitochondrial DNA phylogeny, Onditi *et al.* (2021) proposed a subdivision of the genus into three distinct species groups and assessed the number of species within each group. Among the seven *Lophuromys* specimens we captured, five were sequenced and show a 98–99% similarity with the GenBank sequence KT232249, which corresponds to *L. sikapusi*.

Genus *Mastomys* Thomas, 1915

Among the 107 *Mastomys* individuals captured, 17 were identified as *Mastomys erythroleucus* Temminck, 1853 and three as *M. natalensis* Smith, 1834 based on mitochondrial *Cytb* sequences. The *M. erythroleucus* sequences showed 96-100% similarity with GenBank sequences (e.g., GQ227863), while those of *M. natalensis* showed 99-100% similarity with sequence HE864013. According to the recent phylogeographic study by Hánová *et al.* (2021), *M. erythroleucus* and *M. natalensis* are widespread across sub-Saharan Africa and exhibit a strong geographic structure, with several major clades corresponding to distinct regions. Based on BLAST results and the geographic origin of our samples, the *M. erythroleucus* individuals we sequenced likely fall within the West African clade defined by Hánová *et al.* (2021), which includes specimens from Benin, Ghana, and neighboring countries. The *M. natalensis* individuals cluster within the Central-West African clade from the same study.

Genus *Mus* Linnaeus, 1758

Our *Mus* captures include 49 specimens assigned to three species-level entities, *Mus* (*Nannomys*) *minutoides* Smith, 1834, *Mus* (*Nannomys*) *baoulei* Vermeiren & Verheyen, 1980, and *Mus* (*Nannomys*) sp.

Molecular data have revealed a high level of cryptic diversity within *Mus* in the subgenus *Nannomys* (Bryja *et al.* 2014). Among the 49 individuals captured, 23 yielded usable sequences. Sixteen sequences clustered unambiguously with the molecular operational taxonomic unit MOTU27, clade W (97-99 % similar with GenBank KJ935869) identified by these authors, which corresponds to *Mus* (*Nannomys*) *minutoides*. The seven remaining sequences correspond to MOTU12 (96-99% similar with Genbank EU603991, i.e., *Mus* (*Nannomys*) *baoulei*. Unfortunately, no tissues are available for the remaining 26 specimens. Given the difficulty in identifying this species based only on morphological data we refer them as to *Mus* (*Nannomys*) sp.

Genus *Praomys* Thomas, 1915

Our *Praomys* captures include 143 specimens assigned to two species, *Praomys daltoni* Thomas, 1892 and *Praomys misonnei*.

Recent molecular studies have clarified taxonomy within the genus *Praomys*. Two taxa that were long considered distinct based on ecological and morphological criteria are now recognized as synonyms. *Praomys derooi*, described by Van der Straeten & Verheyen (1978), has been re-evaluated and is currently considered a synonym of *Praomys daltoni* (Bryja *et al.* 2010; Mikula *et al.* 2020; Nicolas *et al.* 2021). According to Mikula *et al.* (2020), *Praomys daltoni* is a monophyletic species comprising four distinct mitochondrial lineages, A, B, C1 + C2 + D, as defined by Bryja *et al.* (2010), in addition to lineage M, newly defined from Mauritania by Mikula *et al.* (2020). Lineage A occurs between the Volta and Niger rivers, while lineage B is found on the left bank of the Niger River, with all records in the savannas of northern Nigeria, Cameroon, and southern Niger. The C+D clade occupies a wide area of the western Sudanian savannas, ranging from Senegal to eastern Ghana and Benin, where it overlaps with lineage A. Lineage D meets C1 along the Niger River in Mali, and lineage C2 (mostly corresponding to populations historically assigned to *P. derooi*) extends from Côte d'Ivoire, where it overlaps with C1, to Benin, where it meets lineage A. Finally, lineage M is restricted to the Saharan mountains of southern Mauritania. Commensal individuals are found in all lineages except lineage M. Twelve of our sequences show a similarity of 99-100% to lineage A, while three others match clade C, which largely corresponds to populations formerly identified as *P. derooi*. We refer to all these lineages collectively as *Praomys daltoni*. An additional eleven specimens (unsequenced) were assigned to this species based on their morphological similarity to sequenced individuals.

Regarding *Praomys misonnei*, Nicolas *et al.* (2012) identified 120 haplotypes grouped into four major clades (I-IV). Clade I is restricted to West Central Africa, Clade II occurs in Benin and Ghana, Clade III is limited to Nigeria, and Clade IV is found in Central and East Africa. Among our 117 individuals of *P. misonnei*, 34 were sequenced and showed 98-100% similarity with Clade II, which includes individuals from Benin and Ghana.

Genus *Rattus* Fischer, 1803

Rattus rattus Linnaeus, 1758 is easy to identify based on morphological grounds. Additionally, 15 specimens with molecular samples out of 23 *Rattus rattus* captured, all have 100% similarity to GenBank reference sequences.

Genus *Stochomys* Thomas, 1926

Stochomys longicaudatus Tullberg, 1893 is also easy to identify based on morphological grounds. However, our species identification was confirmed for five specimens of *Stochomys longicaudatus* out of six specimens captured with 96 to 100% similarity to GenBank reference sequence EU349786. This species has also recently been described for the first time from the Volta Region in eastern Ghana (Decher *et al.* 2021). A recent monograph summarises knowledge of the natural history of this rodent in West and Central Africa (Uebbing 2019).

Genus *Uranomys* Dollman, 1909

Our only specimen of *Uranomys ruddi* Dollman, 1909, a neonate, had 96.12% similarity to GenBank ON419658. *Uranomys ruddi* was first reported from Palimé, Togo (Petter 1963) and subsequently from several other localities in the country (Robbins & Van der Straeten 1996; Schmidt *et al.* 2008; Amori *et al.* 2016). It is rather patchily distributed in West Africa, hence fairly rare in collections.

Family Nesomyidae Major, 1897

Genus *Cricetomys* Waterhouse, 1840

Our *Cricetomys* captures include seven specimens assigned to *Cricetomys gambianus* Waterhouse, 1840, *Cricetomys* sp. 1 and *Cricetomys* sp. Of these seven specimens, three were successfully sequenced. Two sequences correspond to *Cricetomys gambianus* (94-99% similarity with GenBank accession JQ410195), and one to *Cricetomys* sp. 1 (99.61% similarity with GenBank accession JQ410208), as proposed in the revision by Olayemi *et al.* (2012). We gather here under *Cricetomis* sp. three putative species, yet undescribed, which will be studied later. A dedicated taxonomic revision, including molecular data from all type specimens, is urgently needed to clarify the taxonomy of this genus. Given the difficulty of identifying these species based only on morphological data, we refer the four remaining specimens to *Cricetomys* sp., as no ethanol-preserved tissue samples were available for molecular analyses.

Family Sciuridae Fischer, 1817

Genus *Funisciurus* Trouessart, 1880

Only one specimen of *Funisciurus leucostigma* (Temminck, 1853) JCK11056 (FMNH240867) was collected. It has already been published with the revision of the genus *Funisciurus* (Curaudeau *et al.* 2025). That study demonstrated that *F. leucostigma* is the only rope squirrel species occurring in West Africa, including Togo. This study is based on pelage coloration, museum material, citizen-science records, and ecological niche modelling. Accordingly, records previously attributed to *F. pyrropus* (Cuvier, 1833) or *F. anerythrus* (Thomas, 1890) in Togo as included in the checklist of Amori *et al.* (2016), are now considered to refer to *F. leucostigma*.

Genus *Euxerus* Thomas, 1909

Regarding *Euxerus erythropus* É. Geoffroy Saint-Hilaire, 1803 we follow Kryštufek *et al.* (2016) in the use of *Euxerus* instead of *Xerus*. Their revision of African ground squirrels recognized *Euxerus* as a distinct genus within Xerini Osborn, 1910. *Euxerus erythropus* is a widespread West and Central African species typically associated with savanna and open woodland habitats, although it can also occur in cultivated areas (Herron & Waterman 2004). Our only specimen was genetically close to GenBank sequence OR167493 (99.16% similarity).

Order EULIPOTYPHLA Waddell, Okada & Hasegawa, 1999
Family Soricidae Fischer, 1814

Genus *Crocidura* Wagler, 1832

Our *Crocidura* captures include 38 specimens assigned to five species-level entities, *C.* cf. *jouvenetae* (Heim de Balsac, 1958), *C. obscurior* Heim de Balsac, 1958, *C. olivieri* (Lesson, 1827) clade II, *C. foxi* Dollman, 1915, and *Crocidura* sp.

There has been no comprehensive taxonomic revision of the complex formed by *C. crossei* (Thomas, 1895) and *C. jouvenetae*. *Crocidura jouvenetae* was considered a subspecies of *C. crossei* by Heim de Balsac & Meester (1977), Hutterer (1993) and Grubb *et al.* (1998), while Hutterer (2005) considers them as two distinct species based on the larger size of *C. jouvenetae*. However, *Cytb* data suggest that these species are not reciprocally monophyletic, highlighting the need for taxonomic revision of this species complex (Igbokwe *et al.* 2019). Ten specimens from Togo cluster together and they form the sister clade of all specimens from Nigeria (96% sequence similarity). We followed the nomenclature used by Igbokwe *et al.* (2019) and named them *C.* cf. *jouvenetae*. One additional specimen for which *Cytb* sequence data was not available was morphologically assigned to *C.* cf. *jouvenetae*.

Five of our *Crocidura* sequences cluster with *C. obscurior* (96% similarity), a species complex reviewed by Jacquet *et al.* (2014).

Jacquet *et al.* (2015) have demonstrated, based on a combination of mitochondrial and nuclear data, that the species *Crocidura olivieri* Lesson, 1827 is polyphyletic and subdivided into five geographical clades with allopatric or parapatric distributions. Our eight sequences cluster with clade II (100% similarity with GenBank KF690215), which includes samples from Niger to Nigeria, west of the Niger River (Jacquet *et al.* 2015; Igbokwe *et al.* 2019). Following the nomenclature adopted by Jacquet *et al.* (2015), we refer to these specimens as *C. olivieri* clade II.

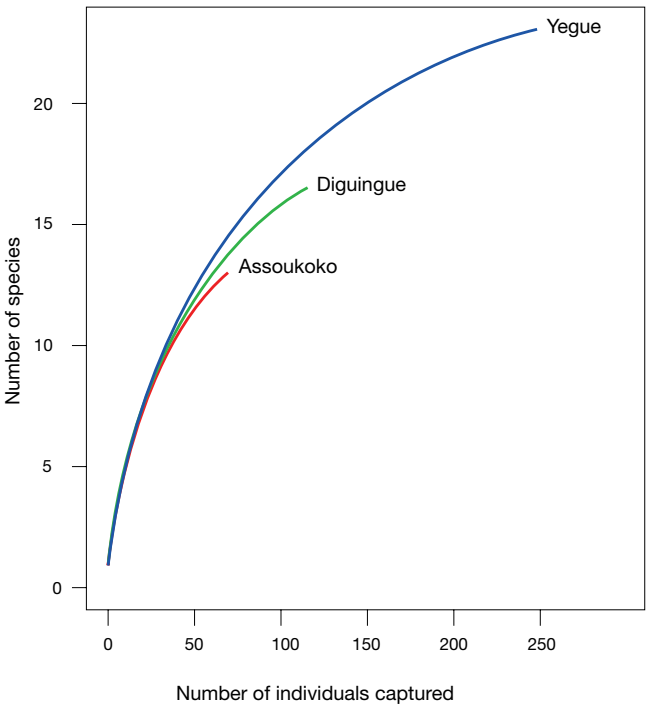


FIG. 2. — Individual-based rarefaction curves of species richness per locality.

The *C. poensis* complex *sensu* Nicolas *et al.* (2019) is a monophyletic group comprising nine distinct lineages (lineages I to IX) corresponding to nine species. Our nine sequences clearly fall within lineage IX (98-99% similarity with GenBank reference sequence MH805963) as defined by these authors, which they assigned to *C. foxi*.

Tissues were not available for the five *Crocidura* specimens referred to as *Crocidura* sp., which prevented molecular identification. We did not attempt a morphological assignment to avoid taxonomic confusion, given the high level of cryptic diversity reported within the genus *Crocidura*.

A complete list of examined material is provided in Appendix 2.

ECOLOGICAL RESULTS

A chi-square test was used to compare the effectiveness of the two main trapping methods used during the study: combined snap and Sherman traps versus pitfall lines. A total of 419 individuals were captured with conventional traps over 11 680 trap-nights, whereas only 13 individuals were captured with pitfall traps over 419 trap-nights. Although conventional traps recorded more captures overall, the difference in capture success was not statistically significant when adjusted for sampling effort ($\chi^2 = 0.15$, df = 1, p = 0.70).

At Assoukoko, trap success was higher in cultivated fields, reaching 4.55% with conventional traps (Sherman and snap traps), compared to only 2.42% in the gallery forest using the same method. In the gallery forest, pitfall traps recorded a higher trap success (3.09%) than conventional traps. At Diguingue, trap success was comparable between gallery forests (1.96% with conventional traps, 3.12% with pitfall traps) and cultivated fields (2.03% with conventional traps). The semi-deciduous dense forest at Diguingue showed the highest trap success with conventional traps (4.17%). At Yegue, a contrasting pattern was observed: trap success was very high in cultivated fields (9.71% with conventional traps), followed by the semi-deciduous dense forest (6.71%), while a marked decline was seen in the gallery forest (1.29%). Overall, across the three sites, Yegue exhibited the highest trap success in cultivated fields.

COMPARISON OF SPECIES RICHNESS BETWEEN HABITATS AND LOCALITIES

Species richness varied notably across habitat types. At Assoukoko, gallery forest harbored the highest species richness, with 11 small mammal species identified among the 46 individuals captured. These included eight rodent species and three shrew species, namely *C. foxi*, *C.* cf. *jouvenetae*, and *C. obscurior*. In contrast, cultivated areas supported only five species, all of which were rodents, with no shrews recorded. *Rattus rattus* was by far the most frequently captured species in these areas, with 17 individuals, followed by four *Lemniscomys striatus*, four *Praomys daltoni*, and one *Mus minutoides*.

At Diguingue, a similar pattern was observed. Gallery forest exhibited higher species richness, with 15 species recorded, including 11 rodents and four shrews, from a total of 71 captures. The shrew species identified were *Crocidura foxi*, *C.* cf. *jouvenetae*, *C. obscurior*, and *C. olivieri* clade II. Additionally, there was a capture of two individuals assigned to *Mus* (*Nannomys*) sp. by their morphology. Cultivated areas hosted 11 species, including two shrews (*C.* cf. *jouvenetae* and *C. olivieri* clade II). Although the semi-deciduous dense forest was less intensively sampled (only eight captures), it revealed the presence of four rodent species, but no shrews.

At Yegue, however, cultivated areas exhibited the highest species richness, with 12 species recorded, including three shrews: *C. foxi*, *C.* cf. *jouvenetae*, and *C. olivieri* clade II. *Mastomys* was by far the most dominant genus, with 89 individuals out of 138 total captures. The semi-deciduous forest hosted 10 species, including two shrew species, while the gallery forest also supported eight species but with a different species composition. In this latter habitat, four shrew species were recorded *C. foxi*, *C. cf. jouvenetae*, *C. obscurior*, and *C. olivieri* clade II alongside only four rodent species.

SPECIES DISTRIBUTION HABITAT

In addition to overall species richness, the distribution of species across habitats and sites reveals ecological preferences and potential indicators of anthropogenic disturbance. Species associated with forested habitats were generally absent from cultivated areas. Several rodents, including *Hylomyscus pamfi*, *Stochomys longicaudatus*, *Gerbilliscus* sp., and *Lophuromys sikapusi*, were captured exclusively in gallery or semi-deciduous dense forests. Similarly, of the 117 individuals of *P. misonnei* captured across the three sites, only six were

found in cultivated areas, and even then, only within plots located inside the core of gallery forest formations, indicating a strong affinity for intact forest habitat. *Cricetomys* sp. 1 was also restricted to gallery forest.

Among other forest-dwelling species, a single *Graphiurus* sp. A was captured in gallery forest, while *Mus baoulei* and *Funisciurus leucostigma* were each represented by one individual also in this habitat.

Species with broad ecological distributions were captured in both natural and modified habitats. *Praomys daltoni* was present at all three sites and occupied a variety of habitats. *Cricetomys gambianus* showed high ecological flexibility, being recorded in gallery forest, semi-deciduous dense forest, and cultivated areas at both Assoukoko and Yegue. *Lemniscomys striatus* occurred across the three sites and across all habitat types, and *Mus minutoides* was likewise found in all habitats surveyed.

Species primarily associated with anthropogenic environments were also identified. *Rattus rattus* was found at Assoukoko, Diguingue, and Yegue, and exclusively in highly anthropized cultivated areas, often near human settlements. *Lemniscomys macculus* and *Graphiurus kelleni* were captured only in cultivated areas, suggesting adaptation to disturbed or open habitats. However, since this is a single capture for these species, we cannot pronounce on their preferred habitat. *Uranomys ruddi* was also captured only once, in a cultivated area at Diguingue.

