

# ***Lactarius* subg. *Lactarius* (Russulaceae) in Indian Himalaya: two new species with morphology and phylogenetic inferences**

Priyanka UNİYAL<sup>a</sup>, Jorinde NUYTINCK<sup>b</sup>, Kanad DAS<sup>c\*</sup>

<sup>a</sup>Department of Botany & Microbiology, H.N.B. Garhwal University, Srinagar, Garhwal – 246174, Uttarakhand, India, email: uniyalsppriyanka@gmail.com

<sup>b</sup>Naturalis Biodiversity Center, P.O. Box 9517, 2300RA Leiden, The Netherlands, email: Jorinde.Nuytinck@naturalis.nl

<sup>c</sup>Cryptogamic Unit, Botanical Survey of India, P.O. Botanic Garden, Howrah – 711103, India, email: daskanadbsi@gmail.com

**Abstract** – The present paper deals with two novel species in *Lactarius* subg. *Lactarius* collected from western Himalaya (India): *Lactarius indozonarius* sp. nov. and *Lactarius thindii* sp. nov. Morphological descriptions, illustrations and nrITS-based phylogenetic analyses of these species are presented.

**Milkcaps / Macrofungi / nrITS / phylogeny / Russulales / taxonomy / western Himalaya**

## INTRODUCTION

*Lactarius* Pers. is a large ectomycorrhizal genus playing a significant role as a mycobiont in different ecosystems, from tropical to temperate and arctic-alpine regions (Verbeken & Nuytinck 2013, Leonardi *et al.* 2016). Species traditionally placed in *L.* subg. *Lactarius* (one of the three subgenera) are characterized in the field by a slimy to viscid or shiny, rarely dry, zonate to azonate pileus, with or without a hairy margin; zonate to azonate pileus context; a dry or sticky stipe which is often scrobiculate and variable latex colour. Microscopically, they typically have an ixocutis to ixotrichoderm (more rarely a cutis to a trichoderm) type of pileipellis (Le *et al.* 2007, Verbeken & Nuytinck 2013). This subgenus is very well represented in all Northern hemisphere continents (Le *et al.* 2007).

In Indian Himalaya, macrofungal explorations by various workers, especially in the past 25 years, reported 29 species so far. These are *Lactarius abbotanus* K. Das & J. R. Sharma, *L. alnicola* A. H. Sm., *L. byssaceus* K. Das & Verbeken, *L. controversus* Pers., *L. dafianus* K. Das *et al.*, *L. deliciosus* (L.) Gray, *L. deterrimus* Gröger, *L. dhakurianus* K. Das *et al.*, *L. elaioviscidus* K. Das & Verbeken, *L. fennoscandicus* Verbeken & Vesterh., *L. indigo* (Schwein.) Fr.,

\* Corresponding author: daskanadbsi@gmail.com

*L. kumaonensis* Uniyal *et al.*, *L. maitlyensis* K. Das *et al.*, *L. mayawatianus* K. Das & J. R. Sharma, *L. olivaceoglutinus* K. Das & Verbeken, *L. olivaceoumbrinus* Hesler & A. H. Sm., *L. paradoxus* Beardslee & Burl., *L. proximellus* Beardslee & Burl., *L. pseudoaspideus* Hesler & A. H. Sm., *L. pubescens* Fr., *L. pyriodorus* K. Das & Verbeken, *L. rubrifluus* Gillet, *L. sanguifluus* (Paulet) Fr., *L. scrobiculatus* (Scop.) Fr., *L. subindigo* Verbeken & E. Horak, *L. subpurpureus* Peck, *L. yazooensis* Hesler & A. H. Sm., *L. yumthangensis* K. Das & Verbeken and *L. zonarius* (Bull.) Fr. (Atri *et al.* 1994, Das & Sharma 2005, Das & Verbeken 2011, Joshi *et al.* 2012, Das 2013, Das *et al.* 2015, Uniyal *et al.* 2017). Among these reported species, several are European and American representatives of *L.* subg. *Lactarius* that are reported from India based on morphological similarity. DNA based studies are needed to confirm species delimitation and intercontinental conspecificity for a large number of these species (*L. alnicola*, *L. deliciosus*, *L. deterrimus*, *L. fennoscandicus*, *L. indigo*, *L. olivaceoumbrinus*, *L. paradoxus*, *L. proximellus*, *L. pseudoaspideus*, *L. pubescens*, *L. rubrifluus*, *L. sanguifluus*, *L. scrobiculatus*, *L. subpurpureus*, *L. yazooensis* and *L. zonarius*). Therefore, collection of more samples and a thorough revision of this group from Indian Himalaya with the support of molecular phylogeny is needed. Unfortunately, macrofungus surveys in Russulaceae have nearly all been undertaken

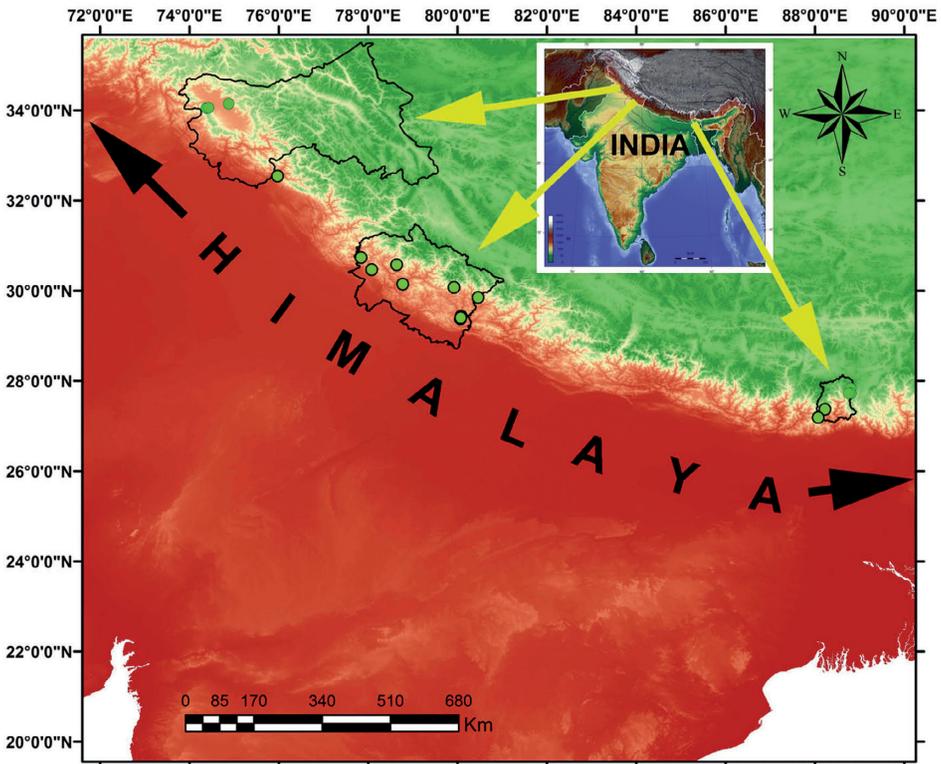


Fig 1. Map of Indian Himalayan region showing sites for the occurrences (only small patches in three states) of *Lactarius* subg. *Lactarius* members with green dots.

in Uttarakhand, Sikkim and Himachal Pradesh (Fig. 1) while the rest of Indian Himalaya remains unexplored.

While undertaking routine macrofungal surveys in temperate forests of Uttarakhand (Figs. 3-6), two interesting taxa were encountered by one (PU) of us. Thorough morphological examination and nrITS-based phylogenetic data of these taxa revealed two novel species within *Lactarius* subg. *Lactarius* and we describe them here as *Lactarius indozonarius* (*L.* sect. *Zonarii*) and *Lactarius thindii* (*L.* sect. *Colorati*).