Within the genus *Mastomys*, *M. erythroleucus* was markedly more abundant than *M. natalensis* across all sites and habitats: 97 individuals versus one at Yegue, 12 versus two at Diguingue, and one versus none at Assoukoko. Both species were found in multiple habitat types, although *M. erythroleucus* appeared more generalist and dominant.

Shrews (*Crocidura*) displayed variable patterns. *C. foxi*, *C.* cf. *jouvenetae*, and *C. obscurior* were widely distributed across all sites and habitats, indicating ecological plasticity. In contrast, *C. olivieri* clade II was captured in multiple habitats of Diguigue and Yegue but not captured at Assoukoko.

Species richness varied according to the sites and habitat types surveyed (Table 3). However, the rarefaction curves (Fig. 2) indicates that species richness did not reach an asymptote at any of the three sampled sites, as evidenced by the absence of a plateau. This is particularly marked for the sites of Assoukoko and Diguingue. Thus, species richness comparisons among sites and habitats should be considered with caution. Despite this limitation, several patterns emerge regarding species distributions. Certain species were found exclusively at a single site. For example, *Graphiurus kelleni*, *G.* sp. A, and *Cricetomys* sp. 1 were recorded only at Diguingue, while *Uranomys ruddi*, *Funisciurus leucostigma*, and *Lemniscomys macculus* were exclusive to Yegue. Likewise, *Euxerus erythropus* was found exclusively at Assoukoko. However, all these species were represented by only a single individual, with the exception of *L. macculus*, for which two individuals were captured. Given their low capture numbers, these species can be considered rare in the sampled communities. Consequently, the absence of certain species at other sites does not necessarily indicate true absence, but may instead reflect low detection probabilities due to rarity or insufficient sampling effort.

Species that showed a broad distribution across all sites include the rodents *Mus minutoides*, *Lemniscomys striatus*, *Praomys daltoni* and shrew species *Crocidura foxi* and *C.* cf. *jouvenetae*, indicating broad ecological tolerance.

Monitoring of *Leimacomys buettneri*

No specimen of *L. buettneri* was captured despite targeted monitoring over several months in Diguingue. Our study was limited by the fact that we did not use insects as bait, although the species is known to be insectivorous. However, we included in our bait smoked fish allowing us to capture other insectivorous species, such as shrews. According to Dieterlen (1976, 2013), *L. buettneri* is known only from two specimens probably collected in the forest-savanna transition zone (ecotone) near the historical Bismarckburg, rather than in dense forest. This species could thus be associated with more open, intermediate habitats rather than strictly forested areas. Furthermore, no specimens have been captured in over a century despite targeted searches in dense forests. By contrast, other strictly forest-dwelling species, such as *Hylomyscus pamfi*, *Stochomys longicaudatus*, and even the rare *Malacomys edwardsi* (Nicolas-Colin & Hounmavo, unpublished data, November 2025), were regularly captured in the same sites using the same methods. This supports the idea that the absence of *L. buettneri* could be due to its habitat preferences rather than methodological limitations.

Contribution of the local community

The local community actively participated in this inventory by capturing 20 small mammal specimens, representing 13 species.

At Assoukoko, nine individuals were collected, including five *Rattus rattus*, one *Praomys daltoni*, one *Mus minutoides*, and one *E. erythropus*, all from the same cultivated habitat; one *C. gambianus* was collected in gallery forest.

At Diguingue six specimens were collected by the community, including three individuals from the genus *Graphiurus*: the first, *G. kelleni* (JCK11281) from cultivated area, the second *Graphiurus* sp. B (AH201), also from cultivated area, the third, *Graphiurus* sp. A, (JCK11228) from gallery forest. Additionally, one *Mastomys natalensis*, one *Lemniscomys striatus*, and one *Gerbilliscus* sp. were captured, the two first species, in cultivated areas and the last in gallery forest.

At Yegue, five individuals were captured by the community, including two *Lophuromys sikapusi* and one *Funisciurus leucostigma* in the gallery forest, as well as one *Mus minutoides* and one *Crocidura olivieri* clade II in cultivated areas.

The involvement of the local community proved to be valuable, as it led to the capture of individuals from five species that were not recorded using conventional traps (Sherman and snap traps) or pitfall traps: *G. kelleni*, *Graphiurus* sp. B, *Graphiurus* sp. A, *Euxerus erythropus* and *Funisciurus leucostigma*. Their contribution thus significantly enriched the faunal inventory.



FIG. 3. — Dorsal and ventral views of three specimens of the genus *Graphiurus* Smuts, 1837: from left to right, JCK11281 (*G. kelleni* (Reuvens, 1890)), AH201 (*Graphiurus* sp. B), and JCK11238 (*Graphiurus* sp. A). Scale bar: 1 cm. Photo credit: Arnaud Delapré (MNHN, France).

DISCUSSION

NEW RECORDS FOR TOGO AND RANGE EXTENSION

We captured 23 rodent species and five shrew species in Assoukoko Forest. Among these 28 species recorded in this survey, five are reported for the first time in Togo. Four of them are already known to science, *Crocidura* cf. *jouvenetae*, *C. obscurior*, *Lemniscomys macculus* and *Gerbilliscus* sp. which is the lineage cleary delimited by Granjon *et al.* (2012), but not yet formally described, while the fifth is a potentially new species belonging to the genus *Graphiurus*.

The presence of *C.* cf. *jouvenetae* in Togo is not surprising, as the species is well documented in West Africa, particularly in Côte d'Ivoire, Guinea, and Liberia (Akpatou *et al.* 2019; Denys *et al.* 2021) and it was recently reported in Nigeria by Igbokwe *et al.* (2019). This latter record extended its range eastward, making its presence in Togo plausible.

Regarding *C. obscurior*, a typically forest-dwelling species, Jacquet *et al.* (2014) examined its genetic and morphological diversity in Guinea, Liberia, Côte d'Ivoire, and southwestern Ghana. The Mammalian Diversity Database (https://www.mammaldiversity.org) and Hutterer (2005) suggest that this species may also be present in Nigeria. This hypothesis is based on the record by Hutterer & Happold (1983) of an African pygmy shrew called *C. bottegi*, a species now considered to be endemic to East Africa. Our record from central Togo represents a significant eastward extension of the known range of *C. obscurior* beyond that documented by Jacquet *et al.* (2014). This finding aligns with the hypothesis that *C. obscurior* is the pygmy shrew present in Nigeria. However, molecular data would be required to confirm this identification.

The discovery of *L. macculus* in Togo (in the gallery forest of Yegue) is also novel, although its presence has been previously confirmed in western Côte d'Ivoire (Lamto), Guinea, southwestern Ghana, and Benin (Hánová *et al.* 2021).

The discovery of *Gerbilliscus* sp. in Togo is also not a surprise because Granjon *et al.* (2012) have already reported it from Guinea, Côte d'Ivoire, and Benin. Our results once again highlight the existence of this lineage and also highlight the urgency of obtaining authorisation for sequencing the tissues of type specimens of *Gerbilliscus kempi* and *Gerbilliscus giffardi*, and all their synonyms, in order to proceed with the formal identification of this lineage, clearly defined since 2012 by Granjon *et al.*, but which has remained without valid taxonomic status since then. It should also be noted that *Gerbilliscus kempi* and *G. guineae* have previously been reported from Togo (Amori *et al.* 2016), further emphasizing the taxonomic complexity of *Gerbilliscus* in the region and the need for molecular data from type material to resolve species limits.

Finally, the most remarkable finding concerns one *Graphiurus* specimen (JCK11238) whose sequence is very different from all *Graphiurus* sequences available in GenBank. The closest similarity (83.61%) was obtained with *Spermophilus*.

Graphiurus is the sole current genus for dormice in sub-Saharan Africa, but the taxonomy of its species remains unstable (Holden 1996; Holden & Levine 2009; Monadjem *et al.* 2015). Currently 15 species are recognized as valid, but 121 species names are available in Mammalian Diversity Database (https://www.mammaldiversity.org), and there is no well-supported molecular phylogeny allowing the evaluation of species boundaries using the phylogenetic species concept. Our preliminary results suggest that we have captured in Togo either a species new to science (or genus, as the branch is deeply divided) or a species for which there is not reference sequence yet in Genbank. A revision of this genus based on molecular data is urgently needed. The availability of the two *Graphiurus* sequences, one of which is unknown in the Genbank database and the other referring to *G. kelleni*, will contribute to this revision. Examination of morphology, tail shape, and coloration of ventral fur, support the possibility that the third specimen of *Graphiurus* (AH 201), for which it was not possible to perform DNA sequencing, may represent a third species of *Graphurius* from our surveys (Fig. 3). Given the low number of dormice specimens in museums around the world, this inventory enriches museum collections of African dormice.

Molecular confirmation of previously reported species

Beyond these five taxa reported for the first time in Togo, including a species of African dormouse potentially new to science, our results provide the first molecular evidence for several other taxa previously reported from the country. For instance, until now, *Mus baoulei* was known in Togo only from a single specimen collected at Ounabé in the Akposso Mountains, mentioned by Robbins & Van der Straeten (1996) and cited by Amori *et al.* (2016). Our study provides the first molecular confirmation of the presence of *M. baoulei* in Togo, based on seven sequences clearly assigned to MOTU12 of Bryja *et al.* (2014), which unambiguously corresponds to this species.

In addition, the identification of 16 genotyped individuals of *Mus minutoides*, with sequences clustering within MOTU27 of Bryja *et al.* (2014), expands knowledge of its range. Although this species was reported by Amori *et al.* (2016) from specimens collected at Pewa, Pagala, Ezimé, and Agou (deposited at MRAC), it was absent from the species list of Robbins & Van der Straeten (1996), and was considered absent from West Africa by Musser & Carleton (2005). Our results thus unequivocally confirm the presence of *M. minutoides* in Togo.

Similarly, our study confirms the presence of *Cricetomys gambianus* in central Togo, based on two new sequences. Although sequences of *C. gambianus* from northern Togo were already available thanks to the work of Olayemi *et al.* (2012), our data represent the first molecular confirmation for central Togo.

Even more remarkable is one of our sequences that clusters with *Cricetomys* sp. 1, an undescribed lineage identified by Olayemi *et al.* (2012). This lineage had, until now, only been detected in southern Togo. The capture of our specimen in the central region significantly extends the known range of this cryptic taxon. According to the distribution maps published by these authors, *C. gambianus* has a predominantly savanna distribution in West Africa, whereas *Cricetomys* sp. 1 is associated with forest habitat. Our findings thus indicate that both taxa occur sympatrically in central Togo. This range extension may also have been facilitated by recent anthropogenic environmental changes, including deforestation and climate change, which are known to affect habitat connectivity and species distributions in West Africa.

Our study also provides the first molecular confirmation for several other taxa previously identified based on morphology only in Togo, including: *Stochomys longicaudatus*, *Uranomys ruddi*, *Rattus rattus*, *Exerus erythropus*, and *Funisciurus leucostigma*. Finally, two species of the genus *Crocidura*, *C. olivieri* clade II and *C. foxi*, although reported in Togo by Amori *et al.* (2016), are confirmed for the first time with molecular data.

Squirrel species sampling limitations

Only two squirrel species were captured: *Funisciurus leucostigma* and *Euxerus erythropus*. The presence of *F. leucostigma* in Togo was not explicitly anticipated by Amori *et al.* (2016), as no voucher specimen or genetic sequence was available prior to our study. Amori *et al.* (2016) listed *F. pyrropus* from Bismarkburg in Togo based on a specimen housed at the ZMB and *F. anerythrus* from Dapaong. *Funisciurus leucostigma* has historically been treated as a subspecies of *F. pyrropus*, but Curaudeau *et al.* (2025) clarified that *F. leucostigma* is the only *Funisciurus* species occurring in West Africa, including Togo, whereas *F. anerythrus* and *F. pyrropus* are restricted to Central Africa east of the Sanaga River. Thus, our record confirms *F. leucostigma* in the country and likely reassigns the historical ZMB specimen from Bismarckburg previously identified as *F. pyrropus* to *F. leucostigma*. This also reinforces that *F. anerythrus* probably does not occur in Togo, a point consistent with previous assessments (Amori *et al.* 2016; Thorington & Hoffmann 2005). Therefore, the specimen collected in Dapaong, which is assigned the name *F. anerythrus*, would probably be *F. leucostigma*.

The number of squirrel species we captured is significantly lower than the six squirrel species (*F. leucogenys* Waterhouse, 1842; *F. pyrropus* Cuvier, 1833; *F. substriatus* de Winton, 1899; *Heliosciurus gambianus* Waterhouse, 1842; *H. rufobrachium* Waterhouse, 1842 and *Protoxerus stangeri* Waterhouse, 1842) reported in Togo by Amori *et al.* (2016) as potentially present in forest Zone IV. This underestimation is likely due to our trapping methods. Medium-sized Sherman traps and ground-based snap traps are largely ineffective at capturing arboreal species of this size. More suitable methods, such as large Sherman or Tomahawk traps placed in trees or the use of shotguns are necessary for better coverage of these taxa (Jacques *et al.* 2017). In addition, widespread deforestation and intense hunting pressure on squirrels in the region may have contributed to their low capture rates, as these pressures can reduce local abundance and increase trap wariness.

Indicators of habitat disturbance

The high frequency of *Mastomys erythroleucus* and *M. natalensis* (113 individuals out of 452 captures) indicates a high level of habitat disturbance. *Mastomys erythroleucus* is a generalist species typically found in dry or moist savannas and agricultural areas (Brouat *et al.* 2007), with strong ability to colonize disturbed habitats (Assefa & Chelmala 2019). This pattern was particularly pronounced at the Yegue site, where 97 individuals of *M. erythroleucus* and only one *M. natalensis* were captured. The dominance of these species, often associated with agricultural or heavily anthropized landscapes, is consistent with local accounts of recent vegetation degradation. Similarly, the capture of several *Rattus rattus* individuals, an allochthonous, highly synanthropic species, reflects increased anthropogenic disturbance at the site. According to local residents, *R. rattus* was not observed in the area a decade ago suggesting recent human-mediated introduction.

Conservation value of Assoukoko Forest

The remarkable small mammal diversity of Assoukoko Forest highlights the importance of this protected area for terrestrial mammal conservation. Despite increasing pressure from human activities, including charcoal production and agricultural expansion, this forest continues to support a diverse and ecologically representative small mammal community. However, there are severe threats. Some sections of the Yegue River gallery forest are only 10-15 m wide as farmers routinely set fires to encourage agricultural growth, and this slowly but steadily destroys this vital and limited habitat. Dowsett-Lemaire & Dowsett (2011) similarly reported that the forest remains a refuge for many bird species despite intense deforestation pressures. The coexistence of forest species like the shrews *Crocidura obscurior*, *C.* cf. *jouvenetae*, and rodents like *Hylomyscus pamfi*, *Praomys misonnei*, and *Cricetomys* sp. 1 supports this hypothesis. These species are sensitive to fragmentation and disturbance, and their presence suggests stable and high-quality forest habitat (Nicolas *et al.* 2010; Jacquet *et al.* 2014).

Thus, despite the obvious signs of degradation, the Assoukoko forest retains high ecological value, especially for forest species. As such, it deserves special attention in terms of conservation, sustainable management, and long-term monitoring efforts.

Monitoring and the absence of *Leimacomys buettneri*

No individuals of *Leimacomys buettneri* were captured despite consistent sampling efforts conducted at Diguingue sites over an eight-month period. This absence suggests local extirpation of the species. The type locality of *L. buettneri*, historically referred to as Bismarckburg, corresponds today to the Adélé area. However, the precise location of collection was never clearly documented, leaving room for multiple hypotheses regarding its actual distribution. It is possible that the original specimens were collected from a different area than what is currently identified as Bismarckburg. Another hypothesis is that the species may have taken refuge in the relatively well-preserved sacred forests near Pagala. It is possible that *L. buettneri* is a savanna species. In this context, it would be advantageous to conduct targeted surveys in the Pagala region, as well as in poorly explored areas of ecoregion IV, including the forests of Misahohé, Akloa, and Deux Bena, and especially targeting savanna habitats.

Authors contribution

The study was conceived by AH, VN, GHS, JCK, TD, and HL. Fieldwork was conducted by AH, JCK, TD, and HL. Molecular lab work and molecular analyses were carried out by AH and VN in France, and by AH, TD, and HL in the USA. As part of his taxonomic revision of the genus *Graphiurus*, KO compared the sequence of specimen JCK11238 with all available reference sequences in the databases accessible to him, suggesting that this specimen may represent a probable new species to science. The photo of the specimens of *Graphiurus* was taken by AD. The first draft of the manuscript was written by AH, VN, JCK, and TD, and then improved by all co-authors. TCD, JCK and VN contributed equally to the manuscript.

Acknowledgements

We sincerely thank the Ministry of Environment of Togo for granting the research permits N° 009/DRF/2023 et N° 010/DRF/ 2023 and facilitating access to the Assoukoko Forest, which made this inventory possible. Our sincere thanks go also to Guillaume K. Ketoh, Director of the Laboratory of Ecology and Ecotoxicology at the University of Lomé, for facilitating student participation in the fieldwork, particularly Mawuegnigan Valentin, Issaka Rakib, and Abdou Rachid. We are especially grateful to Mr. Brousounou Djinsa, an enthusiastic naturalist from Yegue, for his assistance in the field. We further thank A. Krokan (Norwegian Ministry of Foreign Affairs) and her son F. Krokan for their personal and field support in 2022. We also appreciate the logistical support provided by Éric Goulignac (Heidelberg Materials). We are also very grateful to the two anonymous reviewers for their thorough, constructive, and insightful comments, which greatly improved the quality and clarity of this manuscript.

REFERENCES

Akpatou K. B., Bohoussou K. H. & Bene K. J.-C. 2019. — Assessment of terrestrial small mammals in an agro-industrial company concession, Western Liberia. *International Journal of Applied Sciences and Biotechnology* 7 (4): 434-439. https://doi.org/10.3126/ijasbt.v7i4.25581

Amori G., Segniagbeto G. H., Decher J., Assou D., Gippoliti S. & Luiselli L. 2016. — Non-marine mammals of Togo (West Africa): an annotated checklist. *Zoosystema* 38 (2): 201-244. https://doi.org/10.5252/z2016n2a3

Assefa A. & Chelmala S. 2019. — Comparison of rodent community between natural and modified habitats in Kafta-Sheraro National Park and its adjoining villages, Ethiopia: implication

for conservation. *The Journal of Basic and Applied Zoology* 80 (1): 59. https://doi.org/10.1186/s41936-019-0128-9

Brouat C., Loiseau A., Kane M., Bâ K. & Duplantier J. 2007. — Population genetic structure of two ecologically distinct multimammate rats: the commensal *Mastomys natalensis* and the wild *Mastomys erythroleucus* in southeastern Senegal. *Molecular Ecology* 16 (14): 2985-2997. https://doi.org/10.1111/j.1365-294x.2007.03353.x

Bryja J., Granjon L., Dobigny G., Patzenhauerová H., Konečný A., Duplantier J. M., Gauthier P., Colyn M., Durnez L., Lalis A. & Nicolas V. 2010. — Plio-Pleistocene history of West African Sudanian savanna and the phylogeography of the *Praomys daltoni* complex (Rodentia): the environment/geography/genetic interplay. *Molecular Ecology* 19 (21): 4783-4799. https://doi.org/10.1111/j.1365-294x.2010.04847.x

Bryja J., Mikula O., Šumbera R., Meheretu Y., Aghová T., Lavrenchenko L. A., Mazoch V., Oguge N., Mbau J. S., Welegerima K., Amundala N., Colyn M., Leirs H. & Verheyen E. 2014. — Pan-African phylogeny of *Mus* (subgenus *Nannomys*) reveals one of the most successful mammal radiations in Africa. *BMC Evolutionary Biology* 14 (1): 256. https://doi.org/10.1186/s12862-014-0256-2

Čepelka L. & Dokulilová M. 2025. — Ecological roles and forest management implications of small terrestrial mammals in temperate and boreal forests – A review. *Forests* 16 (6): 994. https://doi.org/10.3390/f16060994

Colangelo P., Abiadh A., Aloise G., Amori G., Capizzi D., Vasa E., Annesi F. & Castiglia R. 2015. — Mitochondrial phylogeography of the black rat supports a single invasion of the western Mediterranean basin. *Biological Invasions* 17 (6): 1859-1868. https://doi.org/10.1007/s10530-015-0842-2

Colyn M., Hulselmans J., Sonet G., Oude P., De Winter J., Natta A., Nagy Z. T. & Verheyen E. 2010. — Discovery of a new duiker species (Bovidae: Cephalophinae) from the Dahomey Gap, West Africa. *Zootaxa* 2637 (1): 1-30. https://doi.org/10.11646/zootaxa.2637.1.1

Curaudeau M., Peterhans J. K., Le Flanchec T., Gilissen E., Granjon L., Barale L., Gessain A. & Hassanin A. 2025. — Squirrel reservoirs of monkeypox virus are sister species separated by the Sanaga River (Cameroon), as are the two main viral clades. *One Health* 101157. https://doi.org/10.2139/ssrn.5262613

Davies G. T. O., Kirkpatrick J. B., Cameron E. Z., Carver S. & Johnson C. N. 2019. — Ecosystem engineering by digging mammals: effects on soil fertility and condition in Tasmanian temperate woodland. *Royal Society Open Science* 6 (1): 180621. https://doi.org/10.1098/rsos.180621

Decher J., Norris R. W., Abedi-Lartey M., Oppong J., Hutterer R., Weinbrenner M., Koch M., Podsiadlowski L. & Kilpatrick C. W. 2021. — A survey of small mammals in the Volta Region of Ghana with comments on zoogeography and conservation. *Zoosystema* 43 (14): 253-281. https://doi.org/10.5252/zoosystema2021v43a14. http://zoosystema.com/43/14

Denys C., Jacquet F., Kadjo B., Missoup A. D., Aniskine V., de Bellocq J. G., Soropogui B., Douno M., Sylla M. & Nicolas V. 2021. — Shrews (Mammalia, Eulipotyphla) from a biodiversity hotspot, Mount Nimba (West Africa), with a field identification key to species. *Zoosystema* 43 (30): 729-757. https://doi.org/10.5252/zoosystema2021v43a30. http://zoosystema.com/43/30

Denys C., Lalis A., Aniskin V., Gerbault-Seureau M., Delapre A., Gilissen E., Merker S. & Nicolas V. 2020. — Integrative taxonomy of Guinean *Lemniscomys* species (Mammalia). *Journal of Vertebrate Biology* 69 (2): 20008-1. https://doi.org/10.25225/jvb.20008

Dieterlen F. 1976. — Bemerkungen über *Leimacomys büttneri* Matschie, 1893 (Dendromurinae, Cricetidae, Rodentia). *Säugetierkundliche Mitteilungen* 24: 224-228.

Dieterlen F. 2013. — *Leimacomys buettneri* Büttner's Forest Mouse (Büttner's Togo Mouse), *in* Happold D. C. D. (Ed.): *Mammals of Africa, Vol. III: Rodents, Hares and Rabbits*. London, Bloomsbury: 359-360.

Dowsett-Lemaire F. & Dowsett J. R. 2016. — Exploration ornithologique du Bénin et du Togo de mars à début août 2016. *Miscellaneous Publications* 87: 33.

Dowsett-Lemaire F. & Dowsett R. J. 2011. — Ornithological expedition to Togo 2010–2011. Dowsett-Lemaire *Miscellaneous Report* 78: 1-35.

Ducroz J. F., Volobouev V. & Granjon L. 2001. — An assessment of the systematics of arvicanthine rodents using mitochondrial DNA sequences: evolutionary and biogeographical implications. *Journal of Mammalian Evolution* 8 (3): 173-206. https://doi.org/10.1023/a:1012277012303

Ern H. 1979. — Die Vegetation Togos. Gliederung, Gefährdung, Erhaltung. *Willdenowia* 295-312.

Gaubert P., Njiokou F., Ngua G., Afiademanyo K., Dufour S., Malekani J., Bi S. G., Tougard C., Olayemi A., Danquah E. Djagoun C. A., Kaleme P., Mololo C.N., Stanley W., Luo S. J. & Antunes A. 2016. — Phylogeography of the heavily poached African common pangolin (Pholidota, *Manis tricuspis*) reveals six cryptic lineages as traceable signatures of Pleistocene diversification. *Molecular Ecology* 25 (23): 5975. https://doi.org/10.1111/mec.13886

Godó L., Valkó O., Borza S. & Deák B. 2022. — A global review on the role of small rodents and lagomorphs (clade Glires) in seed dispersal and plant establishment. *Global Ecology and Conservation* 33: e01982. https://doi.org/10.1016/j.gecco.2021.e01982

Granjon L., Colangelo P., Tatard C., Colyn M., Dobigny G. & Nicolas V. 2012. — Intrageneric relationships within *Gerbilliscus* (Rodentia, Muridae, Gerbillinae), with characterization of an additional West African species. *Zootaxa* 3325 (1): 1-25. https://doi.org/10.11646/zootaxa.3325.1.1

Grubb P., Jones T. S., Davies A. G., Edberg E., Starin E. D. & Hill J. E. 1998. — *Mammals of Ghana, Sierra Leone and the Gambia*. Trendrine Press, Cornwall, 265 p.

Hánová A., Konečný A., Nicolas V., Denys C., Granjon L., Lavrenchenko L. A., Šumbera R., Mikula O. & Bryja J. 2021. — Multilocus phylogeny of African striped grass mice (*Lemniscomys*): stripe pattern only partly reflects evolutionary relationships. *Molecular Phylogenetics and Evolution* 155: 107007. https://doi.org/10.1016/j.ympev.2020.107007

Heim de Balsac H. & Meester J. 1977. — Part I: Order Insectivora, *in* Meester J., & Setzer H. W. (eds). *The Mammals of Africa: an Identification Manual*. Smithsonian Institution Press, Washington, D.C.: 1-29.

Herron M. D. & Waterman J. M. 2004. — *Xerus erythropus*. *Mammalian Species* 2004 (748): 1-4. https://doi.org/10.1644/748

Holden M. E. 1996. — *Systematic Revision of Sub-Saharan African Dormice (Rodentia, Myoxidae,* Graphiurus*). Part 1, An Introduction to the Generic Revision, and a Revision of* Graphiurus surdus. American Museum novitates 3157. New York, N.Y, American Museum of Natural History. https://www.biodiversitylibrary.org/item/314697

Holden M. E. & Levine R. S. 2009. — Systematic revision of sub-saharan African Dormice (Rodentia: Gliridae: *Graphiurus*) Part II: description of a new species of *Graphiurus* from the Central Congo Basin, including morphological and ecological niche comparisons with *G. crassicaudatus* and *G. lorraineus*. *Bulletin of the American Museum of Natural History* 2009 (331): 314-355. https://doi.org/10.1206/582-9.1

Hounmavo A., Demos T. C., Lutz H. L., Delapré A., Cornette R., Segniagbeto G. H., Kerbis Peterhans J. C. & Nicolas V. 2026. — Integrative taxonomy of the *Praomys tullbergi* complex (Rodentia, Muridae): a new species from West Africa. *Journal of Mammalogy* 107 (2): 368-382. https://doi.org/10.1093/jmammal/gyag007

Hurst J., Bowes L. D. & O'Keefe E. 1995. — Kyabobo'94: a joint Ghanaian and Oxford University Expedition. Final Report.

KCP Publications, Ghana Wildlife Department, Institute for Renewable Natural Resources, 69 p.

HUTTERER R. 1993. — Order Insectivora, *in* WILSON D. E. & REEDER D. M. (Eds.), *Mammal Species of the World: A Taxonomic and Geographic Reference* (2nd ed.). Smithsonian Institution Press, Washington, DC: 69-130.

HUTTERER R. 2005. — Order Soricomorpha, *in* WILSON D. & REEDER D. (Eds). *Mammal Species of the World: A Taxonomic and Geographic Reference*. Baltimore: Johns Hopkins University Press: 220-310.

HUTTERER R. & HAPPOLD D. C. D. 1983. — The shrews of Nigeria (Mammalia: Soricidae). *Bonner Zoologische Monographien* 18: 5-79.

IGBOKWE J., NICOLAS V., OYEYIOLA A., OBADARE A., ADESINA A. S., AWODIRAN M. O., VAN HOUTTE N., FICHET-CALVET E., VERHEYEN E. & OLAYEMI A. 2019. — Molecular taxonomy of *Crocidura* species (Eulipotyphla: Soricidae) in a key biogeographical region for African shrews, Nigeria. *Comptes rendus Biologies* 342 (3-4): 108-117. https://doi.org/10.1016/j.crvi.2019.03.004

IGBOKWE J. O., AWODIRAN M. O., OLADEJO O. S., OLAYEMI A. O. & AWOPETU J. I. 2016. — Chromosomal analysis of small mammals from Southwestern Nigeria. *Mammal Research* 61 (2): 153-159. https://doi.org/10.1007/s13364-015-0254-9

JACQUES C. N., ZWEEP J. S., SCHEIHING M. E., RECHKEMMER W. T., JENKINS S. E., KLAVER R. W. & DUBAY S. A. 2017. — Influence of trap modifications and environmental predictors on capture success of southern flying squirrels. *Wildlife Society Bulletin* 41 (2): 313-321. https://doi.org/10.1002/wsb.769

JACQUET F., DENYS C., VERHEYEN E., BRYJA J., HUTTERER R., KERBIS PETERHANS J. C., STANLEY W. T., GOODMAN S. M., COULOUX A., COLYN M. & NICOLAS V. 2015. — Phylogeography and evolutionary history of the *Crocidura olivieri* complex (Mammalia, Soricomorpha): from a forest origin to broad ecological expansion across Africa. BMC *Evolutionary Biology* 15 (1). https://doi.org/10.1186/s12862-015-0344-y

JACQUET F., NICOLAS V., COLYN M., KADJO B., HUTTERER R., DECHER J., AKPATOU B., CRUAUD C. & DENYS C. 2014. — Forest refugia and riverine barriers promote diversification in the West African pygmy shrew (*Crocidura obscurior* complex, Soricomorpha). *Zoologica Scripta* 43 (2): 131-148. https://doi.org/10.1111/zsc.12039

KRELL F. 1994. — Bismarckburg–two famous collecting localities in Africa. *Entomologist's Monthly Magazine* 130: 55-58.

KRYŠTUFEK B., MAHMOUDI A., TESAKOV A. S., MATĚJŮ J. & HUTTERER R. 2016. — A review of bristly ground squirrels Xerini and a generic revision in the African genus *Xerus*. *Mammalia* 80 (5): 521-540. https://doi.org/10.1515/mammalia-2015-0073

MIKULA O., NICOLAS V., BORATYŃSKI Z., DENYS C., DOBIGNY G., FICHET-CALVET E., GAGARÉ S., HUTTERER R., NIMO-PAINTSIL S. C. & OLAYEMI A. 2020. — Commensalism outweighs phylogeographical structure in its effect on phenotype of a Sudanian savanna rodent. *Biological Journal of the Linnean Society* 129 (4): 931-949. https://doi.org/10.1093/biolinnean/blz184

MONADJEM A., TAYLOR P. J., DENYS C. & COTTERILL F. P. 2015. — Rodents of sub-Saharan Africa: a biogeographic and taxonomic synthesis. Walter de Gruyter, 1102 p. https://doi.org/10.1515/9783110301915

MUSSER G. G. & CARLETON M. D. 2005. — *In* WILSON D. E. & REEDER D. M. *Mammal species of the world: a taxonomic and geographic reference.* The Johns Hopkins University Press, Baltimore, 2000 p.