## MATERIAL AND METHODS

### Collecting and morphology

Macromorphological characters and field data were recorded in the forest or basecamp from the fresh and dissected young to mature basidiomata. Macrochemical colour reactions on pileus, stipe surface, context and latex were also recorded. Images of the fresh basidiocarps were captured with the help of Canon Power Shot SX 50 HS and Sony Cyber Shot W730 cameras. Color codes and terms follow Methuen Handbook of Color (Kornerup & Wanscher 1978). Micromorphological characters were observed with the help of a light microscope (Olympus CH20i) from dry material mounted in a mixture of 5% KOH, 1% Phloxin and 1% Congo red. Drawings of micromorphological elements were made with a camera lucida at 2000 × magnification. Microphotographs were captured with a digital camera attached to the Olympus CH20i and Olympus-CX21iLED light microscopes. Basidiospores were mounted in Melzer's reagent and measured in lateral view excluding the height of ornamentation and apiculus. Basidiospore measurements are presented as minimum-*mean*-maximum length × minimum-*mean*-maximum width and Q = length/width ratio.

Scanning Electron Microscope (SEM) images of basidiospores were obtained from dry basidiospores that were directly mounted on a double-sided adhesive tape pasted on a metallic specimen-stub and then scanned with silver coating at different magnifications in high vacuum mode (20 KV). SEM-studies were carried out with a ZEISS EVO 18 SPECIAL EDITION model installed at USIC Dept., HNBSGU Srinagar (Garhwal), India. Herbarium codes follow Thiers (continuously updated).

### DNA isolation, amplification and sequencing

Genomic DNA was isolated from 100 mg of (dry) tissue of the basidiomata using the XcelGen Plant Fungal gDNA kit (XG2416-01) following the manufacturer's instructions. The nrITS region was amplified with primer pairs ITS1-F and ITS4-B (White *et al.* 1990). PCR amplification was performed using a thermocycler (ABI, Veriti) programmed for 3 min at 95°C, followed by 1 min at 94°C, 30 cycles of 45 sec at 48°C, 45 seconds at 72°C and a final stage of 10 min at 72°C. The PCR amplicon was purified with exosap enzymatic purification as per the manufacturer instruction (ABI). After the purification the products were subjected to Sanger sequencing using an ABI 3730XL DNA analyzer. Forward and reverse strands were

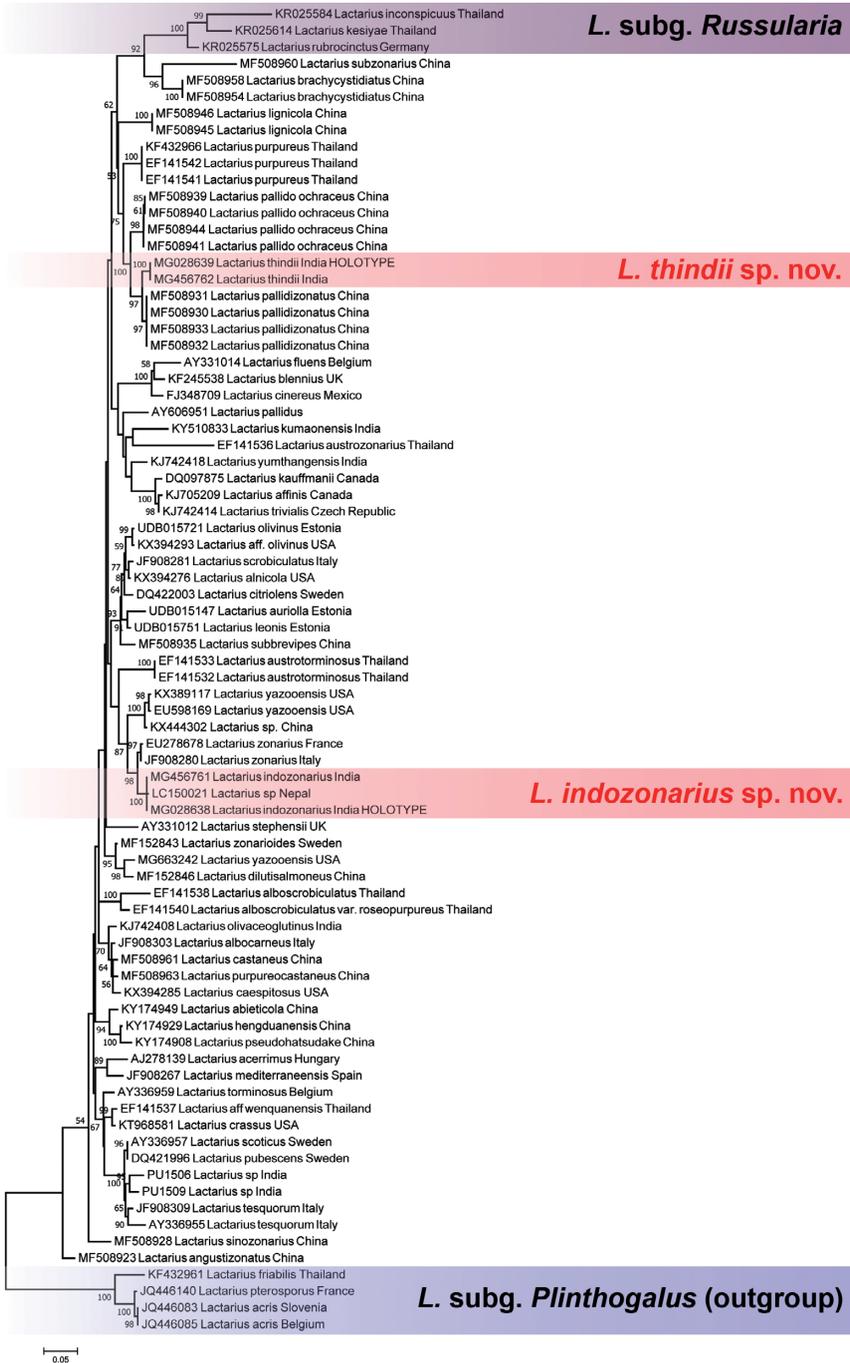


Fig 2. Phylogram generated from ITS-rDNA sequences: The evolutionary history was inferred by using the Maximum Likelihood method in RAxML v. 8.0.24. Bootstrap support values (>50%) obtained from the ML analysis are shown above or below the branches at nodes.

sequenced separately with the ITS1 and ITS4 primers using BDT v3.1 Cycle sequencing kit. Sequences generated in this study are deposited in GenBank.

### Phylogenetic analysis

A dataset with reference nrITS sequences was assembled by considering the sequences from studies on *L.* subg. *Lactarius* (Le *et al.* 2007, Nuytinck *et al.* 2007, Das *et al.* 2015, Uniyal *et al.* 2017, Wang 2017, Shi *et al.* 2018), and a BLAST (Altschul *et al.* 1997) search in GenBank (Clark *et al.* 2016) and UNITE (Kõljalg *et al.* 2013). We aimed for the inclusion of as many Asian representatives as possible. Phylogenetic analyses were carried out to establish the relationships of our new taxa. Species of *L.* subg. *Plinhogalus* were selected as outgroup. Multiple sequence alignment was performed using MAFFT v.7 (Katoh & Standley 2013), applying the E-INS-I strategy. Alignments were manually edited in Bioedit v 7.2.5 (Hall *et al.* 1999). Maximum Likelihood (ML) analyses were conducted in RAxML v. 8.0.24 (Stamatakis 2014) on the CIPRES Science Gateway (Miller *et al.* 2010).

## RESULTS

### Phylogenetic inference

Our ITS-based phylogenetic analysis being conducted with 109 nucleotide sequences (Fig. 2) supports the placement of *L. indozonarius* sp. nov. and *L. thindii* sp. nov. in *L.* subg. *Lactarius*. Although this subgenus is paraphyletic in our analysis with *L.* subg. *Russularia* nested inside, all close relatives of the newly described species are traditionally considered to be members of *L.* subg. *Lactarius*.