NICOLAS V., BARRIÈRE P. & COLYN M. 2003. — Impact of removal pitfall trapping on the community of shrews (Mammalia: Soricidae) in two African tropical forest sites. *Mammalia* 67 (1): 133-138. https://doi.org/10.1515/mamm.2003.67.1.133

NICOLAS V., MBOUMBA J., VERHEYEN E., DENYS C., LECOMPTE E., OLAYEMI A., MISSOUP A. D., KATUALA P. & COLYN M. 2008. — Phylogeographic structure and regional history of *Lemniscomys striatus* (Rodentia: Muridae) in tropical Africa. *Journal of Biogeography* 35 (11): 2074-2089. https://doi.org/10.1111/j.1365-2699.2008.01950.x

NICOLAS V., OLAYEMI A., WENDELEN W. & COLYN M. 2010. — Mitochondrial DNA and morphometrical identification of a new species of *Hylomyscus* (Rodentia: Muridae) from West Africa. *Zootaxa* 2579 (1): 30-44. https://doi.org/10.11646/zootaxa.2579.1.2

NICOLAS V., MISSOUP A. D., DENYS C., KERBIS PETERHANS J., KATUALA P., COULOUX A. & COLYN M. 2011. — The roles of rivers and Pleistocene refugia in shaping genetic diversity in *Praomys misonnei* in tropical Africa: Phylogeography of *Praomys misonnei* in tropical Africa. *Journal of Biogeography* 38 (1): 191-207. https://doi.org/10.1111/j.1365-2699.2010.02399.x

NICOLAS V., SCHAEFFER B., MISSOUP A. D., KENNIS J., COLYN M., DENYS C., TATARD C., CRUAUD C. & LAREDO C. 2012. — Assessment of three mitochondrial genes (16S, Cytb, CO1) for identifying species in the Praomyini tribe (Rodentia: Muridae). *PLoS ONE* 7 (5):e36586. https://doi.org/10.1371/journal.pone.0036586

NICOLAS V., JACQUET F., HUTTERER R., KONEČNÝ A., KAN KOUASSI S., DURNEZ L., LALIS A., COLYN M. & DENYS C. 2019. — Multilocus phylogeny of the *Crocidura poensis* species complex (Mammalia, Eulipotyphla): Influences of the palaeoclimate on its diversification and evolution. *Journal of Biogeography* 46 (5): 871-883. https://doi.org/10.1111/jbi.13534

NICOLAS V., FABRE P-H., BRYJA J., DENYS C., VERHEYEN E., MISSOUP A. D., OLAYEMI A., KATUALA P., DUDU A. & COLYN M. 2020. — The phylogeny of the African wood mice (Muridae, *Hylomyscus*) based on complete mitochondrial genomes and five nuclear genes reveals their evolutionary history and undescribed diversity. *Molecular Phylogenetics and Evolution* 144: 106703. https://doi.org/10.1016/j.ympev.2019.106703

NICOLAS V., MIKULA O., LAVRENCHENKO L. A., ŠUMBERA R., BARTÁKOVÁ V., BRYJOVÁ A., MEHERETU Y., VERHEYEN E., MISSOUP A. D. & LEMMON A. R. 2021. — Phylogenomics of African radiation of Praomyini (Muridae: Murinae) rodents: First fully resolved phylogeny, evolutionary history and delimitation of extant genera. *Molecular Phylogenetics and Evolution* 163: 107263. https://doi.org/10.1016/j.ympev.2021.107263

OATES J. F., WOODMAN N., GAUBERT P., SARGIS E. J., WIAFE E. D., LECOMPTE E., DOWSETT-LEMAIRE F., DOWSETT R. J., GONEDELÉ BI S. & IKEMEH R. A. 2022. — A new species of tree hyrax (Procaviidae: *Dendrohyrax*) from West Africa and the significance of the Niger–Volta interfluvium in mammalian biogeography. *Zoological Journal of the Linnean Society* 194 (2): 527-552. https://doi.org/10.1093/zoolinnean/zlab029

OKSANEN J., SIMPSON G. L., BLANCHET F. G., KINDT R., LEGENDRE P., MINCHIN P. R., O'HARA R. B., SOLYMOS P., STEVENS M. H. H., SZOECS E., WAGNER H., BARBOUR M., BEDWARD M., BOLKER B., BORCARD D., BORMAN T., CARVALHO G., CHIRICO M., DE CACERES M., DURAND S., EVANGELISTA H., FITZJOHN R., FRIENDLY M., FURNEAUX B., HANNIGAN G., HILL M. O., LAHTI L., MARTINO C., MCGLINN D., OUELLETTE M. H., RIBEIRO CUNHA E., SMITH T., STIER A., TER BRAAK C. J. F. & WEEDON J. 2025. — Vegan: Community Ecology Package Version 2.7-2. https://doi.org/10.32614/CRAN.package.vegan

OLAYEMI A., NICOLAS V., HULSELMANS J. A. N., MISSOUP A. D., FICHET-CALVET E., AMUNDALA D., DUDU A., DIERCKX T., WENDELEN WI. M. & LEIRS H. 2012. — Taxonomy of the African giant pouched rats (Nesomyidae: *Cricetomys*): molecular and craniometric evidence support an unexpected high species diversity. *Zoological Journal of the Linnean Society* 165 (3): 700-719. https://doi.org/10.1111/j.1096-3642.2012.00823.x

ONDITI K. O., DEMOS T. C., KERBIS PETERHANS J., CHEN Z.-Z., BRYJA J., LAVRENCHENKO L. A., MUSILA S., VERHEYEN E., VAN DE PERRE F., AKAIBE BD, DE LA SANCHA N. U. & JIANG X.-L. 2021. — Historical biogeography, systematics, and integrative taxonomy of the non-Ethiopian speckled pelage brush-furred

rats (*Lophuromys flavopunctatus* group). *BMC Ecology and Evolution* 21 (1): 89. https://doi.org/10.1186/s12862-021-01813-w

PETTER F. 1963. — Capture d'un *Uranomys* (Rongeurs, Muridés) dans la République du Togo. *Mammalia* 27: 610.

ROBBINS C. B. & VAN DER STRAETEN E. 1996. — Small mammals of Togo and Benin. II. Rodentia. *Mammalia* 60 (2): 231-242. https://doi.org/10.1515/mamm.1996.60.2.231

SCHMIDT D. F., LUDWIG C. A. & CARLETON M. D. 2008. — The Smithsonian Institution African Mammal Project (1961-1972): an annotated gazetteer of collecting localities and summary of its taxonomic and geographic scope. *Smithsonian Contributions to Zoology* 628: 1-244. https://doi.org/10.5479/si.00810282.628

SEGNIAGBETO G. H., ASSOU D., KODA K. D., AGBESSI EK. G., ATSRI K. H., DENDI D., LUISELLI L., DECHER J. & MITTERMEIER R. A. 2017. — Survey of the status and distribution of primates in Togo (West Africa). *Biodiversity* 18 (4): 137-150. https://doi.org/10.1080/14888386.2017.1404930

UEBBING S. 2019. — *Stochomys longicaudatus* (Rodentia: Muridae). *Mammalian Species* 51 (974): 26-33. https://doi.org/10.1093/mspecies/sez004

THORINGTON JR R. W. & HOFFMANN R. S. 2005. — Family sciuridae, *in* WILSON D. & REEDER D. (eds), *Mammal Species of the World. A Taxonomic and Geographic Reference*. Johns Hopkins University Press, Baltimore, 754-818.

VAN DER STRAETEN E. & VERHEYEN W. N. 1978. — Taxonomical notes on the West-African *Myomys* with the description of *Myomys derooi* (Mammalia - Muridae). *Zeitschrift für Säugetierkunde* 43: 31-41.

VERHEYEN W. N., HULSELMANS J. L., DIERCKX T., MULUNGU L., LEIRS H., CORTI M. & VERHEYEN E. 2007. — The characterization of the Kilimanjaro *Lophuromys aquilus* True 1892 population and the description of five new *Lophuromys* species (Rodentia, Muridae). *Koninklijk Belgisch Instituut Voor Natuurwetenschappen. Studiedocumenten* 77: 23-75.

Submitted on 13 August 2025;
accepted on 11 June 2026;
published on 6 November 2026.

APPENDICES

APPENDIX 1. — List of non-volant mammal species reported by Amori *et al.* (2016) from ecological zone IV of Togo. IUCN indicates the conservation status according to the International Union for Conservation of Nature Red List (**LC**, least concern; **DD**, data deficient). Number of specimens refers to the number of voucher specimens reported by Amori *et al.* (2016). Museums indicates the institution(s) holding the voucher specimens. Abbreviations: **MNHN**, Muséum national d'Histoire naturelle, Paris; **MRAC**, Musée royal de l'Afrique centrale, Tervuren; **NMNH**, National Museum of Natural History, Washington, D.C.; **ZMB**, Museum für Naturkunde, Berlin. "Not mentioned" indicates that the information was not provided by Amori *et al.* (2016).

Genus	Species	IUCN	Number of specimens	Museums
Atelerix	*albiventris*	LC	39	NMNH
Graphiurus	*crassicaudatus*	LC	1	NMNH
Graphiurus	*kelleni*	LC	8	NMNH
Graphiurus	*nagtglasii*	LC	9	NMNH, MNHN
Atherurus	*africanus*	LC	Not mentioned	Not mentioned
Arvicanthis	*rufinus*	LC	Not mentioned	MRAC
Dasymys	*rufulus*	LC	2	MNHN
Gerbilliscus	*kempi*	LC	Not mentioned	MRAC, MNHN
Grammomys	*kuru*	LC	Not mentioned	NMNH, MNHN
Hylomyscus	*pamfi*	LC	21	MNHN
Leimacomys	*buettneri*	DD	2	ZMB
Lemniscomys	*striatus*	LC	Not mentioned	NMNH, MRAC
Lophuromys	*sikapusi*	LC	Not mentioned	NMNH, MRAC
Mastomys	*erythroleucus*	LC	Not mentioned	NMNH, MRAC
Mastomys	*natalensis*	LC	2	NMNH
Mus	*minutoides*	LC	Not mentioned	MRAC
Mus	*muscoloides*	LC	44	NMNH, ZMB, MNHN, MRAC
Mus	*setulosus*	LC	3	NMNH, MNHN
Praomys	*daltoni*	LC	3	MRAC
Praomys	*derooi*	LC	Not mentioned	MRAC, NMNH, MNHN
Praomys	*misonnei*	LC	Not mentioned	Not mentioned
Praomys	*tullbergi*	LC	Not mentioned	MRAC, NMNH
Taterillus	*gracilis*	LC	Not mentioned	NMNH, MRAC
Uranomys	*ruddi*	LC	Not mentioned	NMNH, MRAC
Cricetomys	sp.	LC	Not mentioned	NMNH
Cricetomys	*gambianus*	LC	Not mentioned	NMNH, ZMB
Dendromus	*messorius*	LC	Not mentioned	Not mentioned
Funisciurus	*leucogenys*	LC	6	ZMB, NMNH
Funisciurus	*pyrropus*	LC	Not mentioned	ZMB
Funisciurus	*substriatus*	LC	Not mentioned	ZMB, NMNH
Heliosciurus	*gambianus*	LC	Not mentioned	ZMB, NMNH
Heliosciurus	*rufobrachium*	LC	Not mentioned	NMNH
Protoxerus	*stangeri*	LC	6	ZMB, NMNH
Euxerus	*erythropus*	LC	Not mentioned	ZMB, NMNH
Thryonomys	*swinderianus*	LC	12	ZMB
Crocidura	*crossei*	LC	3	NMNH
Crocidura	*grandiceps*	LC	2	NMNH
Crocidura	*lamottei*	LC	Not mentioned	Not mentioned
Crocidura	*olivieri*	LC	6	NMNH, MNHN
Suncus	*megalura*	LC	Not mentioned	Not mentioned

Appendix 2. — List of all specimens of rodent and shrew examined in three sites (**A**, Assoukoko, **D**, Diguingue and **Y**, Yegue), and tree habitat type (**Hab.)**: **GF**, gallery forest; **CF**, cultivated field; **SF**, semi-deciduous dense forest; Age: **Ad**, adult; **Sad.**, sub adult; **Juv.**, juvenile. Measurements: **TOT**, total length (HB + TAIL); **HB**, head and body length; **TAIL**, tail length; **HF**, hindfoot length; **EAR**, ear length; **Weight**, body weight; **Lat.**, latitude; **Long.**, longitude; **Inst.**, institution where the specimen is deposited **ÒUL**, université de Lomé; **FMNH**, Field Museum of Natural History); **GenBank**, GenBank accession number (sequence generated in this study); **ID**, field identification number.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK11162	*Cricetomys* sp.	Assoukoko	3/1/2023	J. C. Kerbis Peterhans, H. L. Lutz, T. C. Demos, A. Hounmavo, D. Broussounou	684	650	304	59	NA	NA	F	Ad.	GF	8.014	0.635	FMNH	–
JCK10839	*Cricetomys* sp.	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	245	164	81	37.5	19	105	F	Juv.	CF	8.177	0.656	FMNH	–
JCK10840	*Cricetomys* sp.	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	241	156	85	35	18.5	108	M	Juv.	CF	8.177	0.656	FMNH	–
JCK10841	*Cricetomys gambianus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	251	158	93	35	20	115	M	Juv.	CF	8.177	0.656	FMNH	PX147195
JCK10842	*Cricetomys* sp.	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	246	150	96	35	20	110	F	Juv.	CF	8.177	0.656	FMNH	–
JCK10843	*Cricetomys gambianus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	245	156	89	35	19	113	M	Juv.	OP	8.177	0.656	FMNH	PX147196
AH100	*Cricetomys* sp. 1	Diguingue	11/26/2022	A. Hounmavo, D. Broussounou	672	310	362	65	38	NA	M	Ad.	GF	8.072	0.647	UL	PX147194
JCK11168	*Crocidura* cf. *jouvenetae*	Assoukoko	3/4/2023	J. C. Kerbis Peterhans, H. L. Lutz, T. C. Demos, A. Hounmavo, D. Broussounou	113	67.5	45.5	10.5	8.5	54	M	Ad.	GF	8.014	0.635	FMNH	PX147212
JCK10778	*Crocidura* cf. *jouvenetae*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	121	69	52	11	10	5.9	M	Ad.	CF	8.177	0.656	FMNH	PX147213
JCK10890	*Crocidura* cf. *jouvenetae*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	120	73	47	11	8	4.65	M	Ad.	CF	8.177	0.656	FMNH	PX147214
JCK11068	*Crocidura* cf. *jouvenetae*	Yegue	2/20/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	112	66	46	10.5	9	4.1	–	Ad.	OP	8.143	0.640	FMNH	PX147216
JCK11077	*Crocidura* cf. *jouvenetae*	Yegue	2/22/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	115	69	46	10.5	8.5	5.2	M	Ad.	GF	8.146	0.646	FMNH	PX147217
JCK11211	*Crocidura* cf. *jouvenetae*	Diguingue	4/26/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	106	60	46	13	8	3.8	M	Juv.	GF	8.077	0.644	UL	PX147201
JCK11220	*Crocidura* cf. *jouvenetae*	Diguingue	4/28/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	102	59	43	9	8	3.2	–	Juv.	GF	8.077	0.644	UL	PX147202
JCK11261	*Crocidura* cf. *jouvenetae*	Diguingue	7/30/2023	A. Hounmavo, D. Broussounou	119	69	50	6	*	6	–	Ad.	CF	8.079	0.645	UL	PX147205
JCK11269	*Crocidura* cf. *jouvenetae*	Diguingue	9/6/2023	A. Hounmavo, D. Broussounou	119	72	47	12	9	6.3	F	Ad.	GF	8.077	0.644	UL	PX147206
JCK11273	*Crocidura* cf. *jouvenetae*	Diguingue	9/7/2023	A. Hounmavo, D. Broussounou	121	76	45	9	9	60	–	Ad.	GF	8.077	0.644	UL	PX147209
JCK11276	*Crocidura* cf. *jouvenetae*	Diguingue	9/9/2023	A. Hounmavo, D. Broussounou	120	70	50	11	8	5.3	F	Ad.	GF	8.077	0.644	UL	PX147380
AH cro	*Crocidura foxi*	Assoukoko	11/21/2022	A. Hounmavo, D. Broussounou	149	88	61	15	6	NA	–	Ad.	GF	8.014	0.635	UL	PX147197
JCK10837	*Crocidura foxi*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	160	107	53	14	9	16.5	F	Sad.	GF	8.145	0.647	FMNH	PX147220
JCK10852	*Crocidura foxi*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	171	109	62	15	11	21.5	M	Ad.	CF	8.177	0.656	FMNH	PX147221

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK10862	*Crocidura foxi*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	150	94	56	14	9.5	16.5	M	Ad.	GF	8.145	0.647	FMNH	PX147222
JCK10863	*Crocidura foxi*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	154	95	59	15	11	17.5	M	Ad.	GF	8.145	0.647	FMNH	PX147223
JCK11073	*Crocidura foxi*	Yegue	2/21/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	144	92	52	15	9	11.2	F	Ad.	GF	8.146	0.646	FMNH	PX147224
JCK11076	*Crocidura foxi*	Yegue	2/22/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	159	102	57	17.5	10	20.5	M	Ad.	GF	8.145	0.646	FMNH	PX147225
TCD5508	*Crocidura foxi*	Assoukoko	3/6/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	167	101	66	15	11	22	M	Ad.	GF	8.014	0.635	FMNH	PX147226
JCK11272	*Crocidura foxi*	Diguingue	9/7/2023	A. Hounmavo, D. Broussounou	152	91	61	15	NA	15	–	Ad.	GF	8.077	0.644	UL	PX147208
JCK10905	*Crocidura obscurior*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	101	61	40	11	7	4.9	M	Ad.	GF	8.145	0.647	FMNH	PX147218
JCK11067	*Crocidura obscurior*	Yegue	2/20/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	98	65.5	32.5	9.5	8.5	3.5	M	Ad.	GF	8.146	0.646	FMNH	PX147198
TCD5513	*Crocidura obscurior*	Assoukoko	3/7/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	107	69	38	9	6	5.5	M	Ad.	GF	8.014	0.635	FMNH	PX147227
TCD5514	*Crocidura obscurior*	Assoukoko	3/7/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	103	64	39	10	5	7	M	Ad.	GF	8.014	0.635	FMNH	PX147228
JCK11221	*Crocidura obscurior*	Diguingue	5/21/2023	A. Hounmavo, D. Broussounou	96	59	37	9	8	3.5	F	Juv.	GF	8.077	0.644	UL	PX147203
JCK10907	*Crocidura olivieri*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	210	141	69	21	13	42.5	F	Sad.	GF	8.145	0.647	FMNH	PX147199
JCK11049	*Crocidura olivieri*	Yegue	2/19/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	223	145	78	23.5	13.5	52	M	Ad.	OP	8.143	0.640	FMNH	PX147200
JCK11072	*Crocidura olivieri*	Yegue	2/21/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	212	140	72	22	13	49.5	M	Ad.	GF	8.145	0.646	FMNH	PX147219
JCK11241	*Crocidura olivieri*	Diguingue	6/19/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	208	128	80	20	11	39	M	Ad.	GF	8.077	0.644	UL	PX147204
JCK11245	*Crocidura olivieri*	Diguingue	6/24/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	228	140	88	21	12	49	M	Ad.	CF	8.079	0.645	UL	PX147215
JCK11271	*Crocidura olivieri*	Diguingue	9/7/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	210	124	86	21	13	50	–	Ad.	GF	8.077	0.644	UL	PX147207
JCK11278	*Crocidura olivieri*	Diguingue	9/9/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	214	129	85	21	12	51	M	Ad.	GF	8.077	0.644	UL	PX147210
JCK11280	*Crocidura olivieri*	Yegue	9/10/2023	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	210	134	76	20	12	48	–	Ad.	CF	8.177	0.656	UL	PX147211
*AH*66	*Crocidura* sp.	Assoukoko	11/21/2022	A. Hounmavo, D. Broussounou	175	115	60	16	8	19	M	Sad.	GF	8.014	0.635	UL	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK11081	*Crocidura* sp.	Yegue	2/23/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	220	131	89	19	16	46	F	Ad.	GF	8.145	0.646	FMNH	–
JCK10857	*Crocidura* sp.	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	170	116	54	14.5	11	13	M	Ad.	CF	8.177	0.656	FMNH	–
JCK10998	*Crocidura* sp.	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	120*	96*	24*	15	NA	NA	–	Ad.	GF	8.148	0.646	FMNH	–
JCK11287	*Crocidura* sp.	Diguingue	10/20/2023	A. Hounmavo, D. Broussounou	168	104	64	14	10	22	–	Ad.	GF	8.077	0.644	UL	–
JCK11056	*Funisciurus leucostigma*	Yegue	2/20/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	232*	163*	69*	40	16	105	F	Sad.	GF	8.177	0.656	FMNH	–
JCK10748	*Gerbilliscus* sp.	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	NA	NA	NA	F	Ad.	OP	8.146	0.646	FMNH	PX147234
JCK10858	*Gerbilliscus* sp.	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	300	149	151	35	23	76	M	Sad.	OP	8.177	0.656	FMNH	PX147235
JCK10870	*Gerbilliscus* sp.	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	228	96	132	30	22	42	F	Sad.	OP	8.146	0.646	FMNH	PX147236
JCK11264	*Gerbilliscus* sp.	Diguingue	8/22/2023	A. Hounmavo, D. Broussounou	266	155	111	34	16	100	F	Ad.	GF	8.077	0.644	UL	PX147237
JCK11290	*Gerbilliscus* sp.	Diguingue	10/21/2023	A. Hounmavo, D. Broussounou	315	165	150	32	17	101	M	Ad.	GF	8.077	0.644	UL	PX147233
JCK11281	*Graphiurus kelleni*	Diguingue	10/15/2023	A. Hounmavo, D. Broussounou	159	99	60	14	13	22	F	Ad.	CF	8.079	0.645	UL	PX147232
JCK11238	*Graphiurus* sp. *A*	Diguingue	6/8/2023	A. Hounmavo, D. Broussounou	133	81	52	15	12	NA	F	Ad.	GF	8.077	0.644	UL	PX147231
JCK10970	*Hylomyscus pamfi*	Yegue	2/16/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	240	101	139	18.5	14	29	M	Ad.	OP	8.143	0.640	FMNH	PX147262
JCK10981	*Hylomyscus pamfi*	Yegue	2/17/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	217	94	123	18.5	15	27	M	Ad.	OP	8.143	0.640	FMNH	PX147263
TCD5501	*Hylomyscus pamfi*	Assoukoko	3/5/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	123	60	63	15	12	5.7	M	Juv.	GF	8.014	0.635	FMNH	PX147264
JCK10976	*Lemniscomys macculus*	Yegue	2/17/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	228	111	117	25.5	16	37	M	Sad.	CF	8.177	0.656	FMNH	PX147251
JCK11059	*Lemniscomys macculus*	Yegue	2/20/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	223	113	110	24	15	35.5	F	Ad.	CF	8.177	0.656	FMNH	PX147252
JCK11172	*Lemniscomys striatus*	Assoukoko	3/6/2023	J. C. Kerbis Peterhans, H. L. Lutz, T. C. Demos, A. Hounmavo, D. Broussounou	277	123	154	28	17	56	F	Ad.	CF	8.011	0.642	FMNH	PX147249
JCK10750	*Lemniscomys striatus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	248	220	28	28	NA	NA	–	Ad.	CF	8.177	0.656	FMNH	–
JCK10752	*Lemniscomys striatus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	187	81	106	24	16	18	F	Juv.	CF	8.177	0.656	FMNH	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK10761	*Lemniscomys striatus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	223	85	138	27	16	36	F	Ad.	CF	8.177	0.656	FMNH	–
JCK10763	*Lemniscomys striatus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	NA	NA	21	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10764	*Lemniscomys striatus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	NA	NA	46	F	Ad.	CF	8.177	0.656	FMNH	–
JCK10767	*Lemniscomys striatus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	191	86	105	24	13	22	F	Juv.	CF	8.177	0.656	FMNH	–
JCK10768	*Lemniscomys striatus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	230	110	120	28	NA	34.5	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10769	*Lemniscomys striatus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	215	105	110	26	15	30.5	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10770	*Lemniscomys striatus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	272	142	130	27	NA	61	M	Ad.	CF	8.177	0.656	FMNH	–
JCK10771	*Lemniscomys striatus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	267	122	145	28	17	56	F	Ad.	CF	8.177	0.656	FMNH	–
JCK10772	*Lemniscomys striatus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	NA	NA	NA	F	Juv.	CF	8.177	0.656	FMNH	–
JCK10773	*Lemniscomys striatus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	205	96	109	25.5	16.5	30	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10832	*Lemniscomys striatus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	230	106	124	27.5	14.5	28.5	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10847	*Lemniscomys striatus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	199	97	102	25	15	26	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10848	*Lemniscomys striatus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	116	25	14	23	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10849	*Lemniscomys striatus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	106	25	16	24	M	Juv.	CF	8.177	0.656	FMNH	–
JCK10906	*Lemniscomys striatus*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	190	102	88	28	17.5	36.5	F	Ad.	GF	8.145	0.647	FMNH	–
JCK10908	*Lemniscomys striatus*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	272	137	135	27.5	18	52	F	Ad.	CF	8.177	0.656	FMNH	–
JCK10913	*Lemniscomys striatus*	Yegue	2/14/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	204	101	103	24	NA	24	F	Sad.	CF	8.177	0.656	FMNH	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK10915	*Lemniscomys striatus*	Yegue	2/14/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	185	93	92	23	18	23	M	Ad.	CF	8.177	0.656	FMNH	–
JCK10920	*Lemniscomys striatus*	Yegue	2/14/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	168	142	26	27	15	29	M	Sad.	CF	8.177	0.656	FMNH	PX147250
JCK10940	*Lemniscomys striatus*	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	NA	NA	NA	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10945	*Lemniscomys striatus*	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	188	87	101	22	14	19.5	M	Ad.	CF	8.177	0.656	FMNH	–
JCK10952	*Lemniscomys striatus*	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	258	128	130	30	15	52	M	Ad.	CF	8.177	0.656	FMNH	–
JCK10953	*Lemniscomys striatus*	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	195	97	98	24	15	20	F	Juv.	CF	8.177	0.656	FMNH	–
JCK10980	*Lemniscomys striatus*	Yegue	2/17/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	222	107	115	23.5	14	27	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10967	*Lemniscomys striatus*	Yegue	2/16/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	204	132	72	21	16	57	F	Ad.	GF	8.148	0.646	FMNH	PX147253
TCD5515	*Lemniscomys striatus*	Assoukoko	3/7/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	257	122	135	27	17	42	F	Ad.	GF	8.014	0.635	FMNH	PX147258
TCD5516	*Lemniscomys striatus*	Assoukoko	3/7/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	121	76	45	20	11	9.6	M	Juv.	GF	8.014	0.635	FMNH	PX147259
TCD5517	*Lemniscomys striatus*	Assoukoko	3/7/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	272	132	140	28	NA	60	F	Ad.	GF	8.014	0.635	FMNH	PX147260
JCK11201	*Lemniscomys striatus*	Diguingue	4/25/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	156	82	74	23	NA	14	M	Sad.	OP	8.079	0.645	UL	PX147238
JCK11208	*Lemniscomys striatus*	Diguingue	4/26/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	279	139	140	28	13	65	M	Ad.	CF	8.079	0.645	UL	–
JCK11213	*Lemniscomys striatus*	Diguingue	4/28/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	272	130	142	29	15	61	F	Sad.	GF	8.077	0.644	UL	PX147239
JCK11216	*Lemniscomys striatus*	Diguingue	4/27/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	262	126	136	28	NA	57	M	Ad.	GF	8.077	0.644	UL	PX147240
JCK11222	*Lemniscomys striatus*	Diguingue	5/21/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	285	140	145	28	18	68	M	Ad.	GF	8.077	0.644	UL	PX147254
JCK11233	*Lemniscomys striatus*	Diguingue	5/25/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	290	146	144	27	17	67	F	Ad.	CF	8.079	0.645	UL	PX147241
JCK11239	*Lemniscomys striatus*	Diguingue	6/12/2023	A. Hounmavo, D. Broussounou	273	134	139	28	16	63	M	Ad.	CF	8.079	0.645	UL	PX147242
JCK11244	*Lemniscomys striatus*	Diguingue	6/23/2023	A. Hounmavo, D. Broussounou	191	134	57	29	16	62	M	Ad.	CF	8.079	0.645	UL	PX147255
JCK11254	*Lemniscomys striatus*	Diguingue	7/29/2023	A. Hounmavo, D. Broussounou	170	80	90	13	NA	16	M	Juv.	CF	8.079	0.645	UL	PX147261
JCK11255	*Lemniscomys striatus*	Diguingue	7/29/2023	A. Hounmavo, D. Broussounou	262	132	130	NA	15	56	M	Ad.	CF	8.079	0.645	UL	PX147256
JCK11258	*Lemniscomys striatus*	Diguingue	7/30/2023	A. Hounmavo, D. Broussounou	230	128	102	27	16	65	F	Ad.	GF	8.077	0.644	UL	PX147243
JCK11260	*Lemniscomys striatus*	Diguingue	7/30/2023	A. Hounmavo, D. Broussounou	237	112	125	27	16	44	M	Ad.	CF	8.079	0.645	UL	PX147244