Delimiting sections and subsections in *Lactarius* is impossible based on ITS sequences only, but the two new species described here fall in well-supported species groups that also make sense morphologically. This is discussed in more detail in the taxonomy section of this manuscript.

### Taxonomy

***Lactarius indozonarius*** Uniyal, K. Das & Nuytinck, **sp. nov.** **Figs 7-20**

*Mycobank*: MB823734

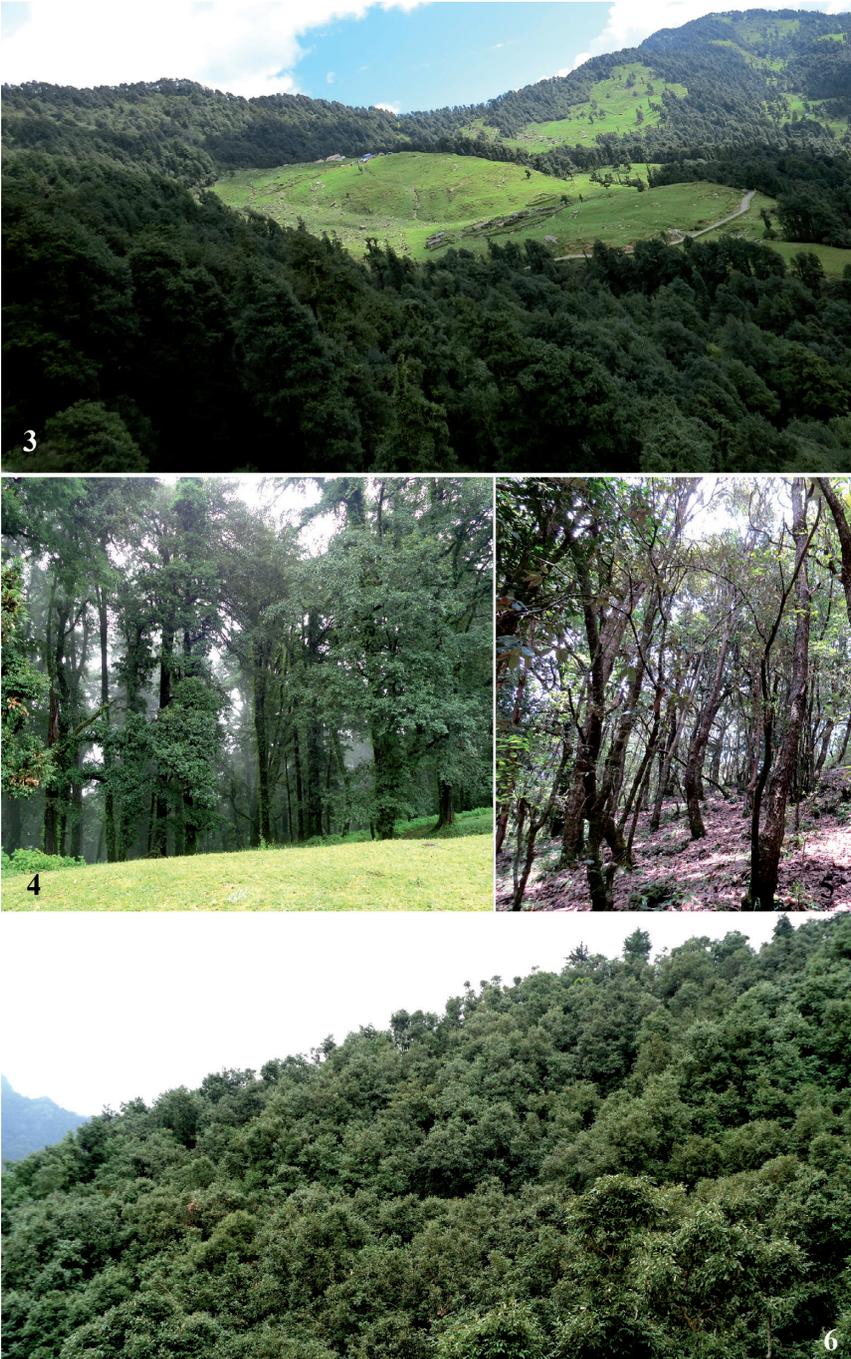
*Genbank*: MG028638 (holotype), MG456761 (paratype)

*Diagnosis*: Differs from *L. zonarius* by its zoned context and smaller basidiospores with higher (up to 1.5 µm) ornamentations and by its nrITS data.

*Holotype*: INDIA: **Uttarakhand**, Pauri Garhwal, Chaubatta, N30° 09.697' E078° 51.195', 1913 m asl., growing in a mixed temperate forest dominated by *Quercus*, *Rhododendron*, *Myrica*, *Cedrus* and *Pinus* sp., 16th July 2016, P. Uniyal, PU-1087 (CAL;).

*Etymology*: Referring to morphological resemblance and close affinity with *L. zonarius*, a European species.

**Pileus** 60-122 mm diam., convex to plano-convex, shallowly depressed, finally infundibuliform; surface viscid to glutinous, zonate, increasingly matted-



Figs 3-6. Habitats. 3-4. Mixed forest of Chopta-Baniyakund; collection site of *Lactarius thindii*. 5-6. Mixed forest of Chaubatta and Chaurikhal dominated by *Quercus leucotrichophora*, collection site of *Lactarius indozonarius*.

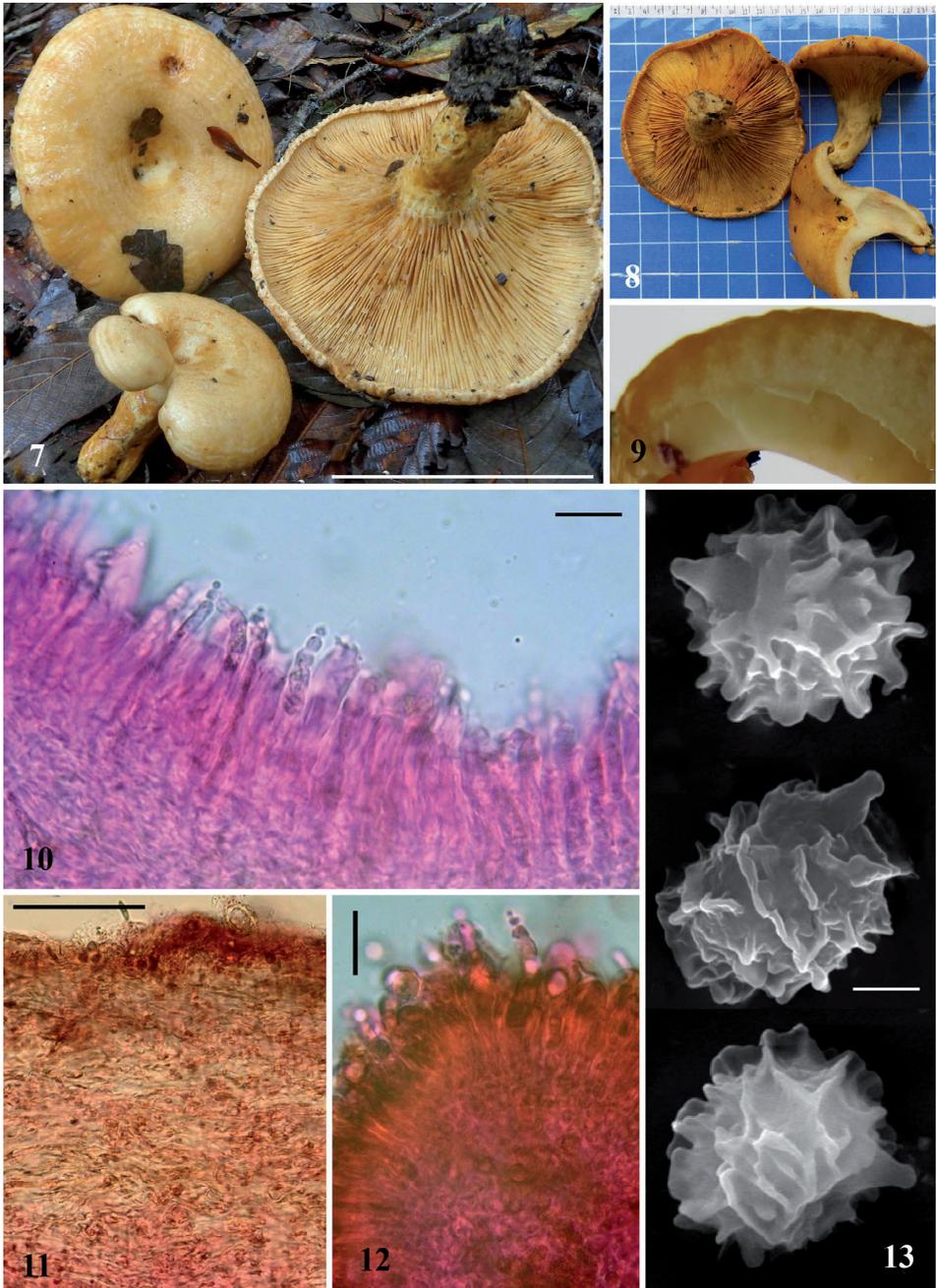
fibrillose towards margin, straw yellow to wax yellow (3B4-5) when young, light yellow (4A4-5) to greyish yellow (4B3) or corn yellow (4B5) with age, finally brownish yellow (5C6-8), sometimes paler near margin, hairs up to 2 mm long, turning brownish yellow (5C7); zonations greyish orange (5B3-4); margin inrolled to decurved, undulate to entire, radially ridged when young, hairy. **Lamellae** subdecurrent, very crowded (15-17 L+l/cm), pale yellow to pastel yellow (3A3-4) when young, greyish yellow (4B4-5) with age, brownish orange (5C5) where damaged, some forked near attachment; lamellulae numerous. **Stipe** 45-72 × 13-29 mm, cylindrical, equal, scrobiculated, pale yellow (3A3) to greyish yellow (3B4-5), yellowish white (3A2) near attachment; scrobicules watery or yellowish white (3A2), becoming greyish yellow to yellowish orange (4B5-7) on handling or with age. **Context** 7-10 mm thick, white to yellowish white (2A2), zonate, zonations watery, continuations in zones of pileus surface, gradually disappearing toward centre; unchanging with FeSO<sub>4</sub> and KOH, turning pastel red (8A4-5) at first with guaiac, then brownish red (8C8); taste mild to slightly acid; stuffed to hollow in stipe. **Latex** scarce, white, unchanging; taste acrid on tissue and in isolation. **Odor** pleasant, fruity. **Spore deposit** yellowish white (1A2).