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK11263	*Lemniscomys striatus*	Diguingue	7/31/2023	A. Hounmavo, D. Broussounou	NA	120	NA	26	NA	51	F	Ad.	CF	8.079	0.645	UL	–
JCK11265	*Lemniscomys striatus*	Diguingue	8/24/2023	A. Hounmavo, D. Broussounou	228	107	121	25	14	42	M	Ad.	GF	8.077	0.644	UL	PX147257
JCK11266	*Lemniscomys striatus*	Diguingue	8/25/2023	A. Hounmavo, D. Broussounou	260	128	132	27	15	NA	–	Ad.	GF	8.077	0.644	UL	PX147245
JCK11267	*Lemniscomys striatus*	Diguingue	8/25/2023	A. Hounmavo, D. Broussounou	262	120	142	23	15	57	M	Ad.	GF	8.077	0.644	UL	PX147246
JCK11268	*Lemniscomys striatus*	Diguingue	8/26/2023	A. Hounmavo, D. Broussounou	161	70	91	23	13	18	–	Ad.	GF	8.077	0.644	UL	PX147247
JCK11275	*Lemniscomys striatus*	Diguingue	9/9/2023	A. Hounmavo, D. Broussounou	170	79	91	23	14	13	F	Ad.	GF	8.077	0.644	UL	PX147248
JCK11296	*Lemniscomys striatus*	Diguingue	10/23/2023	A. Hounmavo, D. Broussounou	180	97	83	21	13	27	F	Ad.	CF	8.079	0.645	UL	–
JCK11157	*Lophuromys sikapusi*	Yegue	2/28/2023	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	25	14	NA	M	Ad.	GF	8.145	0.647	FMNH	PX147190
JCK11158	*Lophuromys sikapusi*	Yegue	2/28/2023	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	231	148	83	25	16	NA	F	Ad.	GF	8.145	0.647	FMNH	PX147191
JCK10986	*Lophuromys sikapusi*	Yegue	2/17/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	215	140	75	23	17	73	M	Ad.	GF	8.177	0.656	FMNH	PX147192
JCK11075	*Lophuromys sikapusi*	Yegue	2/21/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	204	136	68	24	17	68.5	M	Ad.	GF	8.145	0.646	FMNH	PX147193
JCK11079	*Lophuromys sikapusi*	Yegue	2/23/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	158	129	29	23	18	79	M	Ad.	GF	8.145	0.646	FMNH	–
JCK11283	*Lophuromys sikapusi*	Diguingue	10/19/2023	A. Hounmavo, D. Broussounou	196	121	75	22	16	53	F	Sad.	GF	8.077	0.644	UL	PX147189
JCK11288	*Lophuromys sikapusi*	Diguingue	10/20/2023	A. Hounmavo, D. Broussounou	225	145	80	22	19	78	F	Sad.	GF	8.077	0.644	UL	–
JCK10747	*Mastomys erythroleucus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	164	86	78	22	18	19	M	Juv.	OP	8.146	0.646	FMNH	–
JCK10815	*Mastomys erythroleucus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	102	25	NA	23	F	Sad.	OP	8.146	0.646	FMNH	–
JCK10975	*Mastomys erythroleucus*	Yegue	2/17/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	199	102	97	24	16	30	M	Sad.	OP	8.146	0.645	FMNH	–
JCK10746	*Mastomys erythroleucus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	261	143	118	24	19	74	F	Ad.	CF	8.177	0.656	FMNH	–
JCK10749	*Mastomys erythroleucus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	177	137	80	21	16	20	M	Juv.	OP	8.146	0.646	FMNH	–
JCK10751	*Mastomys erythroleucus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	224	127	105	25	18	39	M	Sad.	OP	8.146	0.646	FMNH	–
JCK10753	*Mastomys erythroleucus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	149	135	67	21	NA	19	M	Juv.	OP	8.146	0.646	FMNH	–
JCK10754	*Mastomys erythroleucus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	207	125	111	23	16	37	F	Sad.	OP	8.146	0.646	FMNH	–
JCK10755	*Mastomys erythroleucus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	147	77	21	NA	NA	F	Juv.	OP	8.146	0.646	FMNH	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK10756	*Mastomys erythroleucus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	164	99	81	24	15	15	–	Juv.	CF	8.177	0.656	FMNH	–
JCK10757	*Mastomys erythroleucus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	116	124	96	22	13	28	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10758	*Mastomys erythroleucus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	126	21	105	24	17	31	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10759	*Mastomys erythroleucus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, Mawugnigan Valentin, R. Issaka, D. Broussounou	211	105	106	22	16	40	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10760	*Mastomys erythroleucus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	186	93	93	23	17	26	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10762	*Mastomys erythroleucus*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	NA	NA	NA	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10783	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	124	70	54	19.5	13	7.1	F	Juv.	CF	8.177	0.656	FMNH	–
JCK10787	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	171	91	80	21	16	21	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10788	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	200	101	99	21	17	32	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10789	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	203	96	107	24	18	29	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10792	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	199	95	104	23	17	33	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10793	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	155	75	80	20	29	17	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10794	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	176	87	89	22	16	21	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10795	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	265	137	128	22	20	61	F	Ad.	CF	8.177	0.656	FMNH	PX147369
JCK10796	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	218	103	115	24	17	28	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10797	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	225	127	98	22	18	NA	F	Ad.	CF	8.177	0.656	FMNH	–
JCK10798	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	205	100	105	23	18	33	M	Sad.	CF	8.177	0.656	FMNH	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK10799	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	268	135	133	24	21	96	M	Ad.	CF	8.177	0.656	FMNH	PX147370
JCK10800	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	174	87	87	20	17	22	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10801	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	200	102	98	23	15	30	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10807	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	191	103	88	22	17	26	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10808	*Mastomys erythroleucus*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	202	106	96	22.5	19	25	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10816	*Mastomys erythroleucus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	100	29	71	21	15	17.5	F	Juv.	CF	8.177	0.656	FMNH	–
JCK10817	*Mastomys erythroleucus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	182	90	92	12	14	27.5	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10818	*Mastomys erythroleucus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	131	57	74	21	15	21	F	Juv.	CF	8.177	0.656	FMNH	–
JCK10819	*Mastomys erythroleucus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	223	114	109	22	18	38	M	Juv.	CF	8.177	0.656	FMNH	–
JCK10820	*Mastomys erythroleucus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	103	20	16	26	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10821	*Mastomys erythroleucus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	204	101	103	23	17	34	F	Juv.	CF	8.177	0.656	FMNH	–
JCK10822	*Mastomys erythroleucus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	200	100	100	23	17	31	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10823	*Mastomys erythroleucus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	193	100	93	22	15	31.5	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10826	*Mastomys erythroleucus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	213	100	113	25	17	32	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10827	*Mastomys erythroleucus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	183	80	103	28	17	29	F	Juv.	CF	8.177	0.656	FMNH	–
JCK10828	*Mastomys erythroleucus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	188	86	102	22	16	30	F	Juv.	CF	8.177	0.656	FMNH	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK10829	*Mastomys erythroleucus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	205	105	100	22	15	40	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10830	*Mastomys erythroleucus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	210	99	111	21	16	37	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10833	*Mastomys erythroleucus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	190	100	90	19	NA	29	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10856	*Mastomys erythroleucus*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	125	69	56	19	13	7.4	M	neo-nate	CF	8.177	0.656	FMNH	–
JCK10861	*Mastomys erythroleucus*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	NA	NA	NA	M	Sad.	CF	8.177	0.656	FMNH	PX147371
JCK10864	*Mastomys erythroleucus*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	192	97	95	23	19	33	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10865	*Mastomys erythroleucus*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	214	104	110	24	NA	32	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10866	*Mastomys erythroleucus*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	165	81	84	21	19	20	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10867	*Mastomys erythroleucus*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	177	87	90	21	17	22	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10868	*Mastomys erythroleucus*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	212	110	102	20	16	47	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10869	*Mastomys erythroleucus*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	250	125	125	24	17	65	F	Ad.	CF	8.177	0.656	FMNH	PX147372
JCK10873	*Mastomys erythroleucus*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	154	76	78	19	17	16	M	Juv.	CF	8.177	0.656	FMNH	–
JCK10874	*Mastomys erythroleucus*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	200	98	102	21	17	28	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10875	*Mastomys erythroleucus*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	NA	NA	NA	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10876	*Mastomys erythroleucus*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	190	100	90	22	19	29	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10886	*Mastomys erythroleucus*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	270	147	123	26	18.5	84	M	Ad.	CF	8.177	0.656	FMNH	PX147373
JCK10895	*Mastomys erythroleucus*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	162	79	83	20	12	18	F	Sad.	CF	8.177	0.656	FMNH	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK10896	*Mastomys erythroleucus*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	162	87	75	21	17	16	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10897	*Mastomys erythroleucus*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	182	88	94	22	12	28	–	Sad.	CF	8.177	0.656	FMNH	–
JCK10898	*Mastomys erythroleucus*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	197	92	105	22	21	32	F	Sad.	CF	8.177	0.656	FMNH	–
JCK10899	*Mastomys erythroleucus*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	209	100	109	23	18	38	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10901	*Mastomys erythroleucus*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	163	76	87	22	16	29	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10902	*Mastomys erythroleucus*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	185	87	98	22	18	28	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10903	*Mastomys erythroleucus*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	190	90	100	22	16	29	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10904	*Mastomys erythroleucus*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	90	12	NA	NA	–	–	CF	8.177	0.656	FMNH	–
JCK10911	*Mastomys erythroleucus*	Yegue	2/14/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	194	103	91	23.5	18	27	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10912	*Mastomys erythroleucus*	Yegue	2/14/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	210	115	95	24	18	35	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10916	*Mastomys erythroleucus*	Yegue	2/14/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	185	93	92	23	18	23	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10943	*Mastomys erythroleucus*	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	186	94	92	24	21	24.5	M	–	CF	8.177	0.656	FMNH	–
JCK10949	*Mastomys erythroleucus*	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	192	99	93	15	18	27	M	Ad.	CF	8.177	0.656	FMNH	–
JCK10977	*Mastomys erythroleucus*	Yegue	2/17/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	198	105	93	22.5	NA	26	M	Sad.	CF	8.177	0.656	FMNH	–
JCK10979	*Mastomys erythroleucus*	Yegue	2/17/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	262	149	113	23.5	NA	70	F	Sad.	CF	8.177	0.656	FMNH	PX147374
JCK11000	*Mastomys erythroleucus*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	211	101	110	24	NA	30	–	–	CF	8.177	0.656	FMNH	–
JCK11006	*Mastomys erythroleucus*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	202	89	113	22	NA	33	M	Juv.	CF	8.177	0.656	FMNH	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK11007	*Mastomys erythroleucus*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	177	85	92	23	NA	25	M	Juv.	CF	8.177	0.656	FMNH	–
JCK11008	*Mastomys erythroleucus*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	175	88	87	21	NA	22	M	Juv.	CF	8.177	0.656	FMNH	–
JCK11009	*Mastomys erythroleucus*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	190	90	100	22	NA	26	F	Sad.	CF	8.177	0.656	FMNH	–
JCK11010	*Mastomys erythroleucus*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	175	93	82	21	NA	26	M	Juv.	CF	8.177	0.656	FMNH	–
JCK11012	*Mastomys erythroleucus*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	250	135	115	23	NA	64	F	Sad.	CF	8.177	0.656	FMNH	–
JCK11013	*Mastomys erythroleucus*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	190	100	90	23	NA	26	M	Juv.	CF	8.177	0.656	FMNH	–
JCK11014	*Mastomys erythroleucus*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	201	99	102	24	17	35	M	Juv.	CF	8.177	0.656	FMNH	–
JCK11016	*Mastomys erythroleucus*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	NA	NA	NA	–	–	CF	8.177	0.656	FMNH	–
JCK11041	*Mastomys erythroleucus*	Yegue	2/19/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	213	98	115	24	18	27	M	Sad.	CF	8.177	0.656	FMNH	–
JCK11044	*Mastomys erythroleucus*	Yegue	2/19/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	190	92	98	22	18	23	F	Juv.	CF	8.177	0.656	FMNH	–
JCK11046	*Mastomys erythroleucus*	Yegue	2/19/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	183	88	95	23	NA	29	F	Sad.	CF	8.177	0.656	FMNH	–
JCK11047	*Mastomys erythroleucus*	Yegue	2/19/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	195	95	100	23	18	29	F	Sad.	CF	8.177	0.656	FMNH	–
JCK11048	*Mastomys erythroleucus*	Yegue	2/19/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	175	88	87	22	17	22	F	Sad.	CF	8.177	0.656	FMNH	–
JCK11063	*Mastomys erythroleucus*	Yegue	2/20/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	190	90	100	23	18	24	M	Juv.	CF	8.177	0.656	FMNH	–
JCK11065	*Mastomys erythroleucus*	Yegue	2/20/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	192	97	95	24	NA	23	M	Sad.	CF	8.177	0.656	FMNH	–
JCK11066	*Mastomys erythroleucus*	Yegue	2/20/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	195	97	98	21	18	24	M	Juv.	CF	8.177	0.656	FMNH	–
JCK11071	*Mastomys erythroleucus*	Yegue	2/21/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	203	112	91	25	17	25.5	F	Juv.	CF	8.177	0.656	FMNH	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK11074	*Mastomys erythroleucus*	Yegue	2/21/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	198*	101	97*	NA	NA	23.5	F	Juv.	CF	8.177	0.656	FMNH	–
JCK11082	*Mastomys erythroleucus*	Yegue	2/23/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	161	79	82	22	17	17			CF	8.177	0.656	FMNH	–
JCK11058	*Mastomys erythroleucus*	Yegue	2/20/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	116*	92	24*	NA	NA	19	M	Juv.	CF	8.177	0.656	FMNH	–
JCK11061	*Mastomys erythroleucus*	Yegue	2/20/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	125	105	20	20	16	6.2	–	Juv.	CF	8.177	0.656	FMNH	–
TCD5498	*Mastomys erythroleucus*	Assoukoko	3/5/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	204	105	99	23	18	31	F	Sad.	GF	8.014	0.635	FMNH	PX147379
JCK11200	*Mastomys erythroleucus*	Diguingue	4/25/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	240	120	120	24	NA	49	F	Ad.	OP	–	–	UL	PX147360
JCK11203	*Mastomys erythroleucus*	Diguingue	4/25/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	220	111	109	23	15	38	F	Ad.	GF	8.077	0.644	UL	PX147361
JCK11204	*Mastomys erythroleucus*	Diguingue	4/25/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	207	108	99	NA	NA	33	F	Ad.	GF	8.077	0.644	UL	–
JCK11207	*Mastomys erythroleucus*	Diguingue	4/26/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	250	130	120	24	15	59	M	Ad.	CF	8.079	0.645	UL	PX147375
JCK11217	*Mastomys erythroleucus*	Diguingue	4/27/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	223	112	111	21	21	37	F	Ad.	GF	8.077	0.644	UL	PX147362
JCK11218	*Mastomys erythroleucus*	Diguingue	4/27/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	200	104	96	22	17	30	F	Ad.	OP	–	–	UL	PX147377
JCK11219	*Mastomys erythroleucus*	Diguingue	4/27/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	224	115	109	27	NA	35	F	Ad.	OP	–	–	UL	PX147363
JCK11242	*Mastomys erythroleucus*	Diguingue	6/20/2023	A. Hounmavo, D. Broussounou	250	133	117	25	12	59	M	Ad.	CF	8.079	0.645	UL	PX147365
JCK11243	*Mastomys erythroleucus*	Diguingue	6/20/2023	A. Hounmavo, D. Broussounou	270	135	135	25	19	66	M	Ad.	CF	8.079	0.645	UL	PX147378
JCK11256	*Mastomys erythroleucus*	Diguingue	7/29/2023	A. Hounmavo, D. Broussounou	289	142	117	NA	*	67	M	Ad.	CF	8.079	0.645	UL	PX147366
JCK11257	*Mastomys erythroleucus*	Diguingue	7/29/2023	A. Hounmavo, D. Broussounou	274	121	153	27	16	63	M	Ad.	CF	8.079	0.645	UL	PX147367
JCK11270	*Mastomys erythroleucus*	Diguingue	9/6/2023	A. Hounmavo, D. Broussounou	260	133	127	24	17	94	M	Ad.	GF	8.077	0.644	UL	PX147368
*AH*76	*Mastomys natalensis*	Diguingue	11/24/2022	A. Hounmavo, D. Broussounou	156	77	79	22	12	NA	F	Sad.	CF	8.079	0.645	UL	PX147359
JCK11055	*Mastomys natalensis*	Yegue	2/20/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	238	124	114	25	18	55	M	Ad.	CF	8.177	0.656	FMNH	PX147376
JCK11240	*Mastomys natalensis*	Diguingue	6/14/2023	A. Hounmavo, D. Broussounou	229	119	110	22	16	56	M	Ad.	CF	8.079	0.645	UL	PX147364
JCK10766	*Mus (Nannomys)* sp.	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	97	66	31	12.5	9	7.2	M	Ad.	OP	8.146	0.646	FMNH	–
JCK10781	*Mus (Nannomys)* sp.	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	94	51	43	12	9	3.8	F	Juv.	OP	8.146	0.646	FMNH	–
JCK10782	*Mus (Nannomys)* sp.	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	97	52	45	12	9	4.3	F	Sad.	OP	8.146	0.646	FMNH	–
JCK10786	*Mus (Nannomys)* sp.	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	92	52	40	12	7	3.5	F	Sad.	OP	8.146	0.646	FMNH	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK10790	*Mus (Nannomys)* sp.	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	97	52	45	12	9	4.4	F	Sad.	OP	8.146	0.646	FMNH	–
JCK10791	*Mus (Nannomys)* sp.	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	41	11	NA	NA	–	–	CF	8.177	0.656	FMNH	–
JCK10824	*Mus (Nannomys)* sp.	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	99	53	46	6	9	4	M	Sad.	OP	8.146	0.646	FMNH	–
JCK10825	*Mus (Nannomys)* sp.	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	105	66	39	7	NA	7	F	Ad.	OP	8.146	0.646	FMNH	–
JCK10854	*Mus (Nannomys)* sp.	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	104	58	46	13.5	9	4.3	F	Ad.	OP	8.146	0.646	FMNH	–
JCK10855	*Mus (Nannomys)* sp.	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	79	44	35	11.5	7	2	M	Juv.	OP	8.146	0.646	FMNH	–
JCK10888	*Mus (Nannomys)* sp.	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	103	56	47	12	8.5	4.1	F	Sad.	OP	8.146	0.646	FMNH	–
JCK10889	*Mus (Nannomys)* sp.	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	45	12.5	NA	NA	F	Ad.	OP	8.146	0.646	FMNH	–
JCK10893	*Mus (Nannomys)* sp.	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	98	61	37	11	9	5	M	Ad.	OP	8.146	0.646	FMNH	–
JCK10894	*Mus (Nannomys)* sp.	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	90	55	35	11.5	9.5	3.5	M	Sad.	OP	8.146	0.646	FMNH	–
JCK10900	*Mus (Nannomys)* sp.	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	86	46	40	NA	12	3.6	F	Sad.	OP	8.146	0.646	FMNH	–
JCK10910	*Mus (Nannomys)* sp.	Yegue	2/14/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	93	50	43	12.5	8	3.5	F	Sad.	OP	8.146	0.646	FMNH	–
JCK10917	*Mus (Nannomys)* sp.	Yegue	2/14/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	100	58	42	9	8	6	M	–	OP	8.145	0.647	FMNH	–
JCK10939	*Mus (Nannomys)* sp.	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	95	50	45	11	NA	4.1	F	Sad.	OP	8.145	0.647	FMNH	–
JCK10978	*Mus (Nannomys)* sp.	Yegue	2/17/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	98	58	40	9	9	3.8	F	Sad.	OP	8.146	0.645	FMNH	–
JCK10999	*Mus (Nannomys)* sp.	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	NA	9	4	F	Ad.	OP	8.148	0.646	FMNH	–
JCK11001	*Mus (Nannomys)* sp.	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	116	64	52	NA	9	6.9	F	Ad.	OP	8.148	0.646	FMNH	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK11042	*Mus (Nannomys)* sp.	Yegue	2/19/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	NA	NA	NA	–	–	OP	8.148	0.646	FMNH	–
JCK11050	*Mus (Nannomys)* sp.	Yegue	2/19/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	90	47	43	11	NA	5.2	F	Sad.	CF	8.177	0.656	FMNH	–
JCK11051	*Mus (Nannomys)* sp.	Yegue	2/19/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	107	59	48	11	10	7	F	Sad.	CF	8.177	0.656	FMNH	–
JCK11212	*Mus (Nannomys)* sp.	Diguingue	4/26/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	103	62	41	13	9	7.1	M	Ad.	GF	8.077	0.644	UL	–
JCK11214	*Mus (Nannomys)* sp.	Diguingue	4/27/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	109	59	50	12	NA	5.6	M	Sad.	GF	8.077	0.644	UL	–
JCK10780	*Mus baoulei*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	100	62	38	13	11	6.5	M	Sad.	OP	8.146	0.646	FMNH	PX147273
JCK10872	*Mus baoulei*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	72	55	17	11	NA	4	F	Ad.	OP	8.146	0.646	FMNH	PX147277
JCK10892	*Mus baoulei*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	104	65	39	13	10	5.3	–	–	OP	8.146	0.646	FMNH	PX147279
TCD5511	*Mus baoulei*	Assoukoko	3/7/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	109	70	39	13	12	6.4	F	Ad.	GF	8.014	0.635	FMNH	PX147286
JCK11202	*Mus baoulei*	Diguingue	4/25/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	93	55	38	12	10	5.4	F	Juv.	OP	–	–	UL	PX147265
JCK11206	*Mus baoulei*	Diguingue	4/25/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	105	69	36	12	NA	6.8	M	Sad.	OP	–	–	UL	PX147267
JCK11253	*Mus baoulei*	Diguingue	7/29/2023	A. Hounmavo, D. Broussounou	*	*	*	8	*	5.7	F	Sad.	CF	8.079	0.645	UL	PX147270
JCK11167	*Mus minutoides*	Yegue	3/4/2023	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	42	11.5	7	NA	M	Ad.	CF	8.177	0.656	FMNH	PX147271
JCK11178	*Mus minutoides*	Assoukoko	3/7/2023	J. C. Kerbis Peterhans, H. L. Lutz, T. C. Demos, A. Hounmavo, D. Broussounou	108	58	50	12	9	5.4	M	Ad.	GF	8.014	0.635	FMNH	PX147272
JCK10784	*Mus minutoides*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	108	65	43	12	9.5	5.6	F	Ad.	OP	8.146	0.646	FMNH	PX147274
JCK10785	*Mus minutoides*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	110	64	46	12.5	8	6	M	Sad.	OP	8.146	0.646	FMNH	PX147275
JCK10860	*Mus minutoides*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	NA	NA	NA	M	Ad.	OP	8.145	0.647	FMNH	PX147276
JCK10891	*Mus minutoides*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	107	62	45	12	10	5.7	M	Ad.	OP	8.146	0.646	FMNH	PX147278
JCK11017	*Mus minutoides*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	108	62	46	12	9	5.9	F	Ad.	CF	8.177	0.656	FMNH	PX147280