**Basidiospores** 7-7.5-8.5 × 6.5-6.9-7.5 μm, subglobose to broadly ellipsoid, (n = 20, Q = 1.07-1.13-1.23), ornamentations ≤1.5 μm high, rounded to triangular warts and connecting ridges never forming a complete reticulum; plage inamyloid. **Basidia** 40-51 × 9-12 μm, subclavate, 2-, 4-spored; sterigmata 4-5 μm long. **Pleuromacrocystidia** abundant, 30-63 × 4-7.5 μm, cylindric to subfusiform, apices capitate to mucronate and constricted, mostly moniliform; contents dense, small crystal-like. **Pleuropsychocystidia** up to 5.5 μm thick, nonemergent to slightly emergent, cylindric, with a round apex and often forked near apex. **Lamellae edge** sterile. **Cheilomacrocystidia** 36-42 × 4-5.5 μm, subcylindric to subfusiform, apex moniliform to constricted; contents dense. **Lactifers in hymenophoral trama** up to 10 μm thick. **Pileipellis** an ixocutis, 80-137 μm thick, composed of densely packed parallel, often shrivelled hyphae; hyphae 1-2.5 μm wide. **Lactifers in pilear trama** up to 9 μm thick.

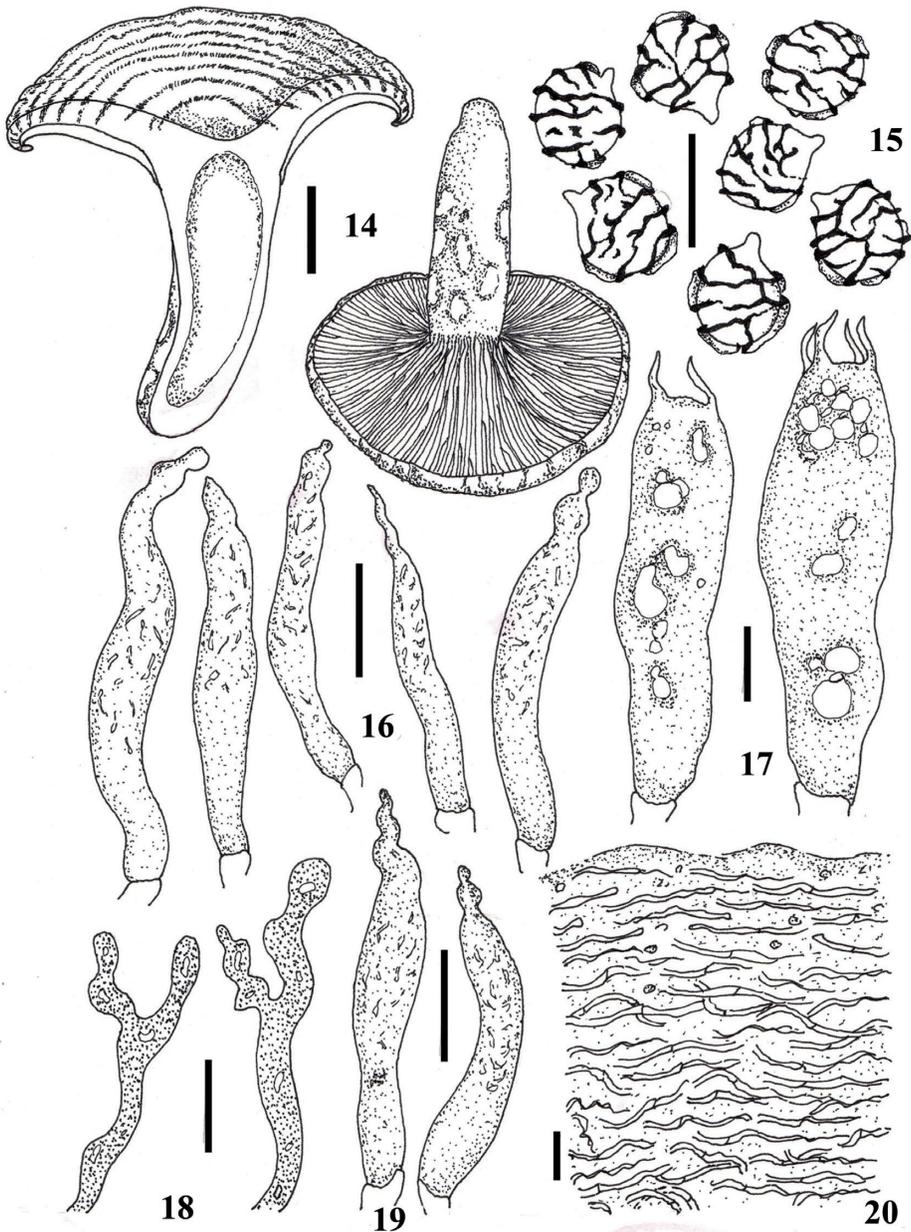
**Ecology:** Growing in association with *Quercus leucotrichophora* in mixed temperate forest dominated by *Quercus*, *Rhododendron*, *Myrica*, *Cedrus* and *Pinus* sp.

**Other collection examined:** INDIA: **Uttarakhand**, Pauri Garhwal, Chaurikhal, N30° 05.681' E078° 43.890', 1989 m asl., 21<sup>st</sup> Aug. 2014, *P. Uniyal*, PU-439 (GUH).

**Comments:** The viscid to glutinous, zonate, yellowish pileus with hairy margin, and the white, unchanging latex, scrobiculate stipe and fruity odour undoubtedly place the present taxon in *Lactarius* sect. *Zonarii*. This is also confirmed in the molecular phylogeny by the placement of *L. indozonarius* as the sister species of *L. zonarius*, although the monophyly of *L. sect. Zonarii* in its traditional morphological sense cannot be confirmed. A Nepalese sample (LC150021) is clustered with *L. indozonarius* but lacks any morphological description. *Lactarius indozonarius* is quite distinct due to the zonations in the pileus context. This character has only been reported in Thai species of *L. sect. Zonarii*: *L. alboscrobiculatus* var. *roseopurpureus* H.T. Le & A. Verbeken, *L. austrotorminosus* H.T. Le & A. Verbeken and *L. austrozonarius* H.T. Le & A. Verbeken (Le *et al.* 2007), but is a generic character for its sister genus *Multifurca* (Wang *et al.* 2018). Microscopically, *L. indozonarius* is characterized by subglobose to broadly ellipsoid basidiospores, abundant macrocystidia and an ixocutis pattern of the pileipellis.



Figs. 7-13. *Lactarius indozonarius* sp. nov. (PU-1087, holotype). 7-8. Fresh basidiomata in field and basecamp. 9. Zonations in context. 10. Pleuromacrocystidia. 11. Transverse section through pileipellis. 12. Cheilomacrocystidia. 13. SEM image of basidiospores. Scale bars 7 = 100 mm; 10 & 12 = 25 µm; 11 = 100 µm; 13 = 2 µm.



Figs. 14-20. *Lactarius indozonarius* sp. nov. (drawings from holotype: PU-1087). 14. Fresh/dissected basidiomata. 15. Basidiospores. 16. Pleuromacrocystidia. 17. Basidia. 18. Pleuropseudocystidia. 19. Cheilomacrocystidia. 20. Pileipellis. Scale bars 14 = 10 mm; 15-19 = 10  $\mu$ m; 20 = 25  $\mu$ m.

*Lactarius indozonarius* is morphologically similar to *L. zonarius* by the presence of a strongly zonate, yellowish pileus, brownish discoloration in lamellae on damage, burning acrid taste, pleasant odour and abundant pleurocystidia. Yet, our

new species is clearly distinct from it due to its hairy pileus and zoned context. Microscopically, *L. zonarius* differs because of its smaller ( $6.9\text{-}7.7 \times 5.8\text{-}6.2 \mu\text{m}$ ) basidiospores with lower ( $0.7 \mu\text{m}$ ) ornamentation (Basso 1999; Heilmann-Clausen *et al.* 1998).

*L. alboscrobiculatus* var. *roseopurpureus* (95% similarity for 100% query coverage in BLAST search) is distinct by its white to cream-coloured pileus, distant lamellae and very low ( $0.5\text{-}0.8 \mu\text{m}$ ) ornamentations on the basidiospores. *L. austrozonarius* differs by its subdistant lamellae, very abundant, white to watery cream-coloured latex and distinctively larger ( $7.2\text{-}12.2 \times 7\text{-}10.2 \mu\text{m}$ ) basidiospores with very high ridges ( $\leq 2.5 \mu\text{m}$ ). *Lactarius indozonarius* shares with *L. austrotorminosus* a hairy cap margin, dense lamellae with brownish orange discoloration and  $\leq 1.5 \mu\text{m}$  high ornamentations on the basidiospores, but the latter species is still recognizable by its pinkish orange basidiomes and smaller (30-80 mm diam.), papillate pileus. Micromorphologically, *L. austrotorminosus* is quite distinct from *L. indozonarius* due to the structure of the pileipellis being a cutis or trichoderm, and the zebroid ornamentation of the basidiospores (Le *et al.* 2007).

***Lactarius thindii*** Uniyal, K. Das & Nuytinck, **sp. nov.**

**Figs 21-33**

*Mycobank*: MB823735

*Genbank*: MG028639 (holotype), MG456762 (paratype)

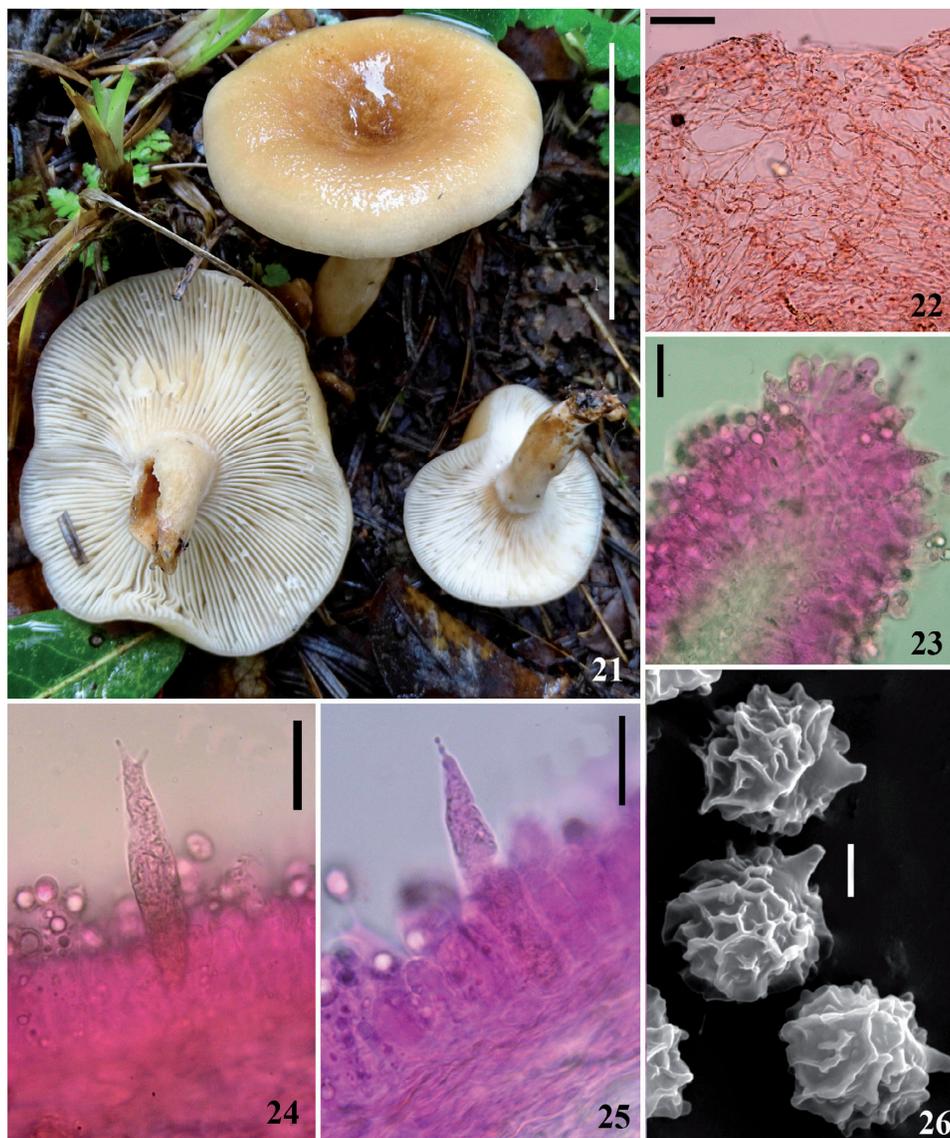
*Diagnosis*: Differs from *Lactarius pallidizonatus* by the presence of an azonate to faintly zonate pileus, non striate pileus margin, unchanging latex, smaller ( $6.0\text{-}7.0 \times 5.0\text{-}6.0 \mu\text{m}$ ) basidiospores and nrITS sequence data.