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK11070	*Mus minutoides*	Yegue	2/20/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	NA	NA	4.4	M	Ad.	CF	8.177	0.656	FMNH	PX147281
TCD5475	*Mus minutoides*	Assoukoko	3/1/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	103	60	43	13	9	5.4	M	Ad.	GF	8.014	0.635	FMNH	PX147282
TCD5476	*Mus minutoides*	Assoukoko	3/1/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	109	61	48	11	9	5.6	M	Ad.	GF	8.014	0.635	FMNH	PX147283
TCD5497	*Mus minutoides*	Assoukoko	3/5/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	100	56	44	11	NA	3.5	–	Juv.	GF	8.014	0.635	FMNH	PX147284
TCD5504	*Mus minutoides*	Assoukoko	3/6/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	71	38	33	10	5	1.5	F	Juv.	CF	8.011	0.642	FMNH	PX147285
TCD5518	*Mus minutoides*	Assoukoko	3/7/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	120	68	52	11	10	8.5	F	Ad.	GF	8.014	0.635	FMNH	PX147287
JCK11205	*Mus minutoides*	Diguingue	4/25/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	105	65	40	12	9	6.5	M	Ad.	OP	–	–	UL	PX147266
JCK11209	*Mus minutoides*	Diguingue	4/26/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	108	57	51	12	NA	5.6	F	Ad.	GF	8.077	0.644	UL	PX147268
JCK11210	*Mus minutoides*	Diguingue	4/26/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	116	63	53	13	9	7.1	F	Ad.	OP	–	–	UL	PX147269
*AH*99	*Praomys daltoni*	Diguingue	11/24/2022	A. Hounmavo, D. Broussounou	190	90	100	20	11	21	M	Ad.	CF	8.079	0.645	UL	–
*AH*92	*Praomys daltoni*	Diguingue	11/24/2022	A. Hounmavo, D. Broussounou	179	82	97	19	14	14	M	Juv.	CF	8.079	0.645	UL	–
*AH*88	*Praomys daltoni*	Diguingue	11/26/2022	A. Hounmavo, D. Broussounou	190	95	95	18	16	26	M	Ad.	CF	8.079	0.645	UL	–
*AH*103	*Praomys daltoni*	Diguingue	11/26/2022	A. Hounmavo, D. Broussounou	228	115	113	20	14	37	M	Ad.	CF	8.079	0.645	UL	–
*AH*86	*Praomys daltoni*	Diguingue	11/24/2022	A. Hounmavo, D. Broussounou	165	75	90	19	13	12	M	Juv.	CF	8.079	0.645	UL	PX147289
*AH*70	*Praomys daltoni*	Diguingue	11/24/2022	A. Hounmavo, D. Broussounou	202	92	110	20	15	24	M	Sad.	CF	8.079	0.645	UL	PX147290
*AH*82	*Praomys daltoni*	Diguingue	11/24/2022	A. Hounmavo, D. Broussounou	170	80	90	19	14	13	F	Juv.	CF	8.079	0.645	UL	–
*AH*73	*Praomys daltoni*	Diguingue	11/24/2022	A. Hounmavo, D. Broussounou	205	95	110	21	15	29	F	Sad.	CF	8.079	0.645	UL	PX147291
*AH*80	*Praomys daltoni*	Diguingue	11/23/2022	A. Hounmavo, D. Broussounou	170	80	90	19	14	13	M	Juv.	CF	8.079	0.645	UL	PX147292
*AH*105	*Praomys daltoni*	Diguingue	11/26/2022	A. Hounmavo, D. Broussounou	210	101	109	20	15	24	M	Ad.	CF	8.079	0.645	UL	PX147293
*AH*N9	*Praomys daltoni*	Assoukoko	11/25/2022	A. Hounmavo, D. Broussounou	230	111	119	26	16	NA	M	Ad.	GF	8.014	0.635	UL	PX147294
*AH*N7	*Praomys daltoni*	Diguingue	11/24/2022	A. Hounmavo, D. Broussounou	199	95	104	21	15	NA	F	Ad.	CF	8.079	0.645	UL	PX147295
*AH*94	*Praomys daltoni*	Diguingue	11/24/2022	A. Hounmavo, D. Broussounou	183	89	94	19	12	30	M	Ad.	CF	8.079	0.645	UL	PX147296
*AH*67	*Praomys daltoni*	Assoukoko	11/21/2022	A. Hounmavo, D. Broussounou	210	96	114	19	15	36	F	Ad.	CF	8.011	0.642	UL	PX147297
*AH*81	*Praomys daltoni*	Diguingue	11/24/2022	A. Hounmavo, D. Broussounou	194	99	95	19	14	28	M	Ad.	CF	8.079	0.645	UL	PX147299
*AH*N6	*Praomys daltoni*	Assoukoko	11/24/2022	A. Hounmavo, D. Broussounou	170	85	85	17	13	NA	F	Ad.	CF	8.011	0.642	UL	PX147300
*AH*63	*Praomys daltoni*	Assoukoko	11/25/2022	A. Hounmavo, D. Broussounou	182	83	99	19	16	23	M	Ad.	CF	8.011	0.642	UL	PX147301
*AH*N8	*Praomys daltoni*	Assoukoko	11/23/2022	A. Hounmavo, D. Broussounou	198	94	104	21	14	NA	M	Ad.	GF	8.014	0.635	UL	PX147303
JCK11166	*Praomys daltoni*	Assoukoko	3/4/2023	J. C. Kerbis Peterhans, H. L. Lutz, T. C. Demos, A. Hounmavo, D. Broussounou	191	97	94	19	13.5	19	M	Ad.	CF	8.011	0.642	FMNH	PX147320
JCK10713	*Praomys daltoni*	Yegue	2/7/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	229	103	126	21.5	15	35	F	Ad.	GF	7.521	1.139	FMNH	PX147330
JCK11284	*Praomys daltoni*	Diguingue	10/19/2023	A. Hounmavo, D. Broussounou	120	64	56	12	NA	9	M	Ad.	GF	8.077	0.644	UL	–
JCK11286	*Praomys daltoni*	Diguingue	10/20/2023	A. Hounmavo, D. Broussounou	175	84	1	23	16	19	F	Sad.	GF	8.077	0.644	UL	–
JCK11292	*Praomys daltoni*	Diguingue	10/22/2023	A. Hounmavo, D. Broussounou	230	107	123	21	NA	13	M	Ad.	CF	8.079	0.645	UL	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK11293	*Praomys daltoni*	Diguingue	10/22/2023	A. Hounmavo, D. Broussounou	233	123	110	25	16	37	M	Ad.	CF	8.079	0.645	UL	–
JCK11295	*Praomys daltoni*	Diguingue	10/23/2023	A. Hounmavo, D. Broussounou	220	117	103	21	NA	43	M	Ad.	GF	8.077	0.644	UL	–
JCK11298	*Praomys daltoni*	Diguingue	10/23/2023	A. Hounmavo, D. Broussounou	285	120	165	24	17	48	M	Ad.	CF	8.079	0.645	UL	–
*AH*101	*Praomys misonnei*	Diguingue	11/26/2022	A. Hounmavo, D. Broussounou	240	114	126	23	16	42	M	Ad.	GF	8.072	0.647	UL	PX147318
*AH*97	*Praomys misonnei*	Diguingue	11/24/2022	A. Hounmavo, D. Broussounou	120	49	71	15	10	6	M	Juv.	CF	8.079	0.645	UL	–
*AH*87	*Praomys misonnei*	Diguingue	11/24/2022	A. Hounmavo, D. Broussounou	200	90	110	18	15	20	F	Juv.	CF	8.079	0.645	UL	–
*AH*78	*Praomys misonnei*	Diguingue	11/23/2022	A. Hounmavo, D. Broussounou	240	110	130	20	15	40	F	Sad.	CF	8.079	0.645	UL	–
*AH*64	*Praomys misonnei*	Assoukoko	11/21/2022	A. Hounmavo, D. Broussounou	241	102	139	22	9	30.4	M	Sad.	GF	8.014	0.635	UL	PX147288
*AH*N1	*Praomys misonnei*	Diguingue	11/24/2022	A. Hounmavo, D. Broussounou	183	83	100	20	15	NA	F	Ad.	CF	8.079	0.645	UL	PX147319
*AH*83	*Praomys misonnei*	Assoukoko	11/26/2022	A. Hounmavo, D. Broussounou	224	94	130	21.5	16	NA	F	Ad.	GF	8.014	0.635	UL	PX147298
*AH*98	*Praomys misonnei*	Diguingue	11/24/2022	A. Hounmavo, D. Broussounou	233	101	132	22	NA	NA	F	Ad.	GF	8.072	0.647	UL	PX147302
JCK11160	*Praomys misonnei*	Assoukoko	3/1/2023	J. C. Kerbis Peterhans, H. L. Lutz, T. C. Demos, A. Hounmavo, D. Broussounou	176	80	96	21.5	14.5	13.5	M	Juv.	GF	8.014	0.635	FMNH	PX147304
JCK11161	*Praomys misonnei*	Assoukoko	3/1/2023	J. C. Kerbis Peterhans, H. L. Lutz, T. C. Demos, A. Hounmavo, D. Broussounou	205	47	158	24.5	18	38.5	F	Ad.	GF	8.014	0.635	FMNH	–
JCK10942	*Praomys misonnei*	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	261	117	144	20	19	39	F	Ad.	OP	8.148	0.646	FMNH	–
JCK10834	*Praomys misonnei*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	250	114	136	22	12	27	F	Ad.	OP	8.145	0.647	FMNH	–
JCK10835	*Praomys misonnei*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	205	87	118	17	14	25	F	Ad.	OP	8.145	0.647	FMNH	–
JCK10836	*Praomys misonnei*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	246	105	141	23	17	32.5	F	Ad.	OP	8.145	0.647	FMNH	–
JCK10859	*Praomys misonnei*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	NA	NA	NA	F	Ad.	OP	8.145	0.647	FMNH	–
JCK10871	*Praomys misonnei*	Yegue	2/12/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	245	106	139	22	13	40	M	Ad.	OP	8.146	0.646	FMNH	–
JCK10887	*Praomys misonnei*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	276	124	152	24	18	45	F	Ad.	OP	8.146	0.646	FMNH	–
JCK10918	*Praomys misonnei*	Yegue	2/14/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	269	126	143	23	18	43	F	Ad.	OP	8.145	0.647	FMNH	–
JCK10938	*Praomys misonnei*	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	266	120	146	24	17.5	45	M	Ad.	OP	8.146	0.646	FMNH	–
JCK10941	*Praomys misonnei*	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	101	20	17	NA	F	Sad.	OP	8.148	0.646	FMNH	–
JCK10944	*Praomys misonnei*	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	240	103	137	20	16	28	M	–	OP	8.148	0.646	FMNH	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK10946	*Praomys misonnei*	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	205	103	102	20	16	22	F	Sad.	OP	8.148	0.646	FMNH	–
JCK10947	*Praomys misonnei*	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	297	132	165	20	16	55	M	–	OP	8.148	0.646	FMNH	–
JCK10948	*Praomys misonnei*	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	258	116	142	17	19	44	F	Ad.	OP	8.148	0.646	FMNH	–
JCK10950	*Praomys misonnei*	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	245	101	144	18	NA	31	F	Sad.	OP	8.148	0.646	FMNH	–
JCK10951	*Praomys misonnei*	Yegue	2/15/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	194	87	107	17	16	17	F	Sad.	OP	8.148	0.646	FMNH	–
JCK10959	*Praomys misonnei*	Yegue	2/16/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	249	132	117	24	18	50	M	Ad.	OP	8.146	0.646	FMNH	–
JCK10960	*Praomys misonnei*	Yegue	2/16/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	267	120	147	22	18	44	M	Ad.	OP	8.148	0.646	FMNH	–
JCK10961	*Praomys misonnei*	Yegue	2/16/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	156	71	85	19	15	10	M	Juv.	OP	8.148	0.646	FMNH	–
JCK10962	*Praomys misonnei*	Yegue	2/16/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	150	65	85	19	16	10	F	Juv.	OP	8.148	0.646	FMNH	–
JCK10963	*Praomys misonnei*	Yegue	2/16/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	142	61	81	19	13	10	F	Juv.	OP	8.148	0.646	FMNH	–
JCK10964	*Praomys misonnei*	Yegue	2/16/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	164	59	105	22	12	18	F	Juv.	OP	8.148	0.646	FMNH	–
JCK10965	*Praomys misonnei*	Yegue	2/16/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	266	118	148	24	17	46	F	Ad.	OP	8.148	0.646	FMNH	–
JCK10966	*Praomys misonnei*	Yegue	2/16/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	280	123	157	24	17	48	M	Ad.	OP	8.148	0.646	FMNH	–
JCK10968	*Praomys misonnei*	Yegue	2/16/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	192	80	112	20	17	18	F	Juv.	OP	8.148	0.646	FMNH	–
JCK10969	*Praomys misonnei*	Yegue	2/16/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	270	110	160	24	17	49.5	F	Ad.	OP	8.145	0.647	FMNH	–
JCK10982	*Praomys misonnei*	Yegue	2/17/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	265	115	150	23	15	47	M	Ad.	OP	8.148	0.646	FMNH	–
JCK10983	*Praomys misonnei*	Yegue	2/17/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	202	92	110	21	16	20	F	Juv.	OP	8.148	0.646	FMNH	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK10984	*Praomys misonnei*	Yegue	2/17/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	229	99	130	22	NA	23	F	Sad.	OP	8.148	0.646	FMNH	–
JCK10985	*Praomys misonnei*	Yegue	2/17/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	250	110	140	23	12	38	F	Ad.	OP	8.148	0.646	FMNH	–
JCK11002	*Praomys misonnei*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	202	92	110	22	18	20	F	Juv.	OP	8.148	0.646	FMNH	–
JCK11003	*Praomys misonnei*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	227	102	125	25	18	29.5	F	Sad.	OP	8.148	0.646	FMNH	–
JCK11004	*Praomys misonnei*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	254	112	142	23	18	30.6	F	Ad.	OP	8.148	0.646	FMNH	–
JCK11005	*Praomys misonnei*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	259	122	137	22	NA	40.4	M	Ad.	OP	8.148	0.646	FMNH	–
JCK11011	*Praomys misonnei*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	263	113	150	22	17	31	F	Sad.	OP	8.146	0.645	FMNH	–
JCK11015	*Praomys misonnei*	Yegue	2/18/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	202	87	115	21	18	17	F	Juv.	OP	8.146	0.645	FMNH	–
JCK11037	*Praomys misonnei*	Yegue	2/19/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	205	91	114	22	17	NA	F	Sad.	OP	8.145	0.646	FMNH	–
JCK11043	*Praomys misonnei*	Yegue	2/19/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	232	106	126	22	17	29	F	Sad.	OP	8.148	0.646	FMNH	–
JCK11045	*Praomys misonnei*	Yegue	2/19/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	199	89	110	23	18	17	M	Juv.	OP	8.146	0.645	FMNH	–
JCK11060	*Praomys misonnei*	Yegue	2/20/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	210	100	110	NA	NA	36	F	Ad.	OP	8.148	0.646	FMNH	–
JCK11062	*Praomys misonnei*	Yegue	2/20/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	213	91	122	22	17	26	F	Ad.	OP	8.148	0.646	FMNH	–
JCK11064	*Praomys misonnei*	Yegue	2/20/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	238	100	138	25	20	25	F	Ad.	OP	8.148	0.646	FMNH	–
JCK11083	*Praomys misonnei*	Yegue	2/23/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	255	108	147	23	17	32	F	Sad.	OP	8.145	0.646	FMNH	–
JCK11084	*Praomys misonnei*	Yegue	2/23/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	234	107	127	22	18	34	F	Ad.	OP	8.146	0.646	FMNH	–
JCK11085	*Praomys misonnei*	Yegue	2/23/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	207	93	114	21	17	26	F	Ad.	OP	8.146	0.646	FMNH	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK11078	*Praomys misonnei*	Yegue	2/23/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	176	81	95	22	18	21	F	Sad.	OP	8.145	0.646	FMNH	–
JCK11080	*Praomys misonnei*	Yegue	2/23/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	179	91	88	22	18	19.5	F	Juv.	OP	8.145	0.646	FMNH	–
JCK11057	*Praomys misonnei*	Yegue	2/20/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	21.5	16.5	16	F	Juv.	OP	8.145	0.646	FMNH	–
JCK10774	*Praomys misonnei*	Yegue	2/9/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	142	90	52	19	NA	22	M	–	CF	8.177	0.656	FMNH	PX147331
TCD5477	*Praomys misonnei*	Assoukoko	3/1/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	229	110	119	NA	NA	37	M	Ad.	GF	8.014	0.635	FMNH	PX147321
TCD5478	*Praomys misonnei*	Assoukoko	3/1/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	278	123	155	24	19	50	F	Ad.	GF	8.014	0.635	FMNH	PX147335
TCD5479	*Praomys misonnei*	Assoukoko	3/1/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	208	98	110	19	17	25	F	Sad.	GF	8.014	0.635	FMNH	PX147322
TCD5480	*Praomys misonnei*	Assoukoko	3/2/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	110*	72	38*	20	14	9.4	F	Juv.	GF	8.014	0.635	FMNH	–
TCD5481	*Praomys misonnei*	Assoukoko	3/2/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	271	122	149	22	NA	41	F	Ad.	GF	8.014	0.635	FMNH	–
TCD5482	*Praomys misonnei*	Assoukoko	3/2/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	167*	124	43*	22	19	42	F	Ad.	GF	8.014	0.635	FMNH	–
TCD5483	*Praomys misonnei*	Assoukoko	3/2/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	153	72	81	19	14	NA	F	Juv.	GF	8.014	0.635	FMNH	PX147336
TCD5484	*Praomys misonnei*	Assoukoko	3/2/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	NA	NA	NA	NA	21	9.3	F	Juv.	GF	8.014	0.635	FMNH	–
TCD5485	*Praomys misonnei*	Assoukoko	3/2/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	151	74	77	19	14	8.7	F	Juv.	GF	8.014	0.635	FMNH	–
TCD5486	*Praomys misonnei*	Assoukoko	3/2/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	292	128	164	23	17	44	F	Ad.	GF	8.014	0.635	FMNH	–
TCD5487	*Praomys misonnei*	Assoukoko	3/3/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	NA	NA	150	22	NA	NA	F	Ad.	GF	8.014	0.635	FMNH	–
TCD5488	*Praomys misonnei*	Assoukoko	3/3/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	270	120	150	21	19	46.5	M	Ad.	GF	8.014	0.635	FMNH	–
TCD5489	*Praomys misonnei*	Assoukoko	3/3/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	260	116	144	22	NA	42	F	Ad.	GF	8.014	0.635	FMNH	–

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
TCD5490	*Praomys misonnei*	Assoukoko	3/3/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	265	118	147	21	17	37	F	Ad.	GF	8.014	0.635	FMNH	–
TCD5491	*Praomys misonnei*	Assoukoko	3/3/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	253	112	141	22	18	33	F	Ad.	GF	8.014	0.635	FMNH	–
TCD5492	*Praomys misonnei*	Assoukoko	3/3/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	156	73	83	20	14	9.5	M	Juv.	GF	8.014	0.635	FMNH	–
TCD5493	*Praomys misonnei*	Assoukoko	3/3/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	273	120	153	22	199	46	F	Ad.	GF	8.014	0.635	FMNH	–
TCD5494	*Praomys misonnei*	Assoukoko	3/4/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	143	68	75	18.5	13	8.6	F	Juv.	GF	8.014	0.635	FMNH	–
TCD5495	*Praomys misonnei*	Assoukoko	3/4/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	234	99	135	24	18	38	F	Ad.	GF	8.014	0.635	FMNH	–
TCD5496	*Praomys misonnei*	Assoukoko	3/4/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	139	76	63	20.5	13.5	9.5	F	Juv.	GF	8.014	0.635	FMNH	–
TCD5499	*Praomys misonnei*	Assoukoko	3/5/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	265	107	158	22	19	35	F	Ad.	GF	8.014	0.635	FMNH	–
TCD5500	*Praomys misonnei*	Assoukoko	3/5/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	256	111	145	22	19	41	F	Ad.	GF	8.014	0.635	FMNH	–
TCD5505	*Praomys misonnei*	Assoukoko	3/6/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	119	60	59	16	10	5.8	F	Juv.	GF	8.014	0.635	FMNH	–
TCD5506	*Praomys misonnei*	Assoukoko	3/6/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	170	78	92	20	14.5	11	M	Juv.	GF	8.014	0.635	FMNH	–
TCD5507	*Praomys misonnei*	Assoukoko	3/6/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	263	114	149	23	NA	37	F	Ad.	GF	8.014	0.635	FMNH	–
TCD5512	*Praomys misonnei*	Assoukoko	3/7/2023	T. C. Demos, J. C. Kerbis Peterhans, H. L. Lutz, A. Hounmavo, D. Broussounou	255	108	147	23	20	42	M	Ad.	GF	8.014	0.635	FMNH	–
JCK11215	*Praomys misonnei*	Diguingue	4/27/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	224	109	115	23	16	36	F	Ad.	GF	8.077	0.644	UL	–
JCK11223	*Praomys misonnei*	Diguingue	5/22/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	242	109	133	22	16	35	F	Ad.	GF	8.077	0.644	UL	PX147323
JCK11224	*Praomys misonnei*	Diguingue	5/22/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	222	101	121	22	15	28	F	Ad.	GF	8.077	0.644	UL	–
JCK11225	*Praomys misonnei*	Diguingue	5/22/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	145	73	72	19	14	12	M	Juv.	GF	8.077	0.644	UL	PX147324
JCK11226	*Praomys misonnei*	Diguingue	5/22/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	246	120	126	22	18	47	F	Ad.	GF	8.077	0.644	UL	PX147305

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK11227	*Praomys misonnei*	Diguingue	5/23/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	246	107	139	23	17	38	F	Ad.	GF	8.077	0.644	UL	PX147325
JCK11228	*Praomys misonnei*	Diguingue	5/23/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	261	120	141	23	19	50	M	Ad.	GF	8.077	0.644	UL	PX147326
JCK11229	*Praomys misonnei*	Diguingue	5/24/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	280	125	155	24	19	49	F	Ad.	GF	8.077	0.644	UL	PX147327
JCK11230	*Praomys misonnei*	Diguingue	5/24/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	261	125	136	24	NA	45	F	Ad.	GF	8.077	0.644	UL	PX147306
JCK11231	*Praomys misonnei*	Diguingue	5/25/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	147	69	78	19	13	88	F	Juv.	GF	8.077	0.644	UL	PX147328
JCK11232	*Praomys misonnei*	Diguingue	5/25/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	149	72	77	19	14	10	M	Sad.	GF	8.077	0.644	UL	PX147329
JCK11234	*Praomys misonnei*	Diguingue	5/26/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	180	85	95	21	13	15	F	Sad.	GF	8.077	0.644	UL	PX147307
JCK11235	*Praomys misonnei*	Diguingue	5/26/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	286	137	149	24	18	67	M	Ad.	GF	8.077	0.644	UL	PX147337
JCK11236	*Praomys misonnei*	Diguingue	5/26/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	180	89	94	22	15	18	M	Juv.	GF	8.077	0.644	UL	PX147308
JCK11237	*Praomys misonnei*	Diguingue	5/26/2023	A. Hounmavo, D. Broussounou, V. Mawougnigan	246	189	127	24	13	50	M	Ad.	GF	8.077	0.644	UL	PX147309
JCK11246	*Praomys misonnei*	Diguingue	7/27/2023	A. Hounmavo, D. Broussounou	223	112	111	NA	11	30	F	Ad.	GF	8.077	0.644	UL	PX147310
JCK11247	*Praomys misonnei*	Diguingue	7/27/2023	A. Hounmavo, D. Broussounou	252	115	137	24	*	32	F	Ad.	GF	8.077	0.644	UL	PX147332
JCK11248	*Praomys misonnei*	Diguingue	7/27/2023	A. Hounmavo, D. Broussounou	NA	NA	NA	NA	NA	15	–	–	GF	8.077	0.644	UL	PX147311
JCK11249	*Praomys misonnei*	Diguingue	7/27/2023	A. Hounmavo, D. Broussounou	284	119	165	23	17	47	F	Ad.	GF	8.077	0.644	UL	PX147333
JCK11250	*Praomys misonnei*	Diguingue	7/28/2023	A. Hounmavo, D. Broussounou	157	72	85	20	13	11	M	Juv.	GF	8.077	0.644	UL	PX147312
JCK11251	*Praomys misonnei*	Diguingue	7/28/2023	A. Hounmavo, D. Broussounou	256	115	141	21	18	36	F	Ad.	GF	8.077	0.644	UL	PX147313
JCK11252	*Praomys misonnei*	Diguingue	7/29/2023	A. Hounmavo, D. Broussounou	255	122	133	24	*	44	M	Ad.	GF	8.077	0.644	UL	PX147334
JCK11259	*Praomys misonnei*	Diguingue	7/30/2023	A. Hounmavo, D. Broussounou	280	125	155	22	17	54	F	Ad.	GF	8.077	0.644	UL	PX147338
JCK11262	*Praomys misonnei*	Diguingue	7/31/2023	A. Hounmavo, D. Broussounou	276	119	157	24	19	40	F	Ad.	GF	8.077	0.644	UL	PX147314
JCK11274	*Praomys misonnei*	Diguingue	9/7/2023	A. Hounmavo, D. Broussounou	225	101	124	20	16	29	F	Ad.	GF	8.077	0.644	UL	PX147315
JCK11277	*Praomys misonnei*	Diguingue	9/9/2023	A. Hounmavo, D. Broussounou	249	105	144	NA	NA	NA	F	Ad.	GF	8.077	0.644	UL	PX147316
JCK11279	*Praomys misonnei*	Diguingue	9/9/2023	A. Hounmavo, D. Broussounou	294	134	160	25	19	56	M	Ad.	GF	8.077	0.644	UL	PX147317
JCK11282	*Praomys misonnei*	Diguingue	10/19/2023	A. Hounmavo, D. Broussounou	175	79	96	21	15	17	F	Sad.	GF	8.077	0.644	UL	–
JCK11285	*Praomys misonnei*	Diguingue	10/19/2023	A. Hounmavo, D. Broussounou	192	77	115	13	16	20	M	Ad.	GF	8.077	0.644	UL	–
JCK11291	*Praomys misonnei*	Diguingue	10/21/2023	A. Hounmavo, D. Broussounou	210	95	115	11	10	26	F	Sad.	CF	8.079	0.645	UL	–
JCK11294	*Praomys misonnei*	Diguingue	10/23/2023	A. Hounmavo, D. Broussounou	261	131	130	29	NA	53	M	Ad.	GF	8.077	0.644	UL	–
JCK11297	*Praomys misonnei*	Diguingue	10/23/2023	A. Hounmavo, D. Broussounou	275	119	156	24	18	43	F	Ad.	GF	8.077	0.644	UL	–
*AH*68	*Rattus rattus*	Diguingue	11/24/2022	A. Hounmavo, D. Broussounou	191	86	105	19	14	17	F	Sad.	CF	8.079	0.645	UL	PX147344
*AH*61	*Rattus rattus*	Diguingue	11/25/2022	A. Hounmavo, D. Broussounou	267	110	157	23	19	56	M	Ad.	GF	8.072	0.647	UL	PX147345
*AH*72	*Rattus rattus*	Assoukoko	11/22/2022	A. Hounmavo, D. Broussounou	251	115	136	26	19	41	M	Sad.	CF	8.011	0.642	UL	–
*AH*71	*Rattus rattus*	Assoukoko	11/22/2022	A. Hounmavo, D. Broussounou	220	95	125	23	18	20	M	Juv.	CF	8.011	0.642	UL	PX147346
*AH*79	*Rattus rattus*	Assoukoko	11/22/2022	A. Hounmavo, D. Broussounou	248	110	138	25	19	38	F	Sad.	CF	8.011	0.642	UL	–
*AH*77	*Rattus rattus*	Assoukoko	11/22/2022	A. Hounmavo, D. Broussounou	247	118	129	25	18	40	M	Sad.	CF	8.011	0.642	UL	–
*AH*69	*Rattus rattus*	Assoukoko	11/21/2022	A. Hounmavo, D. Broussounou	376	153	223	36	20	NA	M	Ad.	CF	8.011	0.642	UL	–
*AH*65	*Rattus rattus*	Assoukoko	11/22/2022	A. Hounmavo, D. Broussounou	263	123	140	29	19	40	M	Sad.	CF	8.011	0.642	UL	–
*AH*62	*Rattus rattus*	Assoukoko	11/20/2022	A. Hounmavo, D. Broussounou	340	143	197	28	18	88	F	Ad.	CF	8.081	0.641	UL	PX147347
*AH*60	*Rattus rattus*	Assoukoko	11/21/2022	A. Hounmavo, D. Broussounou	355	145	210	25	20	76	M	Ad.	CF	8.011	0.642	UL	–
*AH*N3	*Rattus rattus*	Assoukoko	11/25/2022	A. Hounmavo, D. Broussounou	238	108	130	26	16	NA	F	Ad.	CF	8.011	0.642	UL	PX147348
*AH*N4	*Rattus rattus*	Assoukoko	11/22/2022	A. Hounmavo, D. Broussounou	121	108	13	27	15	NA	F	Ad.	CF	8.011	0.642	UL	PX147349
*AH*N2	*Rattus rattus*	Assoukoko	11/22/2022	A. Hounmavo, D. Broussounou	234	111	123	26	16	NA	M	Ad.	CF	8.011	0.642	UL	PX147351
*AH*N12	*Rattus rattus*	Assoukoko	11/22/2022	A. Hounmavo, D. Broussounou	226	105	121	27	17	NA	F	Ad.	CF	8.011	0.642	UL	PX147350

Appendix 2. — Continuation.

ID	Species	Locality	Date	Collectors	TOT	HB	TAIL	HF	EAR	Weight	Sex	Age	Hab.	Lat.	Long.	Inst.	GenBank
JCK11163	*Rattus rattus*	Assoukoko	3/2/2023	J. C. Kerbis Peterhans, H. L. Lutz, T. C. Demos, A. Hounmavo, D. Broussounou	347	158	189	33	20	101	M	Ad.	CF	8.011	0.642	FMNH	PX147352
JCK11165	*Rattus rattus*	Assoukoko	3/4/2023	J. C. Kerbis Peterhans, H. L. Lutz, T. C. Demos, A. Hounmavo, D. Broussounou	365	170	195	35	22	115	F	Ad.	CF	8.011	0.642	FMNH	PX147353
JCK11169	*Rattus rattus*	Assoukoko	3/5/2023	J. C. Kerbis Peterhans, H. L. Lutz, T. C. Demos, A. Hounmavo, D. Broussounou	417	203	214	38	24	188	M	Ad.	CF	8.011	0.642	FMNH	PX147354
JCK11170	*Rattus rattus*	Assoukoko	3/6/2023	J. C. Kerbis Peterhans, H. L. Lutz, T. C. Demos, A. Hounmavo, D. Broussounou	319	137	182	32	20	71	F	Sad.	CF	8.011	0.642	FMNH	PX147355
JCK11171	*Rattus rattus*	Assoukoko	3/6/2023	J. C. Kerbis Peterhans, H. L. Lutz, T. C. Demos, A. Hounmavo, D. Broussounou	317	141	176	33.5	21	65	F	Juv.	CF	8.011	0.642	FMNH	PX147356
JCK10838	*Rattus rattus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	225	114	111	27	20	30	F	Juv.	CF	8.177	0.656	FMNH	–
JCK10883	*Rattus rattus*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	233	118	115	29.5	21	33.5	M	Juv.	CF	8.177	0.656	FMNH	–
JCK10884	*Rattus rattus*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	267	122	145	28.5	21.5	45	F	Juv.	CF	8.177	0.656	FMNH	PX147357
JCK10885	*Rattus rattus*	Yegue	2/13/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	236	112	124	29	19	30	M	Juv.	CF	8.177	0.656	FMNH	PX147358
*AH*N5	*Stochomys longicaudatus*	Assoukoko	11/21/2022	A. Hounmavo, D. Broussounou	336	141	195	28	16	NA	F	Ad.	GF	8.014	0.635	UL	PX147339
JCK11164	*Stochomys longicaudatus*	Assoukoko	3/3/2023	J. C. Kerbis Peterhans, H. L. Lutz, T. C. Demos, A. Hounmavo, D. Broussounou	192*	135	57*	17.5	32	69	M	Ad.	GF	8.014	0.635	FMNH	PX147341
JCK11173	*Stochomys longicaudatus*	Assoukoko	3/6/2023	J. C. Kerbis Peterhans, H. L. Lutz, T. C. Demos, A. Hounmavo, D. Broussounou	332	139	193	29	18	63	F	Sad.	GF	8.014	0.635	FMNH	PX147340
JCK10831	*Stochomys longicaudatus*	Yegue	2/11/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	NA	NA	NA	29	18	68	M	Ad.	OP	8.145	0.647	FMNH	PX147342
JCK11036	*Stochomys longicaudatus*	Yegue	2/19/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	325	134	191	29	19.5	56	F	Ad.	OP	8.145	0.646	FMNH	PX147343
JCK11289	*Stochomys longicaudatus*	Diguingue	10/20/2023	A. Hounmavo, D. Broussounou	319	130	189	27	19	76	F	Ad.	GF	8.077	0.644	UL	–
JCK10779	*Uranomys ruddi*	Yegue	2/10/2022	J. C. Kerbis Peterhans, H. L. Lutz, R. Issaka, D. Broussounou, V. Mawougnigan	102	62	40	15	11	6.7	M	–	CF	8.177	0.656	FMNH	PX147230
JCK11179	*Euxerus erythropus*	Assoukoko	3/7/2023	J. C. Kerbis Peterhans, H. L. Lutz, T. C. Demos, A. Hounmavo, D. Broussounou	308	168	140	59	15	155	M	Ad.	CF	8.011	0.642	FMNH	PX147229
AH201	*Graphiurus* sp. *B*	Diguingue	11/21/2022	A. Hounmavo, D. Broussounou	–	84	32	15	12	NA	M	Ad.	CF	8.079	0.645	UL	–