*Holotype*: INDIA: **Uttarakhand**: Rudraprayag, Chopta-Baniyakund, N30° 28.914' E079° 10.854', 2630 m asl., growing in mixed subalpine forest dominated by *Quercus*, *Rhododendron*, *Lyonia*, *Myrica* and *Abies* spp., 7<sup>th</sup> Aug. 2017, P. Uniyal, PU-1526 (CAL).

*Etymology*: in **commemoration** of Prof. K. S. Thind for his contributions to the study of Indian mycobiota.

**Pileus** 24-37 mm diam., convex to plano-convex with depressed center, surface smooth, sticky to slightly glutinous, azonate to faintly zonate near margin, light orange (5A4-5) to greyish orange (5B3-4), brownish yellow (5C7-8) at centre, pale yellow (4A3) near margin; margin undulate, decurved. **Lamellae** subdecurrent, rather close (6-7 L/cm), white to yellowish white (2A2), unchanging when damaged; lamellulae 1-2 between two lamellae. **Stipe** 25-36  $\times$  6-11 mm, brownish orange (5C4-5), whitish toward apex. **Context** white to orange white (5A2), greyish orange (5B4) in stipe when mature, stuffed to hollow, unchanging on exposure and with FeSO<sub>4</sub> and KOH; taste faintly acrid; odour insignificant. **Latex** scarce, white, unchanging; taste acrid. **Spore deposit** yellowish white (2A2).

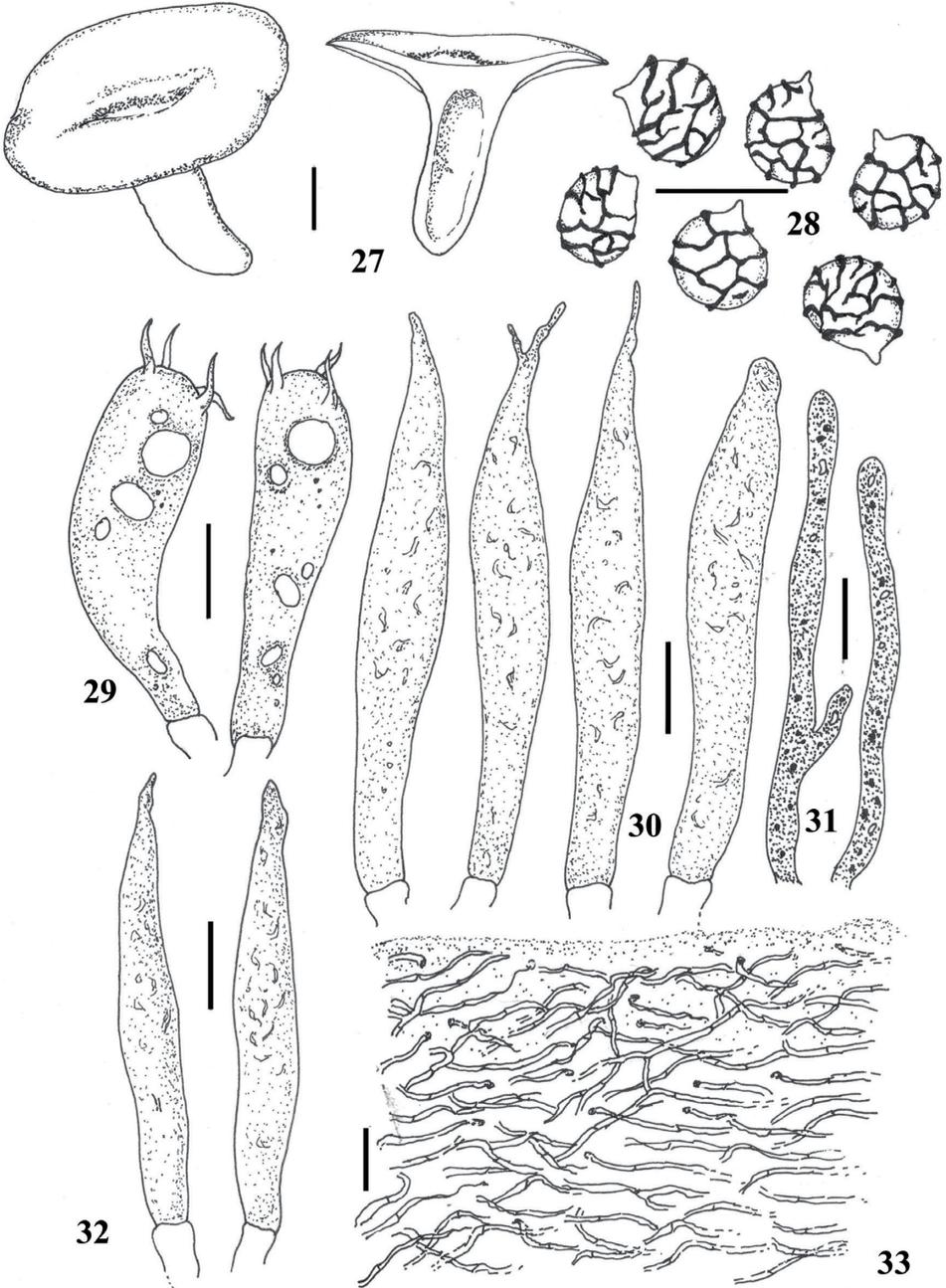
**Basidiospores** 6-6.3-7  $\times$  5-5.2-6  $\mu\text{m}$ , (n = 20, Q = (1) 1.10-1.20-1.27); broadly ellipsoid to ellipsoid, ornamentations composed of some small round to conical warts and ridges ( $\leq 1.2 \mu\text{m}$  high) that form a close incomplete reticulum; plage inamyloid. **Basidia** 33-45  $\times$  9-11  $\mu\text{m}$ , subclavate, 4-spored; sterigmata 3-5  $\mu\text{m}$  long. **Pleuromacrocystidia** 54-82  $\times$  6.5-12  $\mu\text{m}$ , fusiform to subfusiform, often forked near apex; apices acute, capitate and mucronate to moniliform, contents dense, needle shaped. **Pleuropseudocystidia** up to 6.5  $\mu\text{m}$  thick, cylindrical, sometimes forked. **Cheilomacrocystidia** 45-58  $\times$  6.5-8  $\mu\text{m}$ ; apex acute to mucronate. **Lactifers in hymenophoral trama** up to 9  $\mu\text{m}$  thick. **Pileipellis** an ixocutis, 85-165  $\mu\text{m}$  thick; hyphae densely packed, septate, repent to slightly ascending, interwoven in the form of a reticulum, 1-4  $\mu\text{m}$  thick.



Figs. 21-26. *Lactarius thindii* sp. nov. (PU-1526, holotype). **21.** Fresh basidiomata in field. **22.** Transverse section through pileipellis. **23.** Cheilomacrocytidia. **24-25.** Pleuromacrocytidia. **26.** SEM image of basidiospores. Scale bars 21 = 50 mm; 22-25 = 25  $\mu$ m; 26 = 2  $\mu$ m.

*Ecology:* Growing in association with *Quercus semecarpifolia* in a mixed subalpine forest dominated by *Quercus*, *Rhododendron*, *Lyonia*, *Myrica* and *Abies* spp.

*Other collection examined:* INDIA: Uttarakhand, Rudraprayag, Chopta-Baniyakund, N30° 28.998' E079° 10.658', 2653 m asl., 19th July 2015, P. Uniyal, PU-705 (GUH).



Figs. 27-33. *Lactarius thindii* sp. nov. (drawings from holotype: PU-1526). 27. Fresh/dissected basidiomata. 28. Basidiospores. 29. Basidia. 30. Pleuromacrocystidia. 31. Pleuropseudocystidia. 32. Cheilomacrocystidia. 33. Pileipellis. Scale bars 27 = 10 mm; 28-32 = 10 µm; 33 = 25 µm.

*Comments:* *Lactarius thindii* is morphologically characterized by its small sized basidiomes, its sticky to slightly glutinous, greyish orange to brownish yellow pileus that is almost white towards the margin; unchanging, white latex; subglobose to broadly ellipsoid basidiospores with a dense, incomplete reticulum; often forked pleurocystidia and ixocutis type of pileipellis. Molecular data suggest its close relation with *L. purpureus* R. Heim ex R. Heim, *L. pallidizonatus* X.H. Wang and *L. pallido-ochraceus* X.H. Wang. The Asian species *L. purpureus* is unmistakably different from *L. thindii* by its wine red to pink, scaly pileus surface; larger (5.2-8.6 × 4.5-7.9 µm) basidiospores and a cutis to trichoderm type of pileipellis (Verbeken & Horak 2000; Le *et al.* 2007). *Lactarius pallidizonatus* and *L. pallido-ochraceus* share similarities with *L. thindii* in having a faintly zonate to zonate, smooth pileus surface, acrid tasting white latex, ixocutis type of pileipellis and occurrence under fagaceous trees. However, both species are different from *L. thindii* due to presence of hygrophanous pileus surface with sub-transparently striate margin. In addition, *L. pallidizonatus* is distinct by latex changing to watery, larger (7.5-9.0 × 6.0-7.5 µm) basidiospores with lower (0.4-0.6 µm) ornamentations and very thick (200-500 µm) pileipellis. *Lactarius pallido-ochraceus* is different from *L. thindii* due to presence of brownish to greyish ochraceous discoloration in lamellae on bruising and unchanging to glaucous green discolouring latex (Wang 2017).

*Lactarius fraxineus* Romagn. is remarkably similar to *L. thindii* in having crowded, cream-coloured lamellae, white latex, basidiospores (6-7 × 5-6 µm) with incomplete reticulation and an ixocutis type of pileipellis. However, this European species differs from the present taxon by its larger (≤ 85 mm) pileus with scaly surface, scrobiculate stipe and greyish-olive latex discoloration (Basso 1999).

In India, *Lactarius dafianus* is the sole representative of *L. sect. Colorati* and can be confused with *L. thindii* due to its reddish orange pileus with darker centre, dense lamellae and unchanging latex. The former is different on the basis of its watery latex, white spore deposit, abundant isolated warts on the basidiospores and trichodermal pileipellis (Das & Sharma 2005).

*A provisional key to the known species of Lactarius subg. Lactarius from Indian Himalaya*

- |  |                        |
|--|------------------------|
| 1a. Latex strikingly coloured, presence of greenish discoloration in tissue .....  | 2                      |
| 1b. Latex white to watery, absence of greenish discoloration in tissue .....   | 10                     |
| 2a. Pileus greyish blue to blue with silvery lustre .....  | 3                      |
| 2b. Pileus pinkish, brownish or orange .....   | 5                      |
| 3a. Latex vinaceous red, occurrence under <i>Pinus</i> .....   | <i>L. paradoxus</i>    |
| 3b. Latex blue .....   | 4                      |
| 4a. Latex turning green on exposure, basidiospores 7–9.5 × 6–7.5 µm, occurrence under <i>Pinus</i> and <i>Quercus</i> .....            | <i>L. indigo</i>       |
| 4b. Latex unchanging on exposure, basidiospores 6.6–8.3 × 5.4–6.5 µm, occurrence under <i>Castanopsis</i> and <i>Lithocarpus</i> ..... | <i>L. subindigo</i>    |
| 5a. Lamellae subdistant to distant (3-4/cm) .....  | 6                      |
| 5b. Lamellae crowded (12-18/cm) .....  | 7                      |
| 6a. Pileus brownish orange to brown, pleuromacrocytidia scarce, occurrence under <i>Pinus</i> .....                                    | <i>L. rubrifluus</i>   |
| 6b. Pileus dirty pinkish, pleuromacrocytidia abundant, occurrence under <i>Tsuga</i> .....   | <i>L. subpurpureus</i> |
| 7a. Pileus distinctly or faintly zonate .....  | 8                      |
| 7b. Pileus azonate, occurrence under <i>Pinus</i> .....  | <i>L. sanguifluus</i>  |

- 8a. Stipe nonscrobiculate, basidiospore ornamentation consisting of isolated warts and short ridges, never forming reticulum..... 9
- 8b. Stipe scrobiculate, basidiospores ornamentation forming an incomplete reticulum, occurrence under *Pinus*..... *L. deliciosus*
- 9a. Pileus typically azonate or with a few zones near the margin, with mainly bright orange colours and green tinges, occurrence under *Picea* ..... *L. deterrimus*
- 9b. Pileus clearly zonate and brown-orange with sometimes lilac-grey tinges, occurrence under *Picea*..... *L. fennoscandicus*
- 10a. Pileus distinctly to faintly zonate..... 11
- 10b. Pileus azonate..... 23
- 11a. Pileus pubescent to fibrillose..... 12
- 11b. Pileus neither pubescent nor fibrillose..... 15
- 12a. Pileus large ( $\geq 120$  mm diam.)..... 13
- 12b. Pileus smaller ( $\leq 60$  mm) ..... 14
- 13a. Latex changing to sulphur yellow on exposure, occurrence under *Picea* ..... *L. scrobiculatus*
- 13b. Latex unchanging on exposure, occurrence under *Quercus* .... ***L. indozonarius***
- 14a. Latex unchanging on exposure, stipe scrobiculate, occurrence under *Castanopsis*..... *L. byssaceus*
- 14b. Latex changing to lemon yellow on exposure, stipe non scrobiculate, occurrence under *Rhododendron* ..... *L. mayawatianus*
- 15a. Pileus dark olivaceous to vinaceous ..... 16
- 15b. Pileus light orange to pale yellow or pinkish ..... 20
- 16a. Latex turning greenish on exposure..... 17
- 16b. Latex unchanging on isolation..... 19
- 17a. Context turning lilac with KOH, basidiospores with broken to partial reticulum, occurrence under *Abies* ..... *L. olivaceo-umbrinus*
- 17b. Context not turning lilac with KOH, basidiospores with zebroid ornamentation ..... 18
- 18a. Occurrence in subtropical to temperate forest under *Quercus*, pleuromacrocystidia with needle-like contents..... *L. maitlyensis*
- 18b. Occurrence in subalpine forest under *Betula*, pleuromacrocystidia without needle-like contents..... *L. yumthangensis*
- 19a. Latex staining lamellae and context lilac, under *Abies* and *Tsuga* ..... *L. pyriodorus*
- 19b. Latex not staining lamellae and context, under *Quercus*..... *L. proximellus*
- 20a. Pileipellis a trichoderm, occurrence under *Rhododendron* ..... *L. dafricanus*
- 20b. Pileipellis an ixocutis ..... 21
- 21a. Stipe scrobiculate, latex changing to bright grey to pale brownish grey on lamellae edges, occurrence under *Quercus*..... *L. zonarius*
- 21b. Stipe not scrobiculate, latex unchanging..... 22
- 22a. Lamellae white to yellowish white, latex scarce, basidiospores  $6-7 \times 5-6 \mu\text{m}$ , occurrence under *Quercus*..... ***L. thindii***
- 22b. Lamellae pale vinaceous-cinnamon, latex copious, basidiospores  $7-9 \times 6-7.5 \mu\text{m}$ , occurrence under *Quercus* ..... *L. yazooensis*
- 23a. Pileus hairy or fibrillose..... 24
- 23b. Pileus smooth ..... 26
- 24a. Latex immediately changing to yellow on exposure, occurrence under *Quercus*..... *L. abbotanus*
- 24b. Latex unchanging on exposure ..... 25

- 25a. Pileus surface viscid to glutinous, stipe scrobiculate, larger basidiospores (7.5-10 × 6-7.5 µm), occurrence under *Abies* ..... *L. alnicola*
- 25b. Pileus surface dry, stipe non scrobiculate, smaller basidiospores (6.6-7.4 × 4.9-5.2 µm), occurrence under *Betula* ..... *L. pubescens*
- 26a. Lamellae subdistant to distant (2-5/cm) ..... 27
- 26b. Lamellae crowded (17-21/cm) ..... 28
- 27a. Lamellae orange yellow, pileus margin regular, basidiospore ornamentations >0.5 µm high, occurrence under *Rhododendron* and *Quercus* ..... *L. dhakurianus*
- 27b. Lamellae yellowish white, pileus margin crenate, basidiospore ornamentations 2-2.4 µm high, occurrence under *Quercus* ..... *L. kumaonensis*
- 28a. Pileus surface viscid, pileipellis an ixocutis ..... 29
- 28b. Pileus surface very glutinous to slimy, pileipellis an ixotrichoderm ..... 30
- 29a. Lamellae white to yellowish buff, latex copious, changing to greyish, staining lamellae violaceous to brownish lilac ..... *L. pseudoaspideus*
- 29b. Lamellae pinkish, latex scarce, unchanging and not staining the lamellae, occurrence under *Salix* and *Populus* ..... *L. controversus*
- 30a. Latex unchanging, smaller basidiospores (7-8.7 × 6-7.1 µm), occurrence under *Tsuga* ..... *L. elaiovascidus*
- 30b. Latex changing to pinkish on cut lamellae, larger basidiospores (8-10.1 × 6.7-7.8 µm), occurrence under *Abies* ..... *L. olivaceoglutinus*

**Acknowledgements.** The authors are thankful to the Head of the Department of Botany & Microbiology, HNBGU, Srinagar Garhwal for making available the facilities for the study. KD is thankful to the Director of the Botanical Survey of India, Kolkata for providing facilities. PU is thankful to UGC for providing a fellowship. Help rendered from Mr. Aniket Ghosh, Mr. Tahir Mehmood and Mr. Upendra Singh during the field surveys is duly acknowledged.

## REFERENCES

- ALTSCHUL S.F., MADDEN T.L., SCHÄFFER A.A., ZHANG J., ZHANG Z., MILLER W. & LIPMAN D.J., 1997 — Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acid Research* 25 (17): 3389-3402.
- ATRI N.S., SAINI M.K. & SAINI S.S., 1994 — Indian Russulaceae Roze-A checklist. In: Current Researches in Plant Sciences (Eds.: Sarma, T.A., Saini, S.S., Trivedi, M. L. and Sharma, M.). Bishen Singh Mahendra Pal Singh, Dehradun. pp 81-93.
- BASSO M.T., 1999 — *Lactarius* Pers., *Fungi Europaei* 7. Mykoflora. 845 pp.
- CLARK K., KARSCH-MIZRACHI I., LIPMAN D.J., OSTELL J. & SAYERS E.W., 2016 — GenBank. *Nucleic Acid Research* 44: D67-D72.
- DAS K., 2013 — *Lactarius fennoscandicus* (Russulaceae) A new record for India. *Nehumbo* 55: 214-218.
- DAS K. & SHARMA J.R., 2005 — *Russulaceae of Kumaon Himalaya*. Botanical Survey of India, 255 pp.
- DAS K. & VERBEKEN A., 2011 — Three new species of *Lactarius* (Russulaceae) from Sikkim, India. *Cryptogamie Mycologie* 32 (4): 365-381.
- DAS K., VERBEKEN A. & NUYTINCK J., 2015 — Morphological description and phylogenetic placement of four new species of *Lactarius* from India. *Mycotaxon*. 130(1): 105-130.
- HALL T.A., 1999 — BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acid Symposium Series* 41: 95-98.
- HEILMANN-CLAUSEN J., VERBEKEN A. & VESTERHOLT J., 1998 — *The genus Lactarius*. The Danish Mycological Society, Odense. 287 pp.
- JOSHI S., BHATT R.P. & STEPHENSON S.L., 2012 — The current status of the family Russulaceae in the Uttarakhand Himalaya, India. *Mycosphere* 3(4): 486-501.

- KATOH K. & STANDLEY D.M., 2013 — MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Molecular Biology and Evolution* 30: 772-780.
- KÖLJALG U., NILSSON R.H., ABARENKOV K., TEDERSON L., TAYLOR A.F.S., BAHRAM M., BATES S.T., BRUNS T.D., BENGTSSON-PALME J., CALLAGHAN T.M., DOUGLAS B., DRENKHAN T., EBERHARDT U., DUEÑAS M., GREBENC T., GRIFFITH G.W., HARTMANN M., KIRK P.M., KOHOUT P., LARSSON E., LINDAHL B.D., LÜCKING R., MARTÍN M.P., MATHENY P.B., NGUYEN N.H., NISKANEN T., OJA J., PEAY K.G., PEITNER U., PETERSON M., PÖLDMAA K., SAAG L., SAAR I., SCÜBLER A., SCOTT J.A., SENÉS C., SMITH M.E., SUIJA A., TAYLOR D.A., TELLERIA M.T., WEISS M., LARSSON K.-H., 2013 — Towards a unified paradigm for sequence-based identification of fungi. *Molecular Ecology* 22: 5271-5277.
- KORNERUP A. & WANSCHER J.H., 1978 — Methuen handbook of color, 3<sup>rd</sup> Ed., Eyre Methuen Ltd., London, UK.
- LE H.T., NUYTINCK J., VERBEKEN A., SAISAMORN L. & DESJARDIN D.E., 2007 — *Lactarius* in Northern Thailand: 1. *Lactarius* subgenus *Piperites*. *Fungal Diversity* 24: 173-224.
- LEONARDI M., COMANDINI O. & RINALDI A.C., 2016 — Peering into the Mediterranean black box: *Lactifluus rugatus* ectomycorrhizas on *Cistus*. *IMA Fungus* 7(2): 275-284.
- MILLER M.A., PFEIFFER W. & SCHWARTZ T., 2010 — Creating the CIPRES science gateway for inference of large phylogenetic trees. *Proceedings of the Gateway Computing Environments Workshop (GCE)*: 1-8.
- NUYTINCK J., VERBEKEN A. & MILLER S.L., 2007 — Worldwide phylogeny of *Lactarius* section *Deliciosi* inferred from ITS and glyceraldehyde-3-phosphate dehydrogenase gene sequences. *Mycologia* 99(6): 820-832.
- SHI S.-F., WANG X.-H. & BAU T., 2018 — Three new species of *Lactarius* (Russulaceae, Russulales) from Northeast China. *Mycoscience* 59: 206-217.
- STAMATAKIS A., 2014 — RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* 30: 1312-1313.
- THIERS B., (continuously updated) Index Herbariorum: A global directory of public herbaria and associated staff. New York Botanical Garden's Virtual Herbarium. <http://sweetgum.nybg.org/science/ih/>.
- UNIYAL P., DAS K., BHATT R.P., SINGH U. & MEHMOOD T., 2017 — *Lactarius kumaonensis* sp. nov. (Russulaceae) from Uttarakhand, India. *Nordic Journal of Botany* 35: 724-729.
- VERBEKEN A. & HORAK E., 2000 — *Lactarius* (Basidiomycota) in Papua New Guinea. 2. Species of tropical montane rainforests. *Australian Systematic Botany* 13: 649-707.
- VERBEKEN A. & NUYTINCK J., 2013 — Not every milkcap is a *Lactarius*. *Scripta Botanica Belgica* 51: 162-168.
- WANG X.-H., 2017 — Seven new species of *Lactarius* subg. *Lactarius* (Russulaceae) from southwestern China. *Mycosystema* 36: 1463-1482.
- WANG X.-H., HALLING R.E., HOFSTETTER V., LEBEL T. & BUYCK B., 2018 — Phylogeny, biogeography and taxonomic re-assessment of *Multifurca* (Russulaceae, Russulales) using three-locus data. *Plos One* 13(11): e0205840.
- WHITE T.J., BRUNS T., LEE S. & TAYLOR J., 1990 — Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis, M.A., Gelfand D.H., Sninsky J.J. and White T.J. (Eds.) *PCR Protocols: a guide to method and applications*. Academic Press, San Diego, pp. 315-322